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Genome sequence of *Mesorhizobium mediterraneum* strain R31, a nitrogen-fixing rhizobium used as an inoculant for chickpea in Argentina

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ABSTRACT Here, we report the complete genome sequence of *Mesorhizobium mediterraneum* R31, a rhizobial strain recommended and used as a commercial inoculant for chickpea in Argentina. The genome consists of 7.25 Mb, distributed into four circular replicons: a chromosome of 6.72 Mbp and three plasmids of 0.29, 0.17, and 0.07 Mbp.

KEYWORDS *Mesorhizobium mediterraneum*

Inoculating legumes with rhizobia is a sustainable agricultural strategy. Rhizobia fertilize the crop biologically through nitrogen-fixing symbiosis and thus prevent nitrogen deficiency (1, 2). The Instituto Nacional de Tecnología Agropecuaria (INTA) in Argentina provides two strains of different species to inoculate chickpea, *Mesorhizobium ciceri* R30 and *Mesorhizobium mediterraneum* R31 (3). We recently reported the complete annotated genome of the former, *M. ciceri* strain R30 (4); here, we announce the complete annotated genome of the latter, *M. mediterraneum* R31. To date, only two incomplete genomes for *M. mediterraneum* strains (CCBAU 01399 and USDA 3392) are available in NCBI.

A pure culture of *M. mediterraneum* R31, grown in yeast extract-mannitol medium at 30°C with 150 rpm rotation for four days (optical density at 600 nm [OD₆₀₀], 1.0) (5), was the source for the total DNA. It was obtained with a DNeasy Blood & Tissue Kit (Qiagen) for Illumina sequencing and a Promega Wizard HMW DNA Extraction Kit (Promega) for Oxford Nanopore Technologies sequencing. The genome was assembled through a hybrid approach, including short Illumina reads and long Oxford Nanopore reads. Illumina's library was prepared with a Nextera XT DNA Library Preparation Kit and sequenced on Illumina NextSeq 500 with a paired-end 150-bp read configuration. Nanopore's library was prepared with an Oxford Nanopore Technologies Rapid Barcoding sequencing kit (SQK-RBK004) and sequenced in two Flongle flowcells. Data were base-called on Guppy v4.2.2, using the high-accuracy model and the --trim_barcodes option. We obtained 6,441,624 Illumina PE reads predicting a 132-fold coverage, and 119,523 Nanopore long reads which averaged 5,027 bp and predicted an 83-fold coverage. The raw reads were quality controlled by manually inspecting the reports obtained on FastQC (6) (Illumina) and PycoQC (7) (Nanopore). Hybrid genome assembly was performed with the raw reads in the nf-core/bacass pipeline (commit ceebac0) set at default parameters (8). This resulted in four contigs that were closed by manually analyzing the overlapped ends on Geneious software version 2019 2.1 (9). DNA content analysis revealed a genome made up of one large chromosome (6,720,795 bp, G+C content 62.2%) and three minor replicons from the repABC plasmid family (10)

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Emiliano Foresto and Santiago Revale contributed equally to this article. Author order was determined by seniority in working with chickpea nodulating strains.

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(293,882 bp, G+C content 60.1%; 175,478 bp, G+C content 60.2%; and 66,723 bp, G+C 58.6%).

The complete genome, which was annotated in the NCBI Prokaryotic Genomes Annotation Pipeline (PGAP) (11–13), consists of 6,695 protein-coding sequences, two complete ribosomal operons, and 53 tRNAs. The genes for nodulation (*nod*) and nitrogen fixation (*nif* and *fix*) appear to be located on a chromosomal 480 kb symbiosis island (1,850,730 to 2,330,771 bp), flanked by direct repeat sequences (22 nt) identical to those in the ICE region in other mesorhizobia, and adjacent to one of four serine tRNA genes (4, 14). This region also harbors mobile genes (transposases, integrases, and recombinases) probably associated with the island's excision and transfer (15, 16).

This complete genome sequence of an *M. mediterraneum* strain could be crucial for more in-depth research into its symbiotic performance and other biological features of this species.

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DATA AVAILABILITY

The complete genome sequence of *Mesorhizobium mediterraneum* R31 is available at NCBI GenBank under accession numbers [CP088151](#) for the chromosome, [CP088152](#), [CP088153](#), and [CP088154](#) for the plasmids with BioProject ID [PRJNA782313](#). and BioSample accession [SAMN23372083](#). Raw data reads are available at NCBI's Sequence Read Archive under accession numbers [SRR16993324](#) to [SRR16993326](#).

REFERENCES

1. Foresto E, Nieves F, Revale S, Giordano W, Bogino P. 2021. Deciphering the phylogenetic affiliation of rhizobial strains recommended as chickpea inoculants in Argentina. *App Soil Ecol* 166:104069. <https://doi.org/10.1016/j.apsoil.2021.104069>
2. Yang J, Lan L, Jin Y, Yu N, Wang D, Wang E. 2022. Mechanisms underlying legume-rhizobium symbioses. *J Integr Plant Biol* 64:244–267. <https://doi.org/10.1111/jipb.13207>
3. Santos MS, Nogueira MA, Hungria M. 2019. Microbial inoculants: reviewing the past, discussing the present and previewing an outstanding future for the use of beneficial bacteria in agriculture. *AMB Express* 9:205. <https://doi.org/10.1186/s13568-019-0932-0>
4. Foresto E, Revale S, Primo E, Nieves F, Carezzano E, Puente M, Alzari P, Martínez M, Ben-Assaya M, Mornico D, Santoro M, Martínez-Abarca F, Giordano W, Bogino P. 2022. Complete genome sequence of *Mesorhizobium ciceri* strain R30, a rhizobium used as a commercial Inoculant for chickpea in Argentina. *Microbiol Resour Announc* 11:e0077922. <https://doi.org/10.1128/mra.00779-22>
5. Somasegaran PH, Hoben H. 1994. Appendix 3: Media and staining solutions, p 383–384. In *Handbook for rhizobia: methods in legume-rhizobium technology*. xvi. Springer, New York, NY. <https://doi.org/10.1007/978-1-4613-8375-8>
6. Andrews S. 2010. Fastqc: a quality control tool for high throughput sequence data. Available online at. Available from: <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>
7. Leger A, Leonardi T. 2019. pycoQC, interactive quality control for oxford nanopore sequencing. *JOSS* 4:1236. <https://doi.org/10.21105/joss.01236>
8. Ewels PA, Peltzer A, Fillinger S, Patel H, Alneberg J, Wilm A, Garcia MU, Di Tommaso P, Nahnsen S. 2020. The nf-core framework for community-curated bioinformatics pipelines. *Nat Biotechnol* 38:276–278. <https://doi.org/10.1038/s41587-020-0439-x>
9. Kearsse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28:1647–1649. <https://doi.org/10.1093/bioinformatics/bts199>
10. MacLellan SR, Zaheer R, Sartor AL, MacLean AM, Finan TM. 2006. Identification of a megaplasmid centromere reveals genetic structural diversity within the repABC family of basic replicons. *Mol Microbiol* 59:1559–1575. <https://doi.org/10.1111/j.1365-2958.2006.05040.x>
11. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
12. Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. Refseq: an update on prokaryotic genome annotation and curation. *Nucleic Acids Res* 46:851–860. <https://doi.org/10.1093/nar/gkx1068>
13. Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. Refseq: expanding the prokaryotic genome annotation pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>
14. Hill Y, Colombi E, Bonello E, Haskett T, Ramsay J, O'Hara G, Terpolilli J. 2021. Evolution of diverse effective N₂-Fixing Microsymbionts of *Cicer arietinum* following horizontal transfer of the *Mesorhizobium ciceri* CC1192 Symbiosis integrative and conjugative element. *Appl Environ Microbiol* 87:e02558-20. <https://doi.org/10.1128/AEM.02558-20>
15. Ramsay JP, Sullivan JT, Stuart GS, Lamont IL, Ronson CW. 2006. Excision and transfer of the *Mesorhizobium loti* R7A Symbiosis Island requires an Integrase IntS, a novel recombination directionality factor RdfS, and a putative relaxase RlxS. *Mol Microbiol* 62:723–734. <https://doi.org/10.1111/j.1365-2958.2006.05396.x>
16. Haskett TL, Terpolilli JJ, Bekuma A, O'Hara GW, Sullivan JT, Wang P, Ronson CW, Ramsay JP. 2016. Assembly and transfer of tripartite integrative and conjugative genetic elements. *Proc Natl Acad Sci U S A* 113:12268–12273. <https://doi.org/10.1073/pnas.1613358113>