

# Adenine, a Key Player in Biology and Medicinal Chemistry

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| 2  | Adenine, a Key Player in Biology and Medicinal Chemistry   |
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| 3  | L'adénine, un acteur majeur en biologie et en chimie médicinale  |
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#### 25 Abstract:

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29 Adenine is one of the most ubiquitous heterocycles in life. In addition of being one of the four 30 nucleobases constituting DNA and RNA, adenine is also contained in many biological 31 molecules (ATP, SAM, NAD, cAMP, coA...) that have fundamental roles in the functioning of living systems, e.g. energy source, cofactors of enzymes and proteins. As such, the adenine 32 33 has naturally become a privileged scaffold explored in medicinal chemistry for biomedical 34 applications. Many chemical modifications and Structure-Activity Relationships studies have 35 been carried out on the adenine scaffold to result in potent analogues with various biological 36 activities. Today, numerous adenine-based inhibitors are used to treat a wide range of diseases 37 including cancer, viral and bacterial diseases. This review aims to introduce the adenine and 38 discuss adenine-based inhibitors, their design and use for different therapeutic targets through 39 examples of drugs and compounds that reached clinical and preclinical trials.

40

41 <u>Key words:</u> Adenine; medicinal chemistry; inhibitors; anticancer; antiviral and antibacterial
42 agents.

#### 44 <u>Résumé :</u>

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46 L'adénine est l'un des hétérocycles les plus représentés dans la nature. En plus d'être l'une des 47 quatre bases constituant l'ADN et l'ARN, elle est également présente dans de nombreuses 48 molécules biologiques (ATP, SAM, NAD, cAMP, coA...) qui jouent un rôle fondamental dans 49 le fonctionnement des systèmes vivants, comme source d'énergie, cofacteurs d'enzymes ou de 50 protéines. L'adénine est ainsi devenue un hétérocycle largement exploré en chimie médicinale 51 pour de nombreuses applications biomédicales. De multiples modifications chimiques 52 associées à l'étude de Relations Structure-Activité ont été réalisées sur le squelette de l'adénine 53 pour aboutir à des analogues puissants aux activités biologiques variées. Aujourd'hui, de 54 nombreux inhibiteurs analogues d'adénine sont utilisés pour traiter un large éventail de 55 maladies, comme le cancer et les maladies infectieuses. Cette revue a pour but de présenter 56 l'adénine dans son contexte biologique et de discuter d'exemples d'inhibiteurs constitués d'une 57 adénine (ou de l'un de ses analogues) en décrivant leur conception et utilisation pour différentes 58 cibles thérapeutiques.

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Mots clés : adénine ; chimie médicinale ; inhibiteurs ; agents anticancéreux ; antiviraux et
 antibiotiques.

#### 63 Introduction

64

65 The purine ring system, and especially *adenine*, is one of the most ubiquitous natural 66 heterocyclic compounds and plays major roles in various biological processes. In addition of 67 being a nucleobase and building block of the genetic material and its transcription and 68 translation in proteins, adenine stores energy in the form of Adenosine TriPhosphate (ATP) 69 and is found in the structure of cofactors of many enzymes such as ATP, S-Adenosyl-L-70 Methionine (SAM or AdoMet), Nicotinamide Adenine Dinucleotide (NAD), Flavin Adenine 71 Dinucleotide (FAD) and in substrates of metabolic pathways, as acetyl-Coenzyme A (acetyl-72 CoA) (Fig. 1). ATP is both the source of energy at the cellular level, a precursor to DNA and 73 RNA synthesis and a cofactor of protein kinases to phosphorylate their substrates.[1] SAM is 74 used as methyl donor by MethylTransferases (MTases) to methylate histones, DNA, lipids and 75 small biological molecules.[2] NAD+ is a cofactor for redox reactions and has a central role in 76 energy metabolism. It is also a coenzyme for non-redox NAD+-dependent enzymes [CD38, 77 sirtuins and poly(ADP-ribose)] and is a substrate for Nicotinamide Adenine Dinucleotide 78 Kinases (NADK).[3] Some signaling molecules such as cyclic Adenosine MonoPhosphate 79 (cAMP) contain also an adenine scaffold. cAMP is an ubiquitous second messenger that 80 regulates various biological processes, interacting with protein effectors such as Protein Kinase 81 A (PKA), Exchange Proteins Activated by cAMP (EPACs) and Cyclic Nucleotide-Gated 82 (CNG) channels.[4]

Importantly, these biological pathways are deregulated in diseases and the aforementioned protein families represent major therapeutic targets. Thus, the adenine scaffold has been exploited to design and develop therapeutic agents.[5–7] Chemical modifications of the adenine scaffold - giving *adenine analogues* – have been carried out to identify inhibitors of these pathways deregulated in diseases. Kinases and MTases inhibitors, based on the adenine 88 scaffold are used for cancer treatment.[8,9] Adenosine analogues inhibiting DNA/RNA 89 synthesis are widely utilized as anticancer and antiviral agents.[10,11] NAD Analogues are 90 investigated to target NADK in the context of bacterial infection.[12] Finally, cAMP analogues 91 are emerging as modulators of EPACs, playing a role in inflammation, diabetes and 92 cardiovascular diseases.[13] Another example is the use of adenosine receptor agonists as 93 anticancer agents.[14]

94

Herein, we review the highlights of the use of the adenine scaffold for the design of versatile therapeutic agents and its role in biological processes. This review is not meant to be exhaustive and presents its chemical modulation in Medicinal Chemistry through chosen examples of approved drugs and inhibitors in clinical trials and preclinical studies for each selected therapeutic target.



101 **Figure 1.** The adenine scaffold (in blue) in biological molecules and therapeutic agents.

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# 1041. <u>S-Adenosyl-L-Methionine/S-Adenosyl-L-Homocystein</u> (SAM/SAH)

# 105

#### analogues to target SAM-dependent methyltransferases

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S-Adenosyl-*L*-Methionine is one of the most ubiquitous cofactors. It is mainly used by
the SAM-dependent MethylTransferases (MTasaes) as methyl donor to methylate their
substrates (DNA, RNA, proteins, carbohydrates and metabolites). These methylated substrates
contribute for example to the epigenetic regulation, protein function modulation, metabolites
synthesis and degradation etc...[15–19]

112 SAM is a hybrid compound of adenosine bonded to the sulfur atom of methionine. The 113 sulfonium group that bears the methyl group confers the SAM's reactivity. Under the catalytic 114 reaction, the nucleophilic substrate (N-, C-, O- or S-nucleophiles) attacks the electrophilic 115 methyl group through an S<sub>N</sub>2-type mechanism generating S-Adenosyl-L-Homocystein (SAH) 116 as by product (Fig. 2A).[20,21] The methyl group transfer happens in the catalytic site of 117 MTases that is composed of two pockets: a SAM binding site and a substrate binding site.[15] 118 SAM-Dependent MTases are a wide and diversified class of enzymes. They can be 119 classified according to their type of substrate. First, Histone MethylTransferases (HMTs) catalyze the transfer of methyl group on lysine and arginine residues of histones and are 120 121 partitioned into Protein Arginine MethylTransferases (PRMTs) and Lysine 122 MethylTransferases (KMTs) (Fig. 2B).[22,23] Other protein methyltransferases methylate side 123 chains of non-histone proteins, [24] especially N-Terminal MethylTransferases (NTMTs) have 124 raised interest recently.[25,26] Among the DNA MTases, C5 DNA MethylTransferases 125 (DNMTs) catalyze the methylation of C5-position of cytosines (5mC) in DNA (Fig. 2B), which 126 plays a major role in human diseases, and for which we and other have developed several 127 inhibitors.[27] Additional MTases are responsible for the methylation of small molecules such

128 as Catechol *O*-MethylTransferases (COMT) that methylate catecholamines for
129 neurotransmitter degradation.[28]

Due to the plethora of biological functions of MTases but also to their involvement in various diseases, SAM and SAH analogues have been developed and exploited either as MTases inhibitors or as tools to study the functions of MTases.[29]



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Figure 2. A) SAM-dependent MTase-catalyzed methylation reaction; B) Examples of methylation products:
mammalian histone and DNA methylation by HMTs and DNMTs; C) Examples of adenine methylation on RNA;
D) COMT-catalyzed methylation of Levodopa; E) NNMT-catalyzed methylation of nicotinamide.
(Biorender.com was used to prepare part of the Figure.)

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#### 140 **1.1 Histone MethylTransferases (HMT) inhibitors**

HMTs are a broad class of enzymes with more than 50 members of KMTs and 9 PRMTs.[30] KMTs catalyze mono- (Kme1), di- (Kme2) or tri- (Kme3) methylation of lysine residues, whereas PRMTs catalyze monomethylation of arginines (Rme1) or dimethylation of arginines (in a symmetrical (Rme2s) or asymmetrical (Rme2a) manner) (Fig. 2B). HMTs are

involved in various diseases, in particular cancers and infectious diseases, and are thus very
interesting therapeutic targets.[31–35] SAM and SAH scaffolds were exploited to develop
numerous HMT inhibitors.[36] We chose few examples among the large literature to show the
design and application of adenine-based inhibitors (Fig. 3).





151 Figure 3. Examples of some SAM/SAH inhibitors. A) HMT inhibitors; B) DNMT inhibitors; C) NTMT
152 inhibitors; D) NNMT inhibitors; E) COMT inhibitors.

153

SAH is an endogenous inhibitor of many MTases and shows, for example, a similar inhibitory potency for PRMT4, G9a and DOT1L (IC<sub>50</sub>  $\approx$  1.5  $\mu$ M). Hence, co-crystal structures of SAH complexed with different HMTs were exploited to develop HMT inhibitors. DOT1L, the only HMT that catalyzes H3K79 methylation, attracted the interest notably because it is a 158 major target for acute leukemia with Mix Lineage Leukemia (MLL) translocations. 159 Interestingly, DOT1L is a KMT that does not bear the catalytic SET domain but rather shows 160 a similarity in the 3D structure of the catalytic pocket with the PRMT's one, and exhibits a 161 unique hydrophobic subpocket in the adenosine binding site.[37] Following this rational, Yao et al. designed a potent and selective compound, 1 (Fig. 3A), with a methyl substitution on the 162 163 N6 position of SAH (to protrude interactions in the hydrophobic pocket specific of DOT1L) 164 and the C-S bond of SAH was replaced by a C-N bond (DOT1L, IC<sub>50</sub> = 120 nM) (Fig. 3A).[38] 165 In 2011, Epizyme Inc. developed a new potent DOT1L inhibitor EPZ-4777, 2, where the 7-166 deaza SAM mimics is linked to a bulky 4-tert-butylphenyl group to occupy also the substrate 167 pocket (DOT1L, IC<sub>50</sub> =  $0.4 \text{ nM} \pm 0.1$ ).[39,40] This compound is a SAM competitive inhibitor 168 and the 4-tert-butylphenyl group occupies a newly opened hydrophobic subpocket. EPZ-4777 169 efficiently inhibits the proliferation of MLL-rearranged leukemia cell lines but the poor pharmacokinetic properties of the compound blocked its progress. To improve the 170 171 pharmacokinetic parameters, a new compound EPZ-5676, 3 (Pinometostat), was developed. In 172 EPZ-5676 the propyl chain of EPZ-4777 is replaced by a *cis*-cyclobutyl ring and the 173 benzimidazole moiety replaces the aryl-substituted urea group. Moreover, the adenosine part 174 of SAM is conserved in EPZ-5676.[41] The compound is very potent (DOT1L,  $K_i = 0.08 \text{ nM}$ 175  $\pm$  0.03) and is selective towards DOT1L. In addition, EPZ-5676 shows better residence time 176 (>24 h vs 1 h) and entered clinical trials (NCT03701295). Interestingly, since EPZ-4777 weakly 177 inhibits PRMT5 (at 30 µM), it served at starting point for the development of new PRMT5 178 SAM-competitive inhibitors. After extensive SAR exploration, compound JNJ-64619178, 4 179 (by Johnson and Johnson), was developed (Fig. 3A). It is a bisubstrate inhibitor composed of 180 a 7-deaza SAM mimics and a substituted quinoline moiety, occupying the second substrate 181 binding pocket. JNJ-64619178 is potent (PRMT5,  $IC_{50} = 0.13$  nM) and selective towards

182 PRMT5; it displays favorable pharmacokinetic properties[42] and entered clinical trial notably

183 for the treatment of advanced solid tumors (NCT03573310).[9]

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#### 185 **1.2.** DNA MethylTransferases (DNMT) inhibitors

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187 DNA methylation, mediated by DNMTs, contributes to maintain normal cell function and 188 to embryonic development. DNMT1 catalyzes methylation of newly synthesized DNA strands, 189 while DNMT3A/B are responsible for de novo DNA methylation.[43] In humans, DNA 190 methylation mainly takes place on position 5 of cytosine in a CpG dinucleotide context and is 191 generally associated with gene repression when in promoters. Abnormal DNA methylation 192 patterns and alterations in DNMTs activity can induce tumor development and maintenance. DNMTs are thus interesting anticancer targets.[27] Azacitidine (Vidaza<sup>TM</sup>) and Decitabine 193 194 (Dacogen®), are two DNMT inhibitors approved by the FDA (US Food and Drug 195 Administration).[44,45] These drugs are cytosine analogues that replace cytosine in DNA, and 196 in RNA for the former. However, they lack selectivity towards CpGs and show poor stability. 197 More recently, a new class of reversible DNMT1-selective inhibitors has been developed by 198 GlaxoSmithKline (GSK), targeting allosteric sites. The compound GSK-3484862 that contains 199 a dicyanopyridine moiety is promising with its low cytotoxicity and great specificity. [46,47] 200 In addition, adenine analogues have been investigated to mimic SAH. MethylGene Inc. 201 performed structural modifications of SAH and showed that substituting the homocystein with 202 a proline mimics confers potent activity (compound 5 in Fig. 3B), [48] while the 7-deaza SAH, 203 6, showed similar activity as SAH.[49] Finally, DNMT bisubstrate inhibitors with a 204 quinazoline moiety as adenine mimic were developed by our laboratory and others.[50–53] Compound 7 inhibited DNMT3A and DNMT1 at 4 µM and 46 µM respectively (Fig. 3B).[54] 205 206

#### 1.3. N-Terminal MethylTransferase (NTMT) inhibitors

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209 Protein N-Terminal MethylTransferase 1 (NTMT1) methylates the N-terminal amine of 210 proteins and recognizes the canonical motif X-P-K (with X= A, P or S).[55] Dysregulation of 211 NTMT1 is associated with several cancers and aging processes.[56] NTMT inhibitors have 212 been developed notably to study the biological functions of N $\alpha$  methylation. A first NMT1 213 bisubstrate inhibitor NAM-TZ-SPKRIA, 8, has been developed by Zhang *et al.* (Fig. 3C).[57] 214 It is composed of 5'-N SAM analogue coupled to an hexapeptide (SPKRIA) via a triazole 215 linker. The compound is very potent (IC<sub>50</sub>=  $0.81 \pm 0.13 \mu$ M) and shows a 60-fold selectivity 216 over other representative MTases. Later, Chen et al., developed a new series of NTMT1 217 bisubstrate inhibitors.[58] With a new synthetic route, they replaced the first serine residue of 218 the SPKRIA peptide by a proline residue giving the NAM-C3-PPKRIA, 9 (Fig. 3C), exhibiting 219 an improved potency (IC<sub>50</sub> =  $35 \pm 2$  nM) and more than 100-fold selectivity over NTMT1 220 against a panel of MTases. However, the compound showed a poor cell permeability preventing 221 cell-based studies. Nonetheless, this work enabled to obtain the first co-crystal structure of 222 inhibitors with NTMT1 that would help for the development of novel NTMT1 inhibitors.

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#### 1.4. Nicotinamide N-MethylTransferase (NNMT) inhibitors

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NNMT catalyzes methylation of nicotinamide using the SAM cofactor (Fig. 2E). NNMT has a major role in cell detoxification and metabolism. Indeed, NNMT can recognize a broad range of substrates (pyridines, quinolines, diverse heterocyclic metabolites) and catalyzes their methylation, which is followed by their excretion.[59] Nicotinamide is also the precursor of NAD<sup>+</sup>. Moreover, NNMT balances SAM/SAH ratio that impacts indirectly gene expression.[60] Indeed, it has been reported that NNMT is also involved in the development 232 and progression of cancers, neurodegenerative diseases, diabetes and obesity. [61–63] To better 233 understand the role of NNMT in these diseases, several NNMT inhibitors have been developed, 234 as potent bisubstrate compounds composed of SAM and nicotinamide mimics covalently 235 bound through a linker. In 2017, Van Haren et al. developed the first bisubstrate compound 10 236 targeting NNMT (IC<sub>50</sub> = 29.2  $\pm$  4.0  $\mu$ M),[64] followed by other derivatives.[65,66] In 237 particular, Policarpo et al. synthesized a new potent bisubstrate compound 11 with an increased 238 potency in the namomolar range ( $K_i = 0.51 \pm 0.08 \text{ nM}$ ).[67] They figured out that an alkyne 239 linker between the NAM-like and SAM-like fragments was optimal to mimic the transition 240 state geometry. Recently, Gao et al. developed a novel potent bisubstrate compound 12 (IC50 241 =  $3.7 \pm 0.2$  nM) with a *para*-cyano substituted styrene scaffold in the nicotinamide mimic that 242 replaces the common benzamide moiety.[68] However, the compound was not potent in cell-243 based assays probably due to a poor cell permeability. All these findings help the further design 244 and development of new NNMT inhibitors.

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#### 246 1.5. Catechol O-Methyltransferases (COMT) inhibitors

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248 COMT catalyzes methylation of catecholamines (e.g. dopamine and epinephrine) but also 249 of Levodopa (L-dopa) producing 3-O-Methyldopa (Fig. 2D). L-Dopa is the precursor of 250 dopamine and is used to treat the symptoms of Parkinson Disease (PD).[69] Administrating 251 COMT inhibitors avoids L-dopa deactivation and prolongs the effects of PD treatment.[70] 252 COMT bisubstrate inhibitors were developed. Based on the crystal structure of COMT with 253  $Mg^{2+}$ , SAM and 3,5-dinitrocatechol, the bisubstrate compound 13 was designed.[71] 254 Compound 13 is a competitive inhibitor of SAM and a noncompetitive inhibitor of catechol. 255 The potency was increased by rigidifying the linker with a double bond giving compound 14 256  $(IC_{50} = 9 \text{ nM})$ .[72] To avoid hepatotoxicity, the nitro group was replaced by fluoro-substituted 257 phenyl moiety giving the potent compound **15** (IC<sub>50</sub> = 21 nM).[73] Substitution of the OH at 258 position 3 of ribose gave compound **16** (IC<sub>50</sub> = 9 nM).[74]

259

#### 260 **2.** Adenine scaffold in kinase inhibitors

261

262 Protein kinases catalyze the transfer of a  $\gamma$ -phosphate group from ATP to a substrate as for example tyrosine, serine or threonine residues of proteins (Fig. 4A). Approximately 538 263 264 kinases have been identified in the human genome so far.[75] Kinases play major roles in 265 numerous cellular processes such as cell proliferation, transcription, apoptosis and cell 266 survival. However, they are also involved in the development and progression of numerous 267 diseases, especially cancers but also inflammatory, neurological and cardiovascular 268 diseases.[1] Kinases are thus validated drug targets. Over 70 kinase inhibitors targeting more 269 than 20 different protein kinases have been approved with most of them used for cancer 270 treatment.[76] Kinase inhibitors can be classified according to their binding mode: Type I 271 *inhibitors* are ATP-competitive inhibitors and bind to the active conformation of kinases, *Type* 272 II inhibitors bind to the inactive conformation of kinases, Type III inhibitors bind to an 273 allosteric pocket adjacent to the ATP-binding pocket and Type IV inhibitors bind to an 274 allosteric pocket distant to the ATP-binding pocket.[77]



Figure 4. A) Protein phosphorylation-catalyzed reaction by protein kinases; B) Examples of kinase inhibitors
with an adenine analogue scaffold. (Biorender.com was used to prepare part of the Figure.)

279 Most of the kinase inhibitors bind to the ATP-binding site. ATP mimics bind making 280 hydrogen bonds to the kinase hinge backbone similarly to the ATP adenine ring.[77] Different 281 chemical scaffolds of adenine analogues are used (Fig. 4B). Some of them present the *purine* 282 scaffold that occupies the adenine pocket such as Idelalisib, 17, (Zydelig®, by Gilead 283 Sciences), a PhosphoInositide 3-Kinase  $\delta$  (PI3K $\delta$ ) inhibitor approved to treat relapsed Chronic 284 Lymphocytic Leukemia (CLL).[78] Idelalisib establishes a type II binding.[79] The 5fluoroquinazolinone part binds to an induced hydrophobic specificity pocket and the purine 285 286 moiety occupies the adenine pocket by forming a key hydrogen bond between the N6 position 287 of the adenine ring and Val828.[80,81] The pyrazolo aminopyrimidine scaffold is also used as 288 adenine mimic in inhibitors such as in Ibrutinib, 18, (Imbruvica®, by Pharmacyclics Inc.), a Non-Receptor protein Tyrosine Kinase (NRTK) targeting Burton's Tyrosine Kinase (BTK) 289

290 that was approved to treat mantle cell lymphoma and CLL.[82] Crystallography studies showed 291 that Ibrutinib is an irreversible BTK inhibitor, which covalently binds through Michael addition with Cys481 of BTK.[83] However, some insights on the mechanism of action remain to be 292 293 understand, as more recently, the determination of the structure of Ibrutinib with SRC (a non-294 receptor protein tyrosine kinase, PDB ID: 6L8L) showed that Ibrutinib binds via a non-covalent 295 manner to the protein.[84] Pyrrolo pyridine scaffolds are also used, as in Vemurafenib, 19, 296 (Zelboraf®, licensed to Roche), an inhibitor of the serine/threonine B-Raf enzyme (member 297 of the Raf kinase family) approved for late-stage melanoma.[85] The structure of Vemurafenib 298 complexed with V600E B-Raf, determined by X-ray crystallography (PDB ID: 30G7), showed 299 a type I inhibition mechanism. The pyrrolo pyridine core of Vemurafenib mimics the adenine 300 core of ATP by forming hydrogen bonds with hinge residue Cys532 and Gln530. Moreover, 301 the sulfonamide moiety interacts through hydrogen bonds to the DFG residues and the 4-302 chlorophenyl moiety is exposed to the solvent.[86] Since resistance has appeared with 303 mutations in the protein, new inhibitors have been designed (Belvarafenib by Hanmi 304 Pharmaceutical/Genetech and Lifirafenib by Novartis for example).[87] Kinase inhibitors with 305 a fused six-membered ring system such as a *quinazoline*, *quinoline* or *isoquinoline* moiety are 306 the most used ATP mimics (18 FDA approved inhibitors).[77] For instance, Afatinib, 20, 307 (Gilotrif®, by Boehringer Ingelheim), composed of a quinazoline moiety, is an irreversible 308 inhibitor of Human epidermal growth factor receptor 2 (Her2) and Epidermal Growth Factor 309 Receptor (EGFR) kinase. It was the first irreversible kinase inhibitor clinically approved for 310 treatment of Non-Small Cell Lung Carcinoma (NSCLC).[88] A type I binding mode was 311 shown with co-crystal structures of EGFR-Afatinib (PDB ID: 4G5J) and mutant T790M 312 EGFR-Afatinib (PDB ID: 4G5P). The quinazoline core binds with a conserved hydrogen bond 313 to the hinge residue Met793.[89]

It is worth mentioning that kinase inhibitors are also used to treat non-oncologic diseases such as inflammatory diseases, autoimmune diseases, cardiovascular diseases, diabetes and some neurodegenerative disorders.[90] Noteworthy, some promising kinase inhibitors with as adenine scaffold are currently in preclinical trials for Amyotrophic Lateral Sclerosis (ALS).[91]

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# 321 **3.** <u>Nicotinamide Adenine Dinucleotide (NAD) analogues to target</u>

## 322

### <u>Nicotinamide Adenine Dinucleotide Kinases (NADK)</u>

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324 Nicotinamide adenine dinucleotide is an essential cofactor in energy metabolism. NAD is a 325 dinucleotide containing an adenine and nicotinamide linked together through a ribose-326 phosphodiester backbone (Fig. 5A). NAD can be in two forms: in an oxidized form (NAD+) 327 or in a reduced form (NADH). NADK catalyze the phosphorylation of NAD on the 3'OH of 328 the adenosine to NADP, which is further reduced to NADP(H) (Fig. 5B-C).[92,93] NADK is 329 the unique enzyme producing NADP de novo and is essential to control the balance of NADH 330 and NADP(H) in various metabolic pathways. NADK has crucial roles in pathogenic bacteria 331 such as Mycobacterium tuberculosis, Staphylococcus aureus, Bacillus subtilis and Listeria 332 monocytogenes.[12, 94-98] Thus, NADK is a very promising therapeutic target for the 333 development of novel antibiotics.[99,100]







Figure 5. A) Reduction-oxidation reaction of NAD+/NADH; B) NAD+ phosphorylation-catalyzed reaction by
NADK; C) Reduction-oxidation reaction of NADP/NADP(H); D) Examples of NADK inhibitors. (Biorender.com
was used to prepare part of the Figure.)

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340 A series of adenosine derivatives targeting recombinant L. monocytogenes and S. aureus NADKs has been recently developed by the group of Pochet.[12,101–103] This work 341 342 led to the synthesis of the first NADK inhibitor NKI1, 21 (Fig. 5D), showing activity in mice 343 infected with S. aureus.[12] NKI1 is a dinucleoside of adenosine coupled through a propargyl 344 linker. This linker confers a restricted conformation that is essential for the binding to the NAD-345 binding site of bacterial NADK. It was suggested that even more constrained compounds would 346 have better affinity and specificity for NADKs.[104,105] Thus, macrocycle compounds were 347 investigated. The potent compound 22 was developed and the macrocyclization was performed 348 by intramolecular amide bond (Fig. 5D). Compound 22 is active at 55 nM on recombinant 349 NADK from L. monocytogenes (LmNADK1) and represents the best NADK-binder described 350 to date.[106]

352

#### 4. Adenosine analogues as adenosine receptor agonists

Adenosine is a major cell regulator that is involved in numerous physiological processes 353 354 such as the nervous system, vascular function, immune system, and platelet aggregation. More precisely, adenosine interacts with four G Protein-Coupled Receptors (GPCR) Adenosine 355 356 Receptors (ARs): A1, A2A, A2B and A3 (Fig. 6A).[107,108] ARs exhibit seven-passed 357 transmembrane  $\alpha$ -helix structures, the amino terminus being extracellular and the carboxyl terminus intracellular. Receptors A1 and A3 activate G<sub>i</sub> proteins that inhibit the activity of the 358 359 Adenylyl Cyclase (AC) and consequently reduce the intracellular level of cAMP. On the 360 contrary, receptors A2A and A2B are coupled to G<sub>s</sub> proteins that activate AC and induce an 361 increase in the cAMP level. ARs are distributed throughout the body, for example receptors 362 A1 are distributed in the brain, the spinal cord, heart muscles, kidney. Receptors A2A are mainly found in the immune system and the stratum, while receptors A2B are at high 363 364 concentration in the respiratory pathway. Receptors A3 are located into hippocampus, 365 thalamus, astrocytes, epithelial cells, inflammatory cells etc...[109]



<sup>367</sup> 

**Figure 6.** A) Schematic representation of adenosine signaling; B) Examples of adenosine receptor agonists. AC:



By their role, ARs are involved in various pathological functions in several human diseases as for example in central nervous system disorders, cardiovascular diseases, asthma, and cancer. ARs are thus interesting therapeutic targets and several drugs including agonists, antagonists and allosteric enhancers have been developed and approved. The drugs targeting adenosine signaling pathways are described in a recent review by Kutryb-Zając *et al.*[14] A major strategy for the design of agonists of ARs consists in modifying the *N*-6 position of adenine and the 5'-position of ribose.

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- 378

#### 8 4.1 Specific A2A receptor agonist

379 Regadenoson, 23 (Lexiscan<sup>®</sup>), is the only A2A receptor agonist that has been approved by 380 the FDA (Fig. 6B). Regadenoson is a pyrazole derivative of adenosine that induces coronaries 381 dilatation by binding to A2A receptors of the coronary arteries.[110,111] Regadenoson is more 382 than 100 times more potent than adenosine as an agonist of A2A receptor. It is used as a stress 383 agent in conjunction with Myocardial Potential Imaging (MPI) for diagnosis of Coronary 384 Artery Disease (CAD). Regarding its therapeutic use, Regadenoson is currently in phase I/II 385 study for the treatment of COVID-19. Indeed, by inhibiting hyperinflammation, Regadenoson 386 could reduce COVID-19 induced lung injury and ameliorate recovery of patients.[112]

387

#### 388 4.2 Specific A3 receptor agonists

The A3 receptor is a promising therapeutic target because it is highly expressed in the skin of psoriasis patients, in inflamed tissues of arthritic patients and in several solid tumor cells (breast, pancreatic, melanoma...).[113–115] Importantly, A3 agonists inhibit cytokine production and downregulate the NF- $\kappa$ B pathway in cancer and inflamed tissues. Piclidenoson, **24** (Fig. 6B), is an adenosine A3 agonist showing anti-inflammatory effects in arthritis and inflammatory bowel disease.[116,117] A phase II study showed that the drug was safe and well-tolerated for patients with rheumatoid arthritis.[118] Piclidenoson was tested in phase III
to treat patients with moderate to severe plaque psoriasis (NCT03168256).[119]

The A3 receptor is also overexpressed in HepatoCellular Carcinoma (HCC) tissues and it has been shown that the A3 agonist Namodenoson, **25** (Fig. 6B), inhibits tumor cell growth *in vitro* and *in vivo*.[120,121] Namodenoson is currently in phase III to treat advanced HCC (NCT05201404).[119]

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Targeting adenosine receptors has shown promising results, however due to the complexity of the AR signaling and the wide distribution in the body of ARs, only a small amount of adenosinergic drugs have been approved. Development of new drugs is currently explored.

405

# 406 5. <u>cyclic Adenosine MonoPhosphate (cAMP) analogues as Exchange</u> 407 <u>Proteins Activated by cAMP (EPAC) agonist modulators</u>

408

409 cAMP is a versatile chemical second messenger that mediates extracellular signals to 410 intracellular responses in most types of cells. Many biological processes are regulated by 411 cAMP such as cell activation, proliferation, migration or apoptosis regulation.[122] The cAMP 412 response is first initiated by primary signaling molecules (hormones, neurotransmitters or 413 prostaglandins) that activate mainly G-Protein-Coupled Receptors (GPCR), which in turn 414 activate adenylyl cyclases (Fig. 7A). ACs catalyze ATP cyclization to produce cAMP.[123] 415 There are several intracellular receptors for cAMP, as the Protein Kinase A (PKA) family, 416 EPACs and CNG channels.[124-126] Deregulations in local cAMP signaling has been 417 correlated to various pathophysiological disorders such as cancer, diabetes, cardiac dysfunction 418 and immunological diseases.[127–129]

419





422 Figure 7. A) Schematic representation of the intracellular cAMP signaling cascade; B) Examples of cAMP
423 analogues used to target EPACs (Exchange Proteins Activated by cAMP). AC= Adenylyl Cyclase; CNG= Cyclic
424 Nucleotide Gated; PKA= Protein Kinase A. (Biorender.com was used to prepare panel A.)

425

EPACs activate Ras-like small GTPases (Rap1 and Rap2) that are involved in cell-cell 426 427 adhesion. [122, 125, 130] EPAC1 and EPAC2 have been identified as potent targets for several 428 human diseases, as in cancer and cardiovascular diseases, thus small molecules modulating 429 EPAC have been developed. EPAC modulators can be classified into cAMP analogues and 430 non-cyclic nucleotide molecules (for a specific review see Wang et al.).[13] Here we focused 431 on the cAMP analogues (Fig. 7B). cAMP analogues display EPAC agonist activity. Bos et al. 432 first showed that 2'-OH modification increases selectivity EPAC/PKA selectivity. For 433 example, substitution of the 2'-OH by a methoxy group led to compound 26 with about 10- to 434 60- fold selectivity EPAC/PKA.[131] Then, bulky substitution on adenine at the C8-position 435 gave EPAC agonist 27 with an half-maximal EPAC1 activation at 2.2 µM and about 100-fold EPAC/PKA selectivity.[132] However, compound 27 showed poor membrane penetrating 436 437 capability. Then, a prodrug, compound 28 (Fig. 7B) was developed with esterification of the

438 4-phosphate group. Compound 28 was over 100-fold more potent than compound 27 and did
439 not affect EPAC/PKA selectivity but acted also as a PhosphoDiEsterases (PDE) inhibitor.[133]
440

441

#### 6. Adenosine analogues as antiviral and anticancer agents

442

443 Nucleoside analogues are widely used for antiviral and anticancer therapy by targeting DNA 444 or RNA replication of the virus or the rapidly replicating cancer cells. Nucleoside analogues 445 are mainly "chain terminators" and belong to the inhibitors described as anti-metabolites (Fig. 446 8A). After penetration in the cell, the nucleoside analogues are tri-phosphorylated by viral or 447 human cells. The triphosphorylated compounds are then recognized by polymerases and 448 incorporated into the growing DNA/RNA chain. During DNA/RNA chain elongation, the 449 (n+1) nucleoside triphosphate is attached to the 3'-OH of the *n* terminal nucleotide of the 450 growing chain. The 3'-OH group is thus essential for chain elongation and "obligate chain 451 terminators" nucleoside analogues lack the 3'-OH group so that their incorporation 452 consequently results in chain termination.[134] Either the 3'-OH group is substituted with a 453 non-reactive group (-H, -Cl, -F, -Br and -N<sub>3</sub> for example) or the deoxyribose is replaced by 454 arabinose derivatives. In general, only few modifications are performed on the nucleobase in order to facilitate the incorporation of the nucleoside analogues as its natural counterpart.[135] 455

- 456 Several adenosine analogues are used as "chain terminators" for the treatment of cancer or
- 457 viral infection.



458

459 Figure 8. A) Schematic representation of the main mechanisms of nucleoside analogues to inhibit DNA/RNA
460 synthesis; B) Examples of adenosine analogues as antiviral and anticancer agents. (Biorender.com was used to
461 prepare panel A)

462

463 *6.1. Antiviral agents* 

464 One example of chain terminator antiviral agent is Adefovir dipivoxil, 29 (Hepsera®),
465 that was approved by the FDA in 2002 for the treatment of Hepatis B Virus (HBV) (Fig.
466 8B).[136] It is a ester prodrug of a viral replication terminator Adefovir that blocks the HBV

467 reverse transcriptase and is the prototype of acyclic adenosine phosphonate analogues. The 468 phosphonate bond (-PC-) mimics the 5'-monophosphate of natural nucleotides and enables 469 Adefovir to be phosphorylated by kinases to phosphono-diphosphate form. Adefovir 470 diphosphate competes with the natural substrate dATP and its incorporation results in blockage 471 of the viral polymerase causing chain termination. The phosphonate bond is more stable than 472 the phosphate bond and cannot be cleaved by phosphodiesterases.[137] Based on this efficient 473 strategy, several similar acyclic nucleoside phosphonate compounds have been optimized. In 474 parallel, some nucleoside analogues display a "delayed chain termination" mechanism.[138] 475 These compounds exhibit a locked sugar conformation, and after their incorporation in the 476 growing chain, two or three other nucleotides can be further incorporated. Then, due to steric 477 hindrance and displacement of the growing chain in the catalytic site, strand elongation is 478 impeded.[134] This mechanism was developed to avoid exonucleolytic removal of nucleoside 479 analogues. For example, Remdesivir, **30** (Veklury ®), is a delayed chain terminator that has 480 been recently approved by the FDA for the treatment of COVID-19.[139, 140] More precisely, 481 Remdesivir is a prodrug of adenosine, the 1'-cyano group avoids the translocation of 482 coronavirus polymerases. According to molecular modeling data, repulsion between the cyano 483 group and S861 in the protein pocket would inhibit the polymerase translocation to position 484 (*n*+4).[141]

485

#### 486 **6.2.** Anticancer agents

Fludarabine, **31** (F-ara-A), was approved by the FDA for the treatment of B-cell chronic lymphotic leukemia (Fig. 8B). After cell penetration in its nucleoside form, Fludarabine is triphosphorylated (forming F-ara-ATP) and competes with the natural substrate. Incorporation of F-ara-ATP into DNA by DNA polymerase triggers chain termination.[142] Then, to improve solubility, the 5'-monophosphate derivative, **32** (Fludara®), was designed. Fludara® is then

converted into fludarabine by phosphatase (Fig. 8B). Clofarabine, 33 (Clolar<sup>TM</sup>, Genzyme), a 492 493 next-generation deoxyadenosine analogue, was approved by the FDA in 2004 to treat relapsed or refractory Acute Lymphoblastic Leukemia (ALL) (Fig. 8B). Clofarabine showed an 494 495 improved stability in the acidic gastric environment and in plasma, notably thanks to the 496 fluorine group at the 2'-position that increases its stability at acidic pH. After its 497 triphosphorylation by kinases, Clofarabine competes with the natural dATP for binding to 498 DNA polymerase  $-\alpha$  and  $-\varepsilon$ . At high ratios of Clofarabine/dATP, Clofarabine is inserted into 499 internal and terminal sites of DNA and inhibits DNA elongation.[143] The anticancer activity 500 of Clofarabine is also due to the inhibition of ribonucleotide reductase and induction of 501 apoptosis.[144] These compounds belong to the antimetabolites that are used in the treatment 502 of cancer based on the higher proliferation of the cancer cells, however they target all 503 proliferating cells showing the well-known side effects (*e.g.* hair loss).

504

#### 505 Conclusion and Perspectives

506

507 This review highlights the role of the adenine in the biology of cells and shows through 508 examples the design of adenine-based drugs and their use to treat human diseases, such as 509 cancer, neurological or infectious diseases. The adenine scaffold is involved in many essential 510 biological processes and participate to a broad range of mechanisms, as energy source, 511 cofactor, ... As these mechanisms are aberrant in diseases, they constitute potential targets for 512 treatment and thus the adenosine scaffold is a chemical starting point for the design of drugs in 513 medicinal chemistry. Many chemical modifications have been carried out on the heterocycle 514 moiety and on the side chain giving extensive chemical libraries of adenine derivatives with 515 comprehensive SAR studies. Moreover, it enabled to develop and investigate novel methods 516 in purine synthesis.[145–147]



517

Figure 9. A) Schematic representation of the STING pathway; B) Example of fleximer of adenosine; C)
Examples of cGAMP analogues and D) Examples of adenosine derivatives used as fluorescent probes.
(Biorender.com was used to prepare panel B.)

521

522 Importantly, membrane permeability and bioavailability remain an issue and many 523 efforts are carried out to improve the pharmacological properties and extend the adenine analogues repertoire and explore adenine prodrugs. For example, adenosine fleximers, in which 524 525 the purine base is split into two heterocycles connected by a bond allowing rotation, have been 526 developed (Fig. 9A).[148] Thanks to their flexible structure, these derivatives have shown 527 advantages compared to the corresponding purine-base nucleosides, as they can exert new 528 interactions with the protein partner that are not possible with the corresponding nucleoside or 529 their extended structure can give access to new space in the protein pocket.[149] Indeed, the

replacement of a guanine by fleximer in Acyclovir greatly improved its biological activity and the ability of the fleximer analogue to adopt the *syn* conformation was able to inhibit efficiently a SAH hydrolase.[150] These compounds have many promising applications in particular for their antiviral activity but also as molecular probes and antitumor agents.[150]

534 Another emerging and promising topic consists of the development of new modified 535 cyclic dinucleotides targeting STimulator of INterferon Genes protein (STING). STING 536 recognizes 2'3' cyclic GMP-AMP (cGAMP), which is produced by the cyclic GMP-AMP 537 synthase (cGAS) after recognition of cytosolic endogenous or exogenous DNA (Fig. 9B).[151] 538 The STING activation induces a signaling cascade and transcription of antiviral and 539 proinflammatory genes.[152] Several cyclic dinucleotides with adenosine derivatives are 540 developed to activate STING (Fig. 9C), by mimicking cGAMP.[153] Such compounds should 541 be further explored and could be used to improve vaccination, control autoimmune disease or 542 treat cancer.

543 It is worth mentioning that the quinazoline core is also a common mimic of adenine and 544 used more broadly in the design of drugs as we can see in many kinase and DNA 545 methyltransferases inhibitors for example.

Finally, the adenine scaffold is also used to develop fluorescent probes. To confer and modulate fluorescence properties, modifications are mainly carried out on the C8-position of adenine (and few examples with modifications on C2-position) (Fig. 9D). By mimicking the natural non-fluorescent adenine, these compounds have many biological applications such as probing the structure of nucleic acids and their interactions with biological molecules.[154– 156] Finally, purine chemical libraries are extended with the guanine scaffold that is also used in many biological applications including anticancer and antiviral therapy.[5,157]

553 The interest in the design of analogues of cofactors (SAM, NAD, ...) is increasing for 554 both anticancer, antiviral and antibiotics applications. Interesting, the advances in the

technologies to detect RNA modifications (epitranscriptomics) pointed out their role in diseases, increasing the research on the design of analogues to target SAM-based enzymes, ribozymes, deoxyribozymes.[158–162]

To conclude, adenine plays a major role in various biological processes but is also involved in processes that are aberrant in diseases. Therefore, it is worth continuing to develop new inhibitors based on this scaffold.

561

#### 562 **Abbreviations**

- 563 AC=Adenylyl Cyclase
- 564 Acetyl-CoA= Acetyl-CoenzymeA
- 565 ALS=Amyotrophic Lateral Sclerosis
- 566 AR=Adenosine Receptors
- 567 ATP= Adenosine TriPhosphate
- 568 cAMP= cyclic Adenosine MonoPhosphate
- 569 5mC= 5-methylCytosine
- 570 BTK= Burton's Tyrosine Kinase
- 571 CAD= Coronary Artery Disease
- 572 CLL= Chronic Lymphocytic Leukemia
- 573 CNG= Cyclic Nucleotide-Gated channels
- 574 COMT= Catechol *O*-MethylTransferases
- 575 DNMT= C5 DNA MethylTransferases
- 576 EPAC= Exchange Protein Activated by cAMP
- 577 EGFR= Epidermal Growth Factor Receptor
- 578 FAD= Flavin Adenine Dinucleotide
- 579 FDA=U.S Food and Drug Administration

- 580 cGAS= cyclic GMP-AMP Synthase
- 581 cGMP= 2'3' cyclic GMP-AMP
- 582 GPCR= G-Protein-Coupled Receptors
- 583 HBV= Hepatis B Virus
- 584 HCC= HepatoCellular Carcinoma
- 585 Her2= Human epidermal growth factor receptor 2
- 586 HMT= Histone MethylTransferases
- 587 KMT= Lysine MethylTransferase
- 588 MLL= Mix Lineage Leukemia
- 589 MPI= Myocardial Potential Imaging
- 590 MTase= MethylTransferases
- 591 NAD= Nicotinamide Adenine Dinucleotide
- 592 NADK= Nicotinamide Adenine Dinucleotide Kinases
- 593 NNMT= Nicotinamide *N*-MethylTransferase
- 594 NRTK= Non-Receptor protein Tyrosine Kinase
- 595 NSCLC= Non-Small Cell Lung Carcinoma
- 596 NTMT= *N*-Terminal MethylTransferases
- 597 PDE= PhosphoDiEsterases
- 598 PI3K $\delta$ = PhosphoInositide 3-Kinase  $\delta$
- 599 PKA= Protein Kinase A
- 600 KMT= Lysine MethylTransferase
- 601 PRMT= Protein arginine MethylTransferases
- 602 SAH= S-Adenosyl-L-Homocystein
- 603 SAM= S-Adenosyl-L-Methionine (or AdoMet)
- 604 sSAR= Structure-Activity Relationships

| 605 | STING= STimulator of INterferon Genes protein.   |
|-----|--|
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| 607 |  |
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| 618 |  |
| 619 |  |
| 620 |  |
| 621 | Note about the authors   |
| 622 | The authors are three women deeply involved in science. Mrs Alexandra Fillion is a PhD       |
| 623 | student of the Ecole Normale Supérieure de Paris; Dr Sophie Vichier Guerre is a senior       |
| 624 | scientist expert in the Chemistry of nucleobases, nucleotides and nucleic acids; Dr Paola B. |
| 625 | Arimondo, director of research, leads research projects at the interface of Chemistry and    |
| 626 | Biology.   |
| 627 |  |
| 628 |  |

#### 629 **References**

- 630
- 631 [1] D. Fabbro, S. W. Cowan-Jacob, H. Moebitz, Br. J. Pharmacol. 2015, 172 (11), 2675-
- 632 2700. DOI: https://doi.org/10.1111/bph.13096.
- 633 [2] Q. Sun, M. Huang, Y. Wei, Acta Pharm. Sin. B. 2021, 11 (3), 632–650. DOI:
- 634 https://doi.org/10.1016/j.apsb.2020.08.011.
- [3] A. J. Covarrubias, R. Perrone, A. Grozio, E. Verdin, *Nat. Rev. Mol. Cell Biol.* 2021, 22
  (2), 119–141. DOI: https://doi.org/10.1038/s41580-020-00313-x.
- 637 [4] C. Patra, K. Foster, J. E. Corley, M. Dimri, M. F. Brady, in *StatPearls*, StatPearls
- 638 Publishing, Treasure Island (FL) 2023.
- 639 [5] X. Lin, C. Liang, L. Zou, Y. Yin, J. Wang, D. Chen, W. Lan, *Eur. J. Med. Chem.* 2021,
- 640 **214**, 113233. DOI: https://doi.org/10.1016/j.ejmech.2021.113233.
- 641 [6] S. Man, Y. Lu, L. Yin, X. Cheng, L. Ma, *Drug Discov. Today.* 2021, 26 (6), 1490–1500.
  642 DOI: https://doi.org/10.1016/j.drudis.2021.02.020.
- 643 [7] C. Wang, Z. Song, H. Yu, K. Liu, X. Ma, *Acta Pharm. Sin. B.* 2015, 5 (5), 431–441. DOI:
  644 https://doi.org/10.1016/j.apsb.2015.07.002.
- [8] P. Wu, T. E. Nielsen, M. H. Clausen, *Trends Pharmacol. Sci.* 2015, 36 (7), 422–439. DOI:
  https://doi.org/10.1016/j.tips.2015.04.005.
- 647 [9] M. Vieito, V. Moreno, A. Spreafico, I. Brana, J. S. Wang, M. Preis, T. Hernández, S.
- 648 Genta, A. R. Hansen, B. Doger, et al., Clin. Cancer Res. 2023, 29 (18), 3592–3602.
- 649 DOI: https://doi.org/10.1158/1078-0432.CCR-23-0092.
- 650 [10] A. A. Zenchenko, M. S. Drenichev, I. A. Il'icheva, S. N. Mikhailov, Mol. Biol. 2021,
- 651 **55** (*6*), 786–812. DOI: https://doi.org/10.1134/S0026893321040105.
- 652 [11] J. Shelton, X. Lu, J. A. Hollenbaugh, J. H. Cho, F. Amblard, R. F. Schinazi, Chem. Rev.
- 653 2016, **116** (*23*), 14379–14455. DOI: https://doi.org/10.1021/acs.chemrev.6b00209.

- 654 [12] M. Gelin, J. Paoletti, M.-A. Nahori, V. Huteau, C. Leseigneur, G. Jouvion, L. Dugué,
- D. Clément, J.-L. Pons, L. Assairi, *et al.*, *ACS Infect. Dis.* 2020, 6 (3), 422–435. DOI:
  https://doi.org/10.1021/acsinfecdis.9b00368.
- 657 [13] P. Wang, Z. Liu, H. Chen, N. Ye, X. Cheng, J. Zhou, Bioorg. Med. Chem. Lett. 2017,
- 658 **27** (8), 1633–1639. DOI: https://doi.org/10.1016/j.bmcl.2017.02.065.
- 659 [14] B. Kutryb-Zając, A. Kawecka, K. Nasadiuk, A. Braczko, K. Stawarska, E. Caiazzo, P.
- 660 Koszałka, C. Cicala, *Biomed. Pharmacother.* 2023, **165**, 115184. DOI:
- 661 https://doi.org/10.1016/j.biopha.2023.115184.
- 662 [15] J. Martin, Curr. Opin. Struct. Biol. 2002, 12 (6), 783–793. DOI:
- 663 https://doi.org/10.1016/S0959-440X(02)00391-3.
- 664 [16] Y. Dor, H. Cedar, *The Lancet*. 2018, **392** (*10149*), 777–786. DOI:
- 665 https://doi.org/10.1016/S0140-6736(18)31268-6.
- 666 [17] Y. Zhou, Y. Kong, W. Fan, T. Tao, Q. Xiao, N. Li, X. Zhu, *Biomed. Pharmacother*.
- 667 2020, **131**, 110731. DOI: https://doi.org/10.1016/j.biopha.2020.110731.
- 668 [18] X. Zhang, H. Wen, X. Shi, Acta Biochim. Biophys. Sin. 2012, 44 (1), 14–27. DOI:
- 669 https://doi.org/10.1093/abbs/gmr100.
- 670 [19] A. Roberti, A. F. Fernández, M. F. Fraga, *Mol. Metab.* 2021, 45, 101165. DOI:
- 671 https://doi.org/10.1016/j.molmet.2021.101165.
- 672 [20] E. Abdelraheem, B. Thair, R. F. Varela, E. Jockmann, D. Popadić, H. C. Hailes, J. M.
- 673 Ward, A. M. Iribarren, E. S. Lewkowicz, J. N. Andexer, et al., ChemBioChem. 2022,
- 674 **23** (*18*), e202200212. DOI: https://doi.org/10.1002/cbic.202200212.
- 675 [21] A. Struck, M. L. Thompson, L. S. Wong, J. Micklefield, ChemBioChem. 2012, 13 (18),
- 676 2642–2655. DOI: https://doi.org/10.1002/cbic.201200556.
- 677 [22] S. Jahan, J. R. Davie, Adv. Biol. Regul. 2015, 57, 173–184. DOI:
- 678 https://doi.org/10.1016/j.jbior.2014.09.003.

- 679 [23] K. Hyun, J. Jeon, K. Park, J. Kim, *Exp. Mol. Med.* 2017, 49 (4), e324–e324. DOI:
  680 https://doi.org/10.1038/emm.2017.11.
- 681 [24] R. Di Blasi, O. Blyuss, J. F. Timms, D. Conole, F. Ceroni, H. J. Whitwell, ACS Chem.
- 682 *Biol.* 2021, **16** (2), 238–250. DOI: https://doi.org/10.1021/acschembio.0c00771.
- 683 [25] R. Huang, *ChemBioChem*. 2019, **20** (8), 976–984. DOI:
- 684 https://doi.org/10.1002/cbic.201800615.
- 685 [26] K. Diaz, Y. Meng, R. Huang, *Curr. Opin. Chem. Biol.* 2021, 63, 115–122. DOI:
  686 https://doi.org/10.1016/j.cbpa.2021.02.017.
- 687 [27] M. Lopez, J. Gilbert, J. Contreras, L. Halby, P. B. Arimondo, in DNA
- 688 *Methyltransferases Role Funct.* (Eds: A. Jeltsch, R. Z. Jurkowska), Vol. 1389,
- 689 Springer International Publishing, Cham 2022.
- 690 [28] P. Bastos, T. Gomes, L. Ribeiro, in Rev. Physiol. Biochem. Pharmacol. (Eds: B. Nilius,
- 691 P. De Tombe, T. Gudermann, R. Jahn, R. Lill, O. H. Petersen), Vol. 173, Springer
- 692 International Publishing, Cham 2017.
- 693 [29] J. Zhang, Y. G. Zheng, ACS Chem. Biol. 2016, 11 (3), 583–597. DOI:
- 694 https://doi.org/10.1021/acschembio.5b00812.
- 695 [30] S. K. Tewary, Y. G. Zheng, M.-C. Ho, Cell. Mol. Life Sci. 2019, 76 (15), 2917–2932.
- 696 DOI: https://doi.org/10.1007/s00018-019-03145-x.
- 697 [31] M. E. Neganova, S. G. Klochkov, Y. R. Aleksandrova, G. Aliev, Semin. Cancer Biol.
- 698 2022, **83**, 452–471. DOI: https://doi.org/10.1016/j.semcancer.2020.07.015.
- 699 [32] M. Barbachowska, P. B. Arimondo, *Epigenetics*. 2023, 18 (1), 2242689. DOI:
  700 https://doi.org/10.1080/15592294.2023.2242689.
- 701 [33] M. Y. Wang, P. Liow, M. I. T. Guzman, J. Qi, ACS Chem. Biol. 2022, 17 (4), 744–755.
- 702 DOI: https://doi.org/10.1021/acschembio.2c00062.

- 703 [34] M. J. G. Eldridge, M. A. Hamon, *PLOS Pathog.* 2021, 17 (*12*), e1010173. DOI:
  704 https://doi.org/10.1371/journal.ppat.1010173.
- 705 [35] D. Schator, S. Mondino, J. Berthelet, C. Di Silvestre, M. Ben Assaya, C. Rusniok, F.
- 706 Rodrigues-Lima, A. Wehenkel, C. Buchrieser, M. Rolando, *Nat. Commun.* 2023, 14
- 707 (1), 2154. DOI: https://doi.org/10.1038/s41467-023-37885-z.
- 708 [36] A. Talukdar, A. Mukherjee, D. Bhattacharya, J. Med. Chem. 2022, 65 (3), 1662–1684.
  709 DOI: https://doi.org/10.1021/acs.jmedchem.1c01208.
- 710 [37] C. Bon, Y. Si, P. B. Arimondo, in *Histone Modifications in Therapy*, Vol 20, Elsevier
- 711 2020. DOI: https://doi.org/10.1016/B978-0-12-816422-8.00005-2
- 712 [38] Y. Yao, P. Chen, J. Diao, G. Cheng, L. Deng, J. L. Anglin, B. V. V. Prasad, Y. Song, J.
- 713 *Am. Chem. Soc.* 2011, **133** (*42*), 16746–16749. DOI:
- 714 https://doi.org/10.1021/ja206312b.
- 715 [39] S. R. Daigle, E. J. Olhava, C. A. Therkelsen, C. R. Majer, C. J. Sneeringer, J. Song, L.
- 716 D. Johnston, M. P. Scott, J. J. Smith, Y. Xiao, et al., Cancer Cell. 2011, 20 (1), 53–65.
- 717 DOI: https://doi.org/10.1016/j.ccr.2011.06.009.
- 718 [40] A. Basavapathruni, L. Jin, S. R. Daigle, C. R. A. Majer, C. A. Therkelsen, T. J. Wigle,
- 719 K. W. Kuntz, R. Chesworth, R. M. Pollock, M. P. Scott, et al., Chem. Biol. Drug Des.
- 720 2012, **80** (*6*), 971–980. DOI: https://doi.org/10.1111/cbdd.12050.
- 721 [41] S. R. Daigle, E. J. Olhava, C. A. Therkelsen, A. Basavapathruni, L. Jin, P. A. Boriack-
- 722 Sjodin, C. J. Allain, C. R. Klaus, A. Raimondi, M. P. Scott, et al., Blood. 2013, 122 (6),
- 723 1017–1025. DOI: https://doi.org/10.1182/blood-2013-04-497644.
- 124 [42] H. Lin, M. Wang, Y. W. Zhang, S. Tong, R. A. Leal, R. Shetty, K. Vaddi, J. I. Luengo,
- 725 ACS Med. Chem. Lett. 2019, **10** (7), 1033–1038. DOI:
- https://doi.org/10.1021/acsmedchemlett.9b00074.

- 727 [43] W. Ren, L. Gao, J. Song, *Genes*. 2018, 9 (12), 620. DOI:
- 728 https://doi.org/10.3390/genes9120620.
- [44] E. Kaminskas, A. T. Farrell, Y.-C. Wang, R. Sridhara, R. Pazdur, The Oncologist. 2005,
- 730 **10** (*3*), 176–182. DOI: https://doi.org/10.1634/theoncologist.10-3-176.
- 731 [45] S. D. Gore, C. Jones, P. Kirkpatrick, Nat. Rev. Drug Discov. 2006, 5 (11), 891–892.
- 732 DOI: https://doi.org/10.1038/nrd2180.
- 733 [46] M. B. Pappalardi, K. Keenan, M. Cockerill, W. A. Kellner, A. Stowell, C. Sherk, K.
- 734 Wong, S. Pathuri, J. Briand, M. Steidel, *et al.*, *Nat. Cancer.* 2021, **2** (10), 1002–1017.
- 735 DOI: https://doi.org/10.1038/s43018-021-00249-x.
- 736 [47] Q. Chen, B. Liu, Y. Zeng, J. W. Hwang, N. Dai, I. R. Corrêa, M. R. Estecio, X. Zhang,
- 737 M. A. Santos, T. Chen, *et al.*, *NAR Cancer*. 2023, **5** (2), zcad022. DOI:
- 738 https://doi.org/10.1093/narcan/zcad022.
- [48] L. Isakovic, O. M. Saavedra, D. B. Llewellyn, S. Claridge, L. Zhan, N. Bernstein, A.
- 740 Vaisburg, N. Elowe, A. J. Petschner, J. Rahil, et al., Bioorg. Med. Chem. Lett. 2009, 19
- 741 (10), 2742–2746. DOI: https://doi.org/10.1016/j.bmcl.2009.03.132.
- 742 [49] O. M. Saavedra, L. Isakovic, D. B. Llewellyn, L. Zhan, N. Bernstein, S. Claridge, F.
- Raeppel, A. Vaisburg, N. Elowe, A. J. Petschner, et al., Bioorg. Med. Chem. Lett. 2009,
- 744 **19** (10), 2747–2751. DOI: https://doi.org/10.1016/j.bmcl.2009.03.113.
- 745 [50] C. Bon, L. Halby, P. B. Arimondo, *Epigenomics*. 2020, **12** (*17*), 1479–1482. DOI:
  746 https://doi.org/10.2217/epi-2020-0203.
- 747 [51] A. Bouchut, D. Rotili, C. Pierrot, S. Valente, S. Lafitte, J. Schultz, U. Hoglund, R.
- 748 Mazzone, A. Lucidi, G. Fabrizi, et al., Eur. J. Med. Chem. 2019, 161, 277–291. DOI:
- 749 https://doi.org/10.1016/j.ejmech.2018.10.041.

- 750 [52] D. Rotili, D. Tarantino, B. Marrocco, C. Gros, V. Masson, V. Poughon, F. Ausseil, Y.
- 751 Chang, D. Labella, S. Cosconati, *et al.*, *PLoS ONE*. 2014, **9** (5), e96941. DOI:
- 752 https://doi.org/10.1371/journal.pone.0096941.
- 753 [53] F. Nardella, L. Halby, E. Hammam, D. Erdmann, V. Cadet-Daniel, R. Peronet, D.
- 754 Ménard, B. Witkowski, S. Mecheri, A. Scherf, et al., ACS Cent. Sci. 2020, 6 (1), 16–
- 755 21. DOI: https://doi.org/10.1021/acscentsci.9b00874.
- 756 [54] L. Halby, Y. Menon, E. Rilova, D. Pechalrieu, V. Masson, C. Faux, M. A. Bouhlel, M.-
- H. David-Cordonnier, N. Novosad, Y. Aussagues, et al., J. Med. Chem. 2017, 60 (11),
- 758 4665–4679. DOI: https://doi.org/10.1021/acs.jmedchem.7b00176.
- 759 [55] C. E. Schaner Tooley, J. J. Petkowski, T. L. Muratore-Schroeder, J. L. Balsbaugh, J.
- 760 Shabanowitz, M. Sabat, W. Minor, D. F. Hunt, I. G. Macara, *Nature*. 2010, 466 (7310),
- 761 1125–1128. DOI: https://doi.org/10.1038/nature09343.
- 762 [56] R. Huang, *ChemBioChem*. 2019, **20** (8), 976–984. DOI:
- 763 https://doi.org/10.1002/cbic.201800615.
- 764 [57] G. Zhang, S. L. Richardson, Y. Mao, R. Huang, Org. Biomol. Chem. 2015, 13 (14),
- 765 4149–4154. DOI: https://doi.org/10.1039/C5OB00120J.
- 766 [58] D. Chen, G. Dong, N. Noinaj, R. Huang, J. Med. Chem. 2019, 62 (7), 3773–3779.
- 767 DOI: https://doi.org/10.1021/acs.jmedchem.9b00206.
- 768 [59] M. J. Van Haren, J. Sastre Toraño, D. Sartini, M. Emanuelli, R. B. Parsons, N. I.
- 769 Martin, *Biochemistry*. 2016, *55* (*37*), *5307–5315*. DOI:
- 770 https://doi.org/10.1021/acs.biochem.6b00733.
- 771 [60] H. Sperber, J. Mathieu, Y. Wang, A. Ferreccio, J. Hesson, Z. Xu, K. A. Fischer, A.
- 772 Devi, D. Detraux, H. Gu, et al., Nat. Cell Biol. 2015, 17 (12), 1523–1535. DOI:
- 773 https://doi.org/10.1038/ncb3264.

- 774 [61] D. Sartini, S. Morganti, E. Guidi, C. Rubini, A. Zizzi, R. Giuliante, V. Pozzi, M.
- 775 Emanuelli, *Cell Biochem. Biophys.* 2013, **67** (*3*), 865–873. DOI:
- 776 https://doi.org/10.1007/s12013-013-9574-z.
- 777 [62] R. B. Parsons, M.-L. Smith, A. C. Williams, R. H. Waring, D. B. Ramsden, J.
- 778 *Neuropathol. Exp. Neurol.* 2002, **61** (2), 111–124. DOI:
- 779 https://doi.org/10.1093/jnen/61.2.111.
- 780 [63] D. Kraus, Q. Yang, D. Kong, A. S. Banks, L. Zhang, J. T. Rodgers, E. Pirinen, T. C.
- 781 Pulinilkunnil, F. Gong, Y. Wang, et al., Nature. 2014, **508** (7495), 258–262. DOI:
- 782 https://doi.org/10.1038/nature13198.
- 783 [64] M. J. Van Haren, R. Taig, J. Kuppens, J. Sastre Toraño, E. E. Moret, R. B. Parsons, D.
- Sartini, M. Emanuelli, N. I. Martin, *Org. Biomol. Chem.* 2017, **15** (*31*), 6656–6667.
  DOI: https://doi.org/10.1039/C7OB01357D.
- 786 [65] Y. Gao, M. J. Van Haren, E. E. Moret, J. J. M. Rood, D. Sartini, A. Salvucci, M.
- 787 Emanuelli, P. Craveur, N. Babault, J. Jin, et al., J. Med. Chem. 2019, 62 (14), 6597-
- 788 6614. DOI: https://doi.org/10.1021/acs.jmedchem.9b00413.
- 789 [66] N. Babault, A. Allali-Hassani, F. Li, J. Fan, A. Yue, K. Ju, F. Liu, M. Vedadi, J. Liu, J.
- 790 Jin, J. Med. Chem. 2018, **61** (4), 1541–1551. DOI:
- 791 https://doi.org/10.1021/acs.jmedchem.7b01422.
- 792 [67] R. L. Policarpo, L. Decultot, E. May, P. Kuzmič, S. Carlson, D. Huang, V. Chu, B. A.
- 793 Wright, S. Dhakshinamoorthy, A. Kannt, et al., J. Med. Chem. 2019, 62 (21), 9837-
- 794 9873. DOI: https://doi.org/10.1021/acs.jmedchem.9b01238.
- 795 [68] Y. Gao, M. J. Van Haren, N. Buijs, P. Innocenti, Y. Zhang, D. Sartini, R. Campagna, M.
- 796 Emanuelli, R. B. Parsons, W. Jespers, et al., J. Med. Chem. 2021, 64 (17), 12938-
- 797 12963. DOI: https://doi.org/10.1021/acs.jmedchem.1c01094.

- 798 [69] J. P. M. Finberg, J. Neural Transm. 2019, **126** (4), 433–448. DOI:
- 799 https://doi.org/10.1007/s00702-018-1952-7.
- 800 [70] C. Waters, J. Am. Geriatr. Soc. 2000, 48 (6), 692–698. DOI:
- 801 https://doi.org/10.1111/j.1532-5415.2000.tb04732.x.
- 802 [71] B. Masjost, P. Ballmer, E. Borroni, G. Zürcher, F. K. Winkler, R. Jakob-Roetne, F.
- 803 Diederich, *Chem. Eur. J.* 2000, **6** (*6*), 971–982. DOI:
- 804 https://doi.org/10.1002/(SICI)1521-3765(20000317)6:6%3C971::AID-
- 805 CHEM971%3E3.0.CO;2-0
- 806 [72] C. Lerner, B. Masjost, A. Ruf, V. Gramlich, R. Jakob-Roetne, G. Zürcher, E. Borroni,
- 807 F. Diederich, Org Biomol Chem. 2003, 1 (1), 42–49. DOI:
- 808 https://doi.org/10.1039/B208690P.
- 809 [73] R. Paulini, C. Lerner, R. Jakob-Roetne, G. Zürcher, E. Borroni, F. Diederich,
- 810 *ChemBioChem.* 2004, **5** (9), 1270–1274. DOI: https://doi.org/10.1002/cbic.200400084.
- 811 [74] M. Ellermann, R. Jakob-Roetne, C. Lerner, E. Borroni, D. Schlatter, D. Roth, A. Ehler,
- 812 M. G. Rudolph, F. Diederich, Angew. Chem. Int. Ed. 2009, 48 (48), 9092–9096. DOI:
- 813 https://doi.org/10.1002/anie.200904410.
- 814 [75] R. R. Naik, A. K. Shakya, Front. Pharmacol. 2023, 13, 1064472. DOI:
- 815 https://doi.org/10.3389/fphar.2022.1064472.
- 816 [76] R. Roskoski, *Pharmacol. Res.* 2023, **187**, 106552. DOI:
- 817 https://doi.org/10.1016/j.phrs.2022.106552.
- 818 [77] M. M. Attwood, D. Fabbro, A. V. Sokolov, S. Knapp, H. B. Schiöth, Nat. Rev. Drug
- 819 Discov. 2021, **20** (11), 839–861. DOI: https://doi.org/10.1038/s41573-021-00252-y.
- 820 [78] A. Markham, Drugs. 2014, 74 (14), 1701–1707. DOI: https://doi.org/10.1007/s40265-
- 821 014-0285-6.

- 822 [79] A. Berndt, S. Miller, O. Williams, D. D. Le, B. T. Houseman, J. I. Pacold, F. Gorrec,
- 823 W.-C. Hon, P. Ren, Y. Liu, et al., Nat. Chem. Biol. 2010, 6 (2), 117–124. DOI:
- 824 https://doi.org/10.1038/nchembio.293.
- 825 [80] J. R. Somoza, D. Koditek, A. G. Villaseñor, N. Novikov, M. H. Wong, A. Liclican, W.
- 826 Xing, L. Lagpacan, R. Wang, B. E. Schultz, et al., J. Biol. Chem. 2015, 290 (13),
- 827 8439–8446. DOI: https://doi.org/10.1074/jbc.M114.634683.
- 828 [81] P. Wu, Y. Hu, *MedChemComm*. 2012, **3** (11), 1337. DOI:
- 829 https://doi.org/10.1039/c2md20044a.
- 830 [82] N. Engl. J. Med. 2013, **369** (13), 1277–1279. DOI:
- 831 https://doi.org/10.1056/NEJMc1309710.
- 832 [83] A. T. Bender, A. Gardberg, A. Pereira, T. Johnson, Y. Wu, R. Grenningloh, J. Head, F.
- 833 Morandi, P. Haselmayer, L. Liu-Bujalski, *Mol. Pharmacol.* 2017, **91** (3), 208–219.
- B34 DOI: https://doi.org/10.1124/mol.116.107037.
- 835 [84] M. Guo, S. Dai, D. Wu, Y. Duan, J. Li, L. Qu, L. Jiang, Z. Chen, X. Chen, Y. Chen,
- 836 Bioorg. Med. Chem. Lett. 2021, **34**, 127757. DOI:
- 837 https://doi.org/10.1016/j.bmcl.2020.127757.
- 838 [85] G. Kim, A. E. McKee, Y.-M. Ning, M. Hazarika, M. Theoret, J. R. Johnson, Q. C. Xu,
- 839 S. Tang, R. Sridhara, X. Jiang, et al., Clin. Cancer Res. 2014, 20 (19), 4994–5000.
- 840 DOI: https://doi.org/10.1158/1078-0432.CCR-14-0776.
- 841 [86] G. Bollag, P. Hirth, J. Tsai, J. Zhang, P. N. Ibrahim, H. Cho, W. Spevak, C. Zhang, Y.
- 842 Zhang, G. Habets, et al., Nature. 2010, 467 (7315), 596–599. DOI:
- 843 https://doi.org/10.1038/nature09454.
- 844 [87] B. Agianian, E. Gavathiotis, J. Med. Chem. 2018, 61 (14), 5775–5793. DOI:
- 845 https://doi.org/10.1021/acs.jmedchem.7b01306.

- 846 [88] R. T. Dungo, G. M. Keating, Drugs. 2013, 73 (13), 1503–1515. DOI:
- 847 https://doi.org/10.1007/s40265-013-0111-6.
- 848 [89] F. Solca, G. Dahl, A. Zoephel, G. Bader, M. Sanderson, C. Klein, O. Kraemer, F.
- 849 Himmelsbach, E. Haaksma, G. R. Adolf, J. Pharmacol. Exp. Ther. 2012, 343 (2), 342–
- 850 350. DOI: https://doi.org/10.1124/jpet.112.197756.
- 851 [90] Z. Xie, X. Yang, Y. Duan, J. Han, C. Liao, J. Med. Chem. 2021, 64 (3), 1283–1345.
- 852 DOI: https://doi.org/10.1021/acs.jmedchem.0c01511.
- 853 [91] V. Palomo, V. Nozal, E. Rojas-Prats, C. Gil, A. Martinez, Br. J. Pharmacol. 2021, 178
- 854 (6), 1316–1335. DOI: https://doi.org/10.1111/bph.15221.
- 855 [92] E. T. McGuinness, J. R. Butler, Int. J. Biochem. 1985, 17 (1), 1–11. DOI:
- 856 https://doi.org/10.1016/0020-711X(85)90079-5.
- 857 [93] G. Magni, G. Orsomando, N. Raffaelli, *Mini-Rev. Med. Chem.* 2006, *6* (7), 739–746.
  858 DOI: https://doi.org/10.2174/138955706777698688.
- 859 [94] K. Kobayashi, S. D. Ehrlich, A. Albertini, G. Amati, K. K. Andersen, M. Arnaud, K.
- 860 Asai, S. Ashikaga, S. Aymerich, P. Bessieres, et al., Proc. Natl. Acad. Sci. 2003, 100
- 861 (8), 4678–4683. DOI: https://doi.org/10.1073/pnas.0730515100.
- 862 [95] C. M. Sassetti, D. H. Boyd, E. J. Rubin, *Mol. Microbiol.* 2003, 48 (1), 77–84. DOI:
  863 https://doi.org/10.1046/j.1365-2958.2003.03425.x.
- 864 [96] R. R. Chaudhuri, A. G. Allen, P. J. Owen, G. Shalom, K. Stone, M. Harrison, T. A.
- Burgis, M. Lockyer, J. Garcia-Lara, S. J. Foster, et al., BMC Genomics. 2009, 10 (1),
- 866 291. DOI: https://doi.org/10.1186/1471-2164-10-291.
- 867 [97] C. Mary, M. H. Soflaee, R. Kesavan, M. Gelin, H. Brown, G. Zacharias, T. P.
- 868 Mathews, A. Lemoff, C. Lionne, G. Labesse, et al., Mol. Cell. 2022, 82 (17), 3299-
- 869 3311.e8. DOI: https://doi.org/10.1016/j.molcel.2022.06.026.

- 870 [98] J. Paoletti, L. Assairi, M. Gelin, V. Huteau, M.-A. Nahori, O. Dussurget, G. Labesse, S.
- 871 Pochet, *Eur. J. Med. Chem.* 2016, **124**, 1041–1056. DOI:
- 872 https://doi.org/10.1016/j.ejmech.2016.10.033.
- 873 [99] G. Magni, M. Di Stefano, G. Orsomando, N. Raffaelli, S. Ruggieri, Curr. Med. Chem.
- 874 2009, **16** (*11*), 1372–1390. DOI: https://doi.org/10.2174/092986709787846505.
- 875 [100] A. Depaix, J. Kowalska, *Molecules*. 2019, 24 (22), 4187. DOI:
- 876 https://doi.org/10.3390/molecules24224187.
- [101] G. Poncet-Montange, L. Assairi, S. Arold, S. Pochet, G. Labesse, J. Biol. Chem. 2007,
- 878 **282** (*47*), 33925–33934. DOI: https://doi.org/10.1074/jbc.M701394200.
- 879 [102] M. Gelin, G. Poncet-Montange, L. Assairi, L. Morellato, V. Huteau, L. Dugué, O.
- 880 Dussurget, S. Pochet, G. Labesse, *Structure*. 2012, **20** (*6*), 1107–1117. DOI:
- 881 https://doi.org/10.1016/j.str.2012.03.024.
- 882 [103] J. Paoletti, L. Assairi, M. Gelin, V. Huteau, M.-A. Nahori, O. Dussurget, G. Labesse, S.
- 883 Pochet, Eur. J. Med. Chem. 2016, **124**, 1041–1056. DOI:
- 884 https://doi.org/10.1016/j.ejmech.2016.10.033.
- 885 [104] R. Kho, B. L. Baker, J. V. Newman, R. M. Jack, D. S. Sem, H. O. Villar, M. R. Hansen,
- 886 Proteins Struct. Funct. Bioinforma. 2003, **50** (4), 589–599. DOI:
- 887 https://doi.org/10.1002/prot.10316.
- 888 [105] M. Fujii, Y. Kitagawa, S. Iida, K. Kato, M. Ono, *Bioorg. Med. Chem. Lett.* 2015, 25
- 889 (22), 5133–5136. DOI: https://doi.org/10.1016/j.bmcl.2015.10.005.
- 890 [106] D. A. Clément, M. Gelin, C. Leseigneur, V. Huteau, L. Mondange, J.-L. Pons, O.
- 891 Dussurget, C. Lionne, G. Labesse, S. Pochet, *Eur. J. Med. Chem.* 2023, 246, 114941.
- 892 DOI: https://doi.org/10.1016/j.ejmech.2022.114941.
- 893 [107] S. Sheth, R. Brito, D. Mukherjea, L. Rybak, V. Ramkumar, Int. J. Mol. Sci. 2014, 15
- 894 (2), 2024–2052. DOI: https://doi.org/10.3390/ijms15022024.

- 895 [108] F. Vincenzi, S. Pasquini, C. Contri, M. Cappello, M. Nigro, A. Travagli, S. Merighi, S.
- 896 Gessi, P. A. Borea, K. Varani, *Biomolecules*. 2023, **13** (9), 1387. DOI:
- 897 https://doi.org/10.3390/biom13091387.
- 898 [109] A. Saini, R. Patel, S. Gaba, G. Singh, G. D. Gupta, V. Monga, Eur. J. Med. Chem.
- 899 2022, **227**, 113907. DOI: https://doi.org/10.1016/j.ejmech.2021.113907.
- 900 [110] K. O. Elkholy, O. Hegazy, A. Okunade, S. Aktas, T. Ajibawo, *Cureus*. 2021, **13** (1).
- 901 DOI: https://doi.org/10.7759/cureus.12940.
- 902 [111] H. Eggebrecht, Vasc. Health Risk Manag. 2008, 4, 337–340. DOI:
- 903 https://doi.org/10.2147/VHRM.S1798.
- 904 [112] J. Rabin, Y. Zhao, E. Mostafa, M. Al-Suqi, E. Fleischmann, M. R. Conaway, B. J.
- 905 Mann, P. Chhabra, K. L. Brayman, A. Krupnick, *et al.*, *PLOS ONE*. 2023, 18 (8),
  906 e0288920. DOI: https://doi.org/10.1371/journal.pone.0288920.
- 907 [113] P. Fishman, S. Cohen, *Clin. Rheumatol.* 2016, **35** (9), 2359–2362. DOI:
- 908 https://doi.org/10.1007/s10067-016-3202-4.
- 909 [114] K. A. Jacobson, S. Merighi, K. Varani, P. A. Borea, S. Baraldi, M. Aghazadeh Tabrizi,
- 910 R. Romagnoli, P. G. Baraldi, A. Ciancetta, D. K. Tosh, et al., Med. Res. Rev. 2018, 38
- 911 (4), 1031–1072. DOI: https://doi.org/10.1002/med.21456.
- 912 [115] P. Fishman, S. M. Stemmer, A. Bareket-Samish, M. H. Silverman, W. D. Kerns,
- 913 Purinergic Signal. 2023, 19 (3), 513–522. DOI: https://doi.org/10.1007/s11302-023914 09925-2.
- 915 [116] C. Gallo-Rodriguez, X. Ji, N. Melman, B. D. Siegman, L. H. Sanders, J. Orlina, B.
- 916 Fischer, Q. Pu, M. E. Olah, J. Med. Chem. 1994, **37** (5), 636–646. DOI:
- 917 https://doi.org/10.1021/jm00031a014.

- 918 [117] S. Bar-Yehuda, L. Rath-Wolfson, L. Del Valle, A. Ochaion, S. Cohen, R. Patoka, G.
- 2019 Zozulya, F. Barer, E. Atar, S. Piña-Oviedo, *et al.*, *Arthritis Rheum*. 2009, **60** (10),
  3061–3071. DOI: https://doi.org/10.1002/art.24817.
- 921 [118] M. David, L. Akerman, M. Ziv, M. Kadurina, D. Gospodinov, F. Pavlotsky, R.
- 922 Yankova, V. Kouzeva, M. Ramon, M. H. Silverman, et al., J. Eur. Acad. Dermatol.
- 923 Venereol. 2012, 26 (3), 361–367. DOI: https://doi.org/10.1111/j.1468-
- 924 3083.2011.04078.x.
- 925 [119] P. Fishman, *Molecules*. 2022, **27** (*12*), 3680. DOI:
- 926 https://doi.org/10.3390/molecules27123680.
- 927 [120] P. Fishman, Int. J. Oncol. 1992. DOI: https://doi.org/10.3892/ijo\_00000008.
- 928 [121] S. Cohen, S. M. Stemmer, G. Zozulya, A. Ochaion, R. Patoka, F. Barer, S. Bar-Yehuda,
- L. Rath-Wolfson, K. A. Jacobson, P. Fishman, J. Cell. Physiol. 2011, 226 (9), 2438–
  2447. DOI: https://doi.org/10.1002/jcp.22593.
- 931 [122] A. Bhadra, J. L. Hewes, A. Scruggs, C. Zhou, J. Y. Lee, N. Bauer, Adv. Biol. 2021, 5

932 (12), 2101064. DOI: https://doi.org/10.1002/adbi.202101064.

- 933 [123] M. Kamenetsky, S. Middelhaufe, E. M. Bank, L. R. Levin, J. Buck, C. Steegborn, J.
   934 *Mol. Biol.* 2006, 362 (4), 623–639. DOI: https://doi.org/10.1016/j.jmb.2006.07.045.
- 935 [124] D. A. Walsh, J. P. Perkins, E. G. Krebs, J. Biol. Chem. 1968, 243 (13), 3763-3765.
- 936 [125] J. De Rooij, F. J. T. Zwartkruis, M. H. G. Verheijen, R. H. Cool, S. M. B. Nijman, A.
- 937 Wittinghofer, J. L. Bos, *Nature*. 1998, **396** (6710), 474–477. DOI:
- 938 https://doi.org/10.1038/24884.
- 939 [126] K. W. Yau, Proc. Natl. Acad. Sci. 1994, 91 (9), 3481–3483. DOI:
- 940 https://doi.org/10.1073/pnas.91.9.3481.
- 941 [127] M. G. Gold, T. Gonen, J. D. Scott, J. Cell Sci. 2013, **126** (20), 4537–4543. DOI:
- 942 https://doi.org/10.1242/jcs.133751.

- 943 [128] F. Lezoualc'h, L. Fazal, M. Laudette, C. Conte, Circ. Res. 2016, 118 (5), 881–897.
- 944 DOI: https://doi.org/10.1161/CIRCRESAHA.115.306529.
- 945 [129] A. R. Moore, D. A. Willoughby, *Clin. Exp. Immunol.* 2008, **101** (*3*), 387–389. DOI:
  946 https://doi.org/10.1111/j.1365-2249.1995.tb03123.x.
- 947 [130] H. Rehmann, J. De Rooij, J. L. Bos, in *Handb. Cell Signal.*, Elsevier 2010.
- 948 [131] A. E. Christensen, F. Selheim, J. De Rooij, S. Dremier, F. Schwede, K. K. Dao, A.
- 949 Martinez, C. Maenhaut, J. L. Bos, H.-G. Genieser, *et al.*, *J. Biol. Chem.* 2003, 278 (37),
  950 35394–35402. DOI: https://doi.org/10.1074/jbc.M302179200.
- 951 [132] J. M. Enserink, A. E. Christensen, J. De Rooij, M. Van Triest, F. Schwede, H. G.
- 952 Genieser, S. O. Døskeland, J. L. Blank, J. L. Bos, *Nat. Cell Biol.* 2002, 4 (11), 901–
- 953 906. DOI: https://doi.org/10.1038/ncb874.
- 954 [133] M. J. Vliem, B. Ponsioen, F. Schwede, W. Pannekoek, J. Riedl, M. R. H. Kooistra, K.
- Jalink, H. Genieser, J. L. Bos, H. Rehmann, *ChemBioChem*. 2008, 9 (13), 2052–2054.
  DOI: https://doi.org/10.1002/cbic.200800216.
- 957 [134] P. N. Kamzeeva, A. V. Aralov, V. A. Alferova, V. A. Korshun, *Curr. Issues Mol. Biol.*
- 958 2023, **45** (8), 6851–6879. DOI: https://doi.org/10.3390/cimb45080433.
- 959 [135] A. J. Berdis, Front. Mol. Biosci. 2017, 4, 78. DOI:
- 960 https://doi.org/10.3389/fmolb.2017.00078.
- 961 [136] T. M. Dando, G. L. Plosker, *Drugs*. 2003, **63** (20), 2215–2234. DOI:
- 962 https://doi.org/10.2165/00003495-200363200-00007.
- 963 [137] E. D. Clercq, A. Holý, Nat. Rev. Drug Discov. 2005, 4 (11), 928–940. DOI:
- 964 https://doi.org/10.1038/nrd1877.
- 965 [138] E. P. Tchesnokov, A. Obikhod, R. F. Schinazi, M. Gótte, J. Biol. Chem. 2008, 283 (49),
- 966 34218–34228. DOI: https://doi.org/10.1074/jbc.M806797200.

- 967 [139] C. J. Gordon, E. P. Tchesnokov, E. Woolner, J. K. Perry, J. Y. Feng, D. P. Porter, M.
- 968 Götte, J. Biol. Chem. 2020, **295** (20), 6785–6797. DOI:
- 969 https://doi.org/10.1074/jbc.RA120.013679.
- 970 [140] G. Kokic, H. S. Hillen, D. Tegunov, C. Dienemann, F. Seitz, J. Schmitzova, L.
- 971 Farnung, A. Siewert, C. Höbartner, P. Cramer, *Nat. Commun.* 2021, 12 (1), 279. DOI:
  972 https://doi.org/10.1038/s41467-020-20542-0.
- 973 [141] L. Zhang, D. Zhang, X. Wang, C. Yuan, Y. Li, X. Jia, X. Gao, H.-L. Yen, P. P.-H.
- 974 Cheung, X. Huang, *Phys. Chem. Chem. Phys.* 2021, **23** (10), 5852–5863. DOI:
- 975 https://doi.org/10.1039/D0CP05948J.
- 976 [142] P. Huang, A. Sandoval, E. Van Den Neste, M. Keating, W. Plunkett, Leukemia. 2000,
- 977 **14** (8), 1405–1413. DOI: https://doi.org/10.1038/sj.leu.2401845.
- 978 [143] A. Zhenchuk, K. Lotfi, G. Juliusson, F. Albertioni, *Biochem. Pharmacol.* 2009, 78 (11),
  979 1351–1359. DOI: https://doi.org/10.1016/j.bcp.2009.06.094.
- 980 [144] P. L. Bonate, L. Arthaud, W. R. Cantrell, K. Stephenson, J. A. Secrist, S. Weitman, Nat.
- 981 *Rev. Drug Discov.* 2006, **5** (*10*), 855–863. DOI: https://doi.org/10.1038/nrd2055.
- 982 [145] M. Legraverend, *Tetrahedron*. 2008, **64** (*37*), 8585–8603. DOI:
- 983 https://doi.org/10.1016/j.tet.2008.05.115.
- [146] M. Guinan, C. Benckendorff, M. Smith, G. J. Miller, *Molecules*. 2020, 25 (9), 2050.
   DOI: https://doi.org/10.3390/molecules25092050.
- 986 [147] Y. Yoshida, M. Honma, Y. Kimura, H. Abe, *ChemMedChem*. 2021, **16** (5), 743–766.
- 987 DOI: https://doi.org/10.1002/cmdc.202000695.
- 988 [148] K. L. Seley, L. Zhang, A. Hagos, Org. Lett. 2001, 3 (20), 3209–3210. DOI:
- 989 https://doi.org/10.1021/ol0165443.
- 990 [149] K. Seley-Radtke, Antivir. Chem. Chemother. 2018, 26, 204020661875678. DOI:
- 991 https://doi.org/10.1177/2040206618756788.

- 992 [150] K. L. Seley-Radtke, C. H. M. Kutz, J. E. Thames, in *Handb. Chem. Biol. Nucleic Acids*
- 993 (Ed: N. Sugimoto), Springer Nature Singapore, Singapore 2023.
- 994 [151] X. Kong, H. Zuo, H.-D. Huang, Q. Zhang, J. Chen, C. He, Y. Hu, J. Adv. Res. 2023, 44,
- 995 119–133. DOI: https://doi.org/10.1016/j.jare.2022.05.006.
- 996 [152] A. Decout, J. D. Katz, S. Venkatraman, A. Ablasser, Nat. Rev. Immunol. 2021, 21 (9),
- 997 548–569. DOI: https://doi.org/10.1038/s41577-021-00524-z.
- 998 [153] C. Coderch, J. Arranz-Herrero, E. Nistal-Villan, B. De Pascual-Teresa, S. Rius999 Rocabert, *Int. J. Mol. Sci.* 2023, 24 (10), 9032. DOI:
- 1000 https://doi.org/10.3390/ijms24109032.
- 1001 [154] W. Xu, K. M. Chan, E. T. Kool, Nat. Chem. 2017, 9 (11), 1043–1055. DOI:
- 1002 https://doi.org/10.1038/nchem.2859.
- 1003 [155] Y. Saito, R. H. E. Hudson, J. Photochem. Photobiol. C Photochem. Rev. 2018, 36, 48-
- 1004 73. DOI: https://doi.org/10.1016/j.jphotochemrev.2018.07.001.
- 1005 [156] D. Dziuba, P. Didier, S. Ciaco, A. Barth, C. A. M. Seidel, Y. Mély, Chem. Soc. Rev.
- 1006 2021, **50** (*12*), 7062–7107. DOI: https://doi.org/10.1039/D1CS00194A.
- 1007 [157] E. De Clercq, G. Li, *Clin. Microbiol. Rev.* 2016, **29** (*3*), 695–747. DOI:
- 1008 https://doi.org/10.1128/CMR.00102-15.
- 1009 [158] D. Coelho, L. Le Corre, K. Bartosik, L. Iannazzo, E. Braud, M. Ethève-Quelquejeu,
- 1010 *Chem. Eur. J.* 2023, **29** (44), e202301134. DOI:
- 1011 https://doi.org/10.1002/chem.202301134.
- 1012 [159] V. Meynier, L. Iannazzo, M. Catala, S. Oerum, E. Braud, C. Atdjian, P. Barraud, M.
- 1013 Fonvielle, C. Tisné, M. Ethève-Quelquejeu, Nucleic Acids Res. 2022, 50 (10), 5793-
- 1014 5806. DOI: https://doi.org/10.1093/nar/gkac354.
- 1015 [160] T. Okuda, A.-K. Lenz, F. Seitz, J. Vogel, C. Höbartner, Nat. Chem. 2023, 15 (11),
- 1016 1523–1531. DOI: https://doi.org/10.1038/s41557-023-01320-z.

- 1017 [161] A. Bollu, A. Peters, A. Rentmeister, Acc. Chem. Res. 2022, 55 (9), 1249–1261. DOI:
- 1018 https://doi.org/10.1021/acs.accounts.2c00059.
- 1019 [162] R. Micura, C. Höbartner, Chem. Soc. Rev. 2020, 49 (20), 7331–7353. DOI:
- 1020 https://doi.org/10.1039/D0CS00617C.
- 1021
- 1022