



Paenibacillus allorhizoplanea sp. nov. from the rhizoplane of a *Zea mays* root

Peter Kämpfer, André Lipski, Lucie Lamothe, Dominique Clermont, Alexis Criscuolo, John Mcinroy, Stefanie Glaeser

► To cite this version:

Peter Kämpfer, André Lipski, Lucie Lamothe, Dominique Clermont, Alexis Criscuolo, et al.. *Paenibacillus allorhizoplanea* sp. nov. from the rhizoplane of a *Zea mays* root. *Archives of Microbiology*, 2022, 204 (10), pp.630. 10.1007/s00203-022-03225-w . pasteur-03791319

HAL Id: pasteur-03791319

<https://pasteur.hal.science/pasteur-03791319>

Submitted on 29 Sep 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Paenibacillus allorhizoplanae sp. nov. from the rhizoplane of a *Zea mays* root

Peter Kämpfer¹ · André Lipski² · Lucie Lamothe^{3,4} · Dominique Clermont⁵ · Alexis Criscuolo⁴ · John A. McInroy⁶ · Stefanie P. Glaeser¹

Received: 23 August 2022 / Revised: 28 August 2022 / Accepted: 29 August 2022
© The Author(s) 2022

Abstract

A Gram-positive staining, aerobic, endospore-forming bacterial strain, isolated from the rhizosphere of *Zea mays* was studied for its detailed taxonomic allocation. Based on the 16S rRNA gene sequence similarity comparisons, strain JJ-42^T was shown to be a member of the genus *Paenibacillus*, most closely related to the type strain of *Paenibacillus pectinilyticus* (98.8%). The 16S rRNA gene sequence similarity to all other *Paenibacillus* species was below 98.5%. The pairwise average nucleotide identity (ANI) and digital DNA–DNA hybridization (dDDH) values of the JJ-42^T genome assembly against publicly available *Paenibacillus* type strain genomes were below 92% and 47%, respectively. The quinone system of strain JJ-42^T consisted exclusively of menaquinone MK-7. The polar lipid profile consisted of the major components diphosphatidylglycerol, phosphatidylethanolamine, phosphatidylglycerol, three aminophospholipids (APL), and one unidentified lipid. The major fatty acids were iso- and anteiso-branched with the major compound anteiso C_{15:0}. Physiological and biochemical characteristics allowed a further phenotypic differentiation of strain JJ-42^T from the most closely related species. Thus, JJ-42^T represents a novel species of the genus *Paenibacillus*, for which the name *Paenibacillus allorhizoplanae* sp. nov. is proposed, with JJ-42^T (=LMG 32089^T=CCM 9085^T=DSM 111786^T=CIP 111891^T) as the type strain.

Keywords *Paenibacillus allorhizoplanae* · Taxonomy · *Zea mays* · Endophyte · Plant associated · Genome sequence

Introduction

The genus *Paenibacillus*, initially proposed by Ash et al. (1993) is now accommodating more than 280 species (<https://lpsn.dsmz.de/genus/paenibacillus>) isolated from many different sources. It is obvious that species of *Paenibacillus* have been isolated often as endophytes (Carro et al. 2013; Lai et al. 2015; Kittiwongwattana and Thawai 2015; Gao et al. 2015) and from other plant-associated environments as the rhizosphere (Elo et al. 2001; Ma et al. 2007; Kim et al. 2009a, b; Hong et al. 2009; Beneduzi et al. 2010; Zhang et al. 2013; 2015; Wang et al. 2014; Son et al. 2014; Han et al. 2015; Kämpfer et al. 2016, 2017a,b, 2021), seeds (Liu et al. 2015) or the phyllosphere (Rivas et al. 2005, 2006). In general, endospore-forming bacilli, including members of the genus *Paenibacillus*, are of particular interest for their capacity to promote plant growth and form endospores (Grady et al. 2016). The purpose of this study was to analyze strain JJ-42^T in detail for its taxonomic allocation and for the presence of genes associated with plant–growth promotion.

Communicated by Erko Stackebrandt.

✉ Peter Kämpfer
peter.kaempfer@umwelt.uni-giessen.de

- ¹ Institut für angewandte Mikrobiologie, Justus-Liebig-Universität Giessen, Heinrich-Buff-Ring 26–32, 35392 Giessen, Germany
- ² Institut für Ernährungs- und Lebensmittelwissenschaften, Lebensmittelmikrobiologie und –hygiene, Rheinische Friedrich-Wilhelms-Universität Bonn, Bonn, Germany
- ³ CNRS, Institut Français de Bioinformatique, IFB-Core, UMS 3601, Evry, France
- ⁴ Institut Pasteur, Université de Paris, Bioinformatics and Biostatistics Hub, 75015 Paris, France
- ⁵ Institut Pasteur, Université de Paris, CIP–Collection of Institut Pasteur, 75015 Paris, France
- ⁶ Department of Entomology and Plant Pathology, Auburn University, Alabama, USA

Materials and methods

Isolation and culture conditions

Field-grown maize plants (*Zea mays*) grown in Dunbar, Nebraska were manually uprooted a few weeks after planting. Roots, approximately 15 cm in length, were separated from the surrounding soil by vigorous shaking such that only the most tightly adhering soil remained. Bacteria were collected from the root surface by immersing the root in sterile water followed by plating dilutions on Nutrient Agar (Sigma-Aldrich). Strain JJ-42^T was initially isolated at Auburn University in this manner. Subsequent cultivation of JJ-42^T was performed on tryptone soy agar, TSA (Oxoid) at 28 °C for 24 h.

Molecular characterization

16S rRNA gene phylogeny

For a first phylogenetic placement the 16S rRNA gene of strain JJ-42^T was PCR amplified with primer system Eub9f (5'-GAGTTTGATCMTGGCTCAG-3') and Eub1492R (5'-ACGGYTACCTTGTACGACTT-3') (Lane 1991) and sequenced with the Sanger dideoxy sequencing technology using primers Eub9f and E786F (5'-GATTAGATACCCTGGTAG-3'), respectively, as described previously (Schauss et al. 2015). The sequence was manually processed and corrected based on the electropherograms using MEGA11 (Tamura et al. 2021).

A first phylogenetic assignment was performed by BLAST analysis against the EzBioCloud 16S rRNA gene sequence database, including all type strain 16S rRNA gene sequences and BLASTN BLAST + 2.13.0 (reduced to sequences of type material) of the NCBI (<https://blast.ncbi.nlm.nih.gov/>). The 16S rRNA gene sequences of strain JJ-42^T and nonincluded-related type strains were subsequently added to LTP_2020 database of the "All-Species Living Tree" Project (LTP; Yarza et al. 2008) using ARB release 5.2 (Ludwig et al. 2004). The updated database version LTP_04_2021 (released in September 2021) was used. The imported 16S rRNA gene sequences were aligned in the alignment explorer using the pt server generated for the respective database. The alignment was checked manually before subsequent analyses. The sequences were added to the database tree tree_Ltp_all_04_21 containing all type strain 16S rRNA gene sequences of species described until May 2021 using the parsimony quick ad marked tool of ARB and the gap95_q0_to_q5 filter.

All strains of the resulting cluster and the type strain of the type species *Paenibacillus polymyxa* and of two additional species, *Paenibacillus peoriae* and *Paenibacillus*

kribbensis, which clustered with the type species, were included in the analysis. Type strains of two *Cohnella* species, *Cohnella thermotolerans* and *Cohnella xylanilytica*, were used as outgroup. The genus *Cohnella* is a monophyletic genus related to the paraphyletic genus *Paenibacillus*. A maximum-likelihood tree was calculated with RAXML v7.04 (Stamatakis 2006), GTR-GAMMA and rapid bootstrap analysis and a maximum-parsimony tree with DNAPARS v 3.6 (Felsenstein 2005). Both trees were calculated with 100 re-samplings (bootstrap analysis; Felsenstein 1985) and based on the 16S rRNA gene sequences between gene termini 95 to 1475 (*Escherichia coli* numbering, Brosius et al. 1978).

Genomic features

The whole-genome sequencing of strains JJ-42^T was carried out with a NextSeq500 instrument using the Nextera XT DNA library preparation kit (Illumina) and a 2 × 150 bp paired-end protocol, yielding 3,876,943 read pairs (140 × average sequencing depth, 356 bp average insert size). Read processing, genome assembly and scaffolding, and gene prediction and annotation were performed using fq2dna v21.06 (<https://gitlab.pasteur.fr/GIPhy/fq2dna>).

Different plant–beneficial function contributing (PBFC) genes (as listed in Cherif-Silini et al. 2019) were searched using tblastn against the draft genome of JJ-42^T, assessing the presence of different putative PBFC genes related to different features.

Pairwise average nucleotide and amino acid identity (ANI and AAI, respectively) values were computed using OGRI_B (<https://gitlab.pasteur.fr/GIPhy/OGRI>) between the draft genome of JJ-42^T and every *Paenibacillus* type strain genome selected for the 16S rRNA gene sequence based phylogenetic analysis (when publicly available).

Physiology and chemotaxonomy

Detailed phenotypic characterization of strain JJ-42^T was performed in comparison to *P. pectinilyticus* KCTC 13222^T. Cell morphology and motility were observed under a Zeiss light microscope at a magnification of 1000, using cells that had been grown for 3 days at 28 °C on TSA (Oxoid). Gram staining was performed by the modified Hucker method according to Gerhardt et al. (1994). Oxidase activity was tested with an oxidase reagent test kit following the instructions of the manufacturer (bio-Mérieux, France). The KOH test for the determination of spores was carried out according to Moaledj (1986). The growth was tested on different agar media, including R2A (Oxoid), nutrient broth agar (NB, Oxoid), tryptic soy agar (TSA, Becton Dickinson), malt agar (Merck), PYE [0.3% (w/v) yeast extract, and 0.3% (w/v) casein

peptone, respectively, 15 g agar L⁻¹, pH 7.2], CASO agar (Carl Roth), K7 [0.1% (w/v) of yeast extract, peptone, and glucose, 15 g L⁻¹ agar, pH 6.8], medium 65 (M65, according to DSMZ), DEV agar (DEV, Merck), Nutrient agar (NA, Becton Dickinson), Luria–Bertani agar (LB, Sigma-Aldrich), Marine agar 2216 (MA, Becton Dickinson), Columbia agar with sheep blood (Oxoid), and MacConkey agar (Oxoid). The growth was evaluated after 48 h incubation at 28 °C. Temperature-dependent growth was determined on Columbia agar with sheep blood at 4, 10, 15, 20, 25, 28, 30, 36, 45, 50, and 55 °C. pH and salinity-dependent growth was tested in R2A broth incubated at 28 °C. The pH was adjusted to pH 4.5 to 10.5 (1 pH unit intervals incrementing) using HCl and NaOH. For salinity-dependent growth, 1 to 8% (w/v) NaCl was added (in 1% intervals incrementing). The growth was monitored after 72 h of incubation.

Further physiological characterization of the strains was performed with the API20NE and APIZYM test systems according to the instruction of the manufacturer (bioMérieux) and with the methods described by Kämpfer et al. (1991) and Kämpfer (1990). All tests were incubated at 28 °C.

For analyses of quinones and polar lipids cells were grown in half-concentrated nutrient broth at 25 °C for 7 days. Quinones were extracted as described by Minnikin et al. (1984) and by Wiertz et al. (2013), and analyzed with an Agilent 1260 infinity HPLC system. Polar lipids were extracted and analyzed by thin-layer chromatography (TLC) according to Minnikin et al. (1984). Molybdophosphoric acid was used for visualization of all polar lipids. Aminolipids were stained by ninhydrin, molybdenum blue reagent for the detection of phospholipids, and α -naphthol reagent for the detection of sugar-containing lipids.

Fatty acid analysis was performed in parallel with JJ-42^T and *P. pectinilyticus* KCTC 13222^T. The fatty acids were extracted and analyzed as described by Kämpfer & Kroppenstedt (1996). Strains were grown under identical conditions (TS-medium after 72 h incubation at 28 °C) and the cells for extractions were taken from colonies of the same size. Fatty acids were identified with the Sherlock version 2.11, TSBA40 Rev. 4.1.

Results and discussion

Molecular and genome characteristics

The final corrected 16S rRNA gene sequence of strain JJ-42^T (OP023150) had a size of 1,466 nt and spanned gene termini 8 to 1,475 (numbering according to the *Escherichia coli* *rrnB* sequence published by Brosius et al. 1978).

Strain JJ-42^T was placed within the cluster of *Paenibacillus* type strains by BLAST analyses. Strain JJ-42^T showed the highest 16S rRNA gene sequence similarity to the type strain of *Paenibacillus pectinilyticus* (98.8%), followed by the type strains of *Paenibacillus qinlingensis* and *Paenibacillus oryzae* (both 98.4%). Sequence similarities to all other type strains were below 98%. Independent of the applied treeing method, strain JJ-42^T formed a stable cluster (100% bootstrap support) with the type strains of *P. pectinilyticus*, *P. qinlingensis*, *P. oryzae*, and *P. plantarum* (Fig. 1).

The resulting draft genome was made of 8,162,391 bps on 78 contigs (N50, 232,911) with G + C content of 45.07 mol%. A total of 7,004 coding sequences and 103 tRNA was inferred. Genome sequence authenticity was assessed by aligning the 16S rRNA segment derived from Sanger sequencing (OP023150) against the de novo assembly (CAKMMW000000000) using blastn, leading to > 99.4% pairwise sequence similarity.

ANI and AAI values are reported in Table S1, together with the associated digital DNA–DNA hybridization (dDDH) values (formula 2; <https://ggdc.dsmz.de/>). All these estimated pairwise similarity values are far below the commonly admitted species delineation cutoffs (ANI, 95%; AAI, 95%; dDDH, 70%). A phylogenomic classification of these genome sequences was also inferred using JolyTree v2.0 (Criscuolo 2019, 2020), such a tree confirming that strain JJ-42^T formed a stable cluster with the type strains of *P. plantarum*, *P. oryzae*, and *P. pectinilyticus* (Fig. 2).

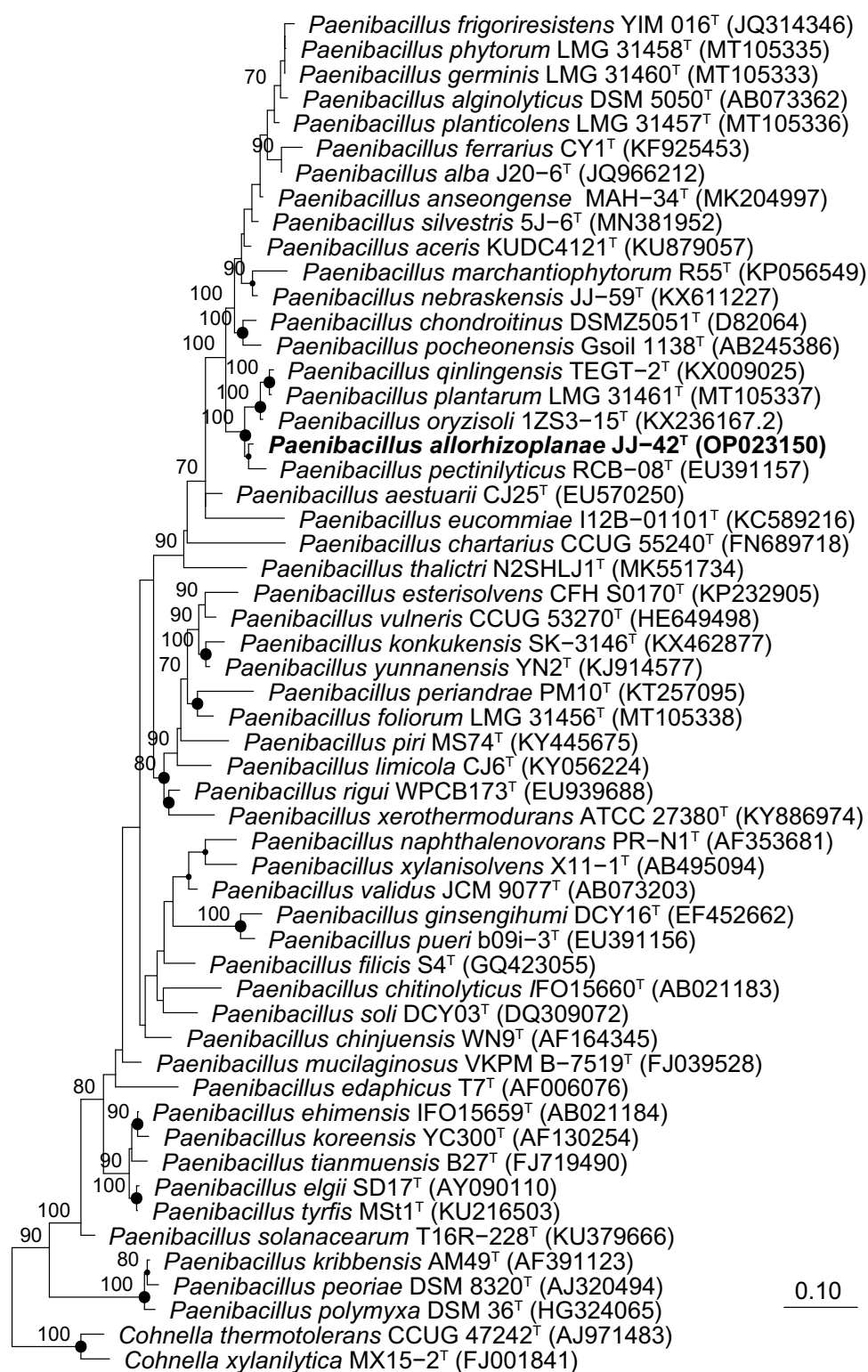
Different plant–beneficial function contributing (PBFC) genes (as listed in Cherif-Silini et al. 2019) were searched using tblastn against the draft genome of JJ-42^T, assessing the presence of different putative PBFC genes related to plant–root colonization (Table S2), nutrient acquisition (Table S3), growth-promoting traits (Table S4), oxidative stress protection (Table S5), drug and heavy metal resistance (Table S6), disease resistance (Table S7), and degradation of aromatic compounds (Table S8). Finally, seven biosynthetic gene clusters for secondary metabolites were found using antiSMASH (Blin et al. 2021) bacterial version (Table S9).

Phenotype

The results of the physiological characterization, performed using methods described previously (Kämpfer 1990; Kämpfer et al. 1991), are given in Table 1 and in the species description.

The polar lipid profile consisted of diphosphatidylglycerol, phosphatidylethanolamine, phosphatidylglycerol, three unidentified aminophospholipids, and one unidentified polar lipid (Fig. S1). The quinone system of strain JJ-42^T

Fig. 1 Maximum-likelihood tree showing the phylogenetic position of strain JJ-42^T among the closest related *Paenibacillus* species. The tree was generated in ARB using RAxML (GTR-GAMMA, rapid bootstrap analysis) and based on the 16S rRNA gene sequences between positions 95–1475 according to *E. coli* numbering (Brosius et al. 1978). GenBank/EMBL/DDBJ accession numbers are given in parentheses. Numbers at branch nodes refer to bootstrap values > 70% (100 replicates). Circle marks nodes that were also present in the maximum-parsimony tree. Larger circles were supported by high bootstrap values in the maximum-parsimony tree. Type strains of *Cohnella* species were used as outgroup. Bar, 0.1 substitutions per nucleotide position



consisted exclusively of menaquinone MK-7. The presence of menaquinone MK-7 as well as the polar lipid profile are in agreement with the characteristics of other species of the genus *Paenibacillus*.

The fatty acids comprised mainly iso- and anteiso-branched fatty acids and the fatty acid profile was very similar to the most closely related *Paenibacillus* species. The detailed fatty acid profile obtained from cells grown is shown in Table S10.

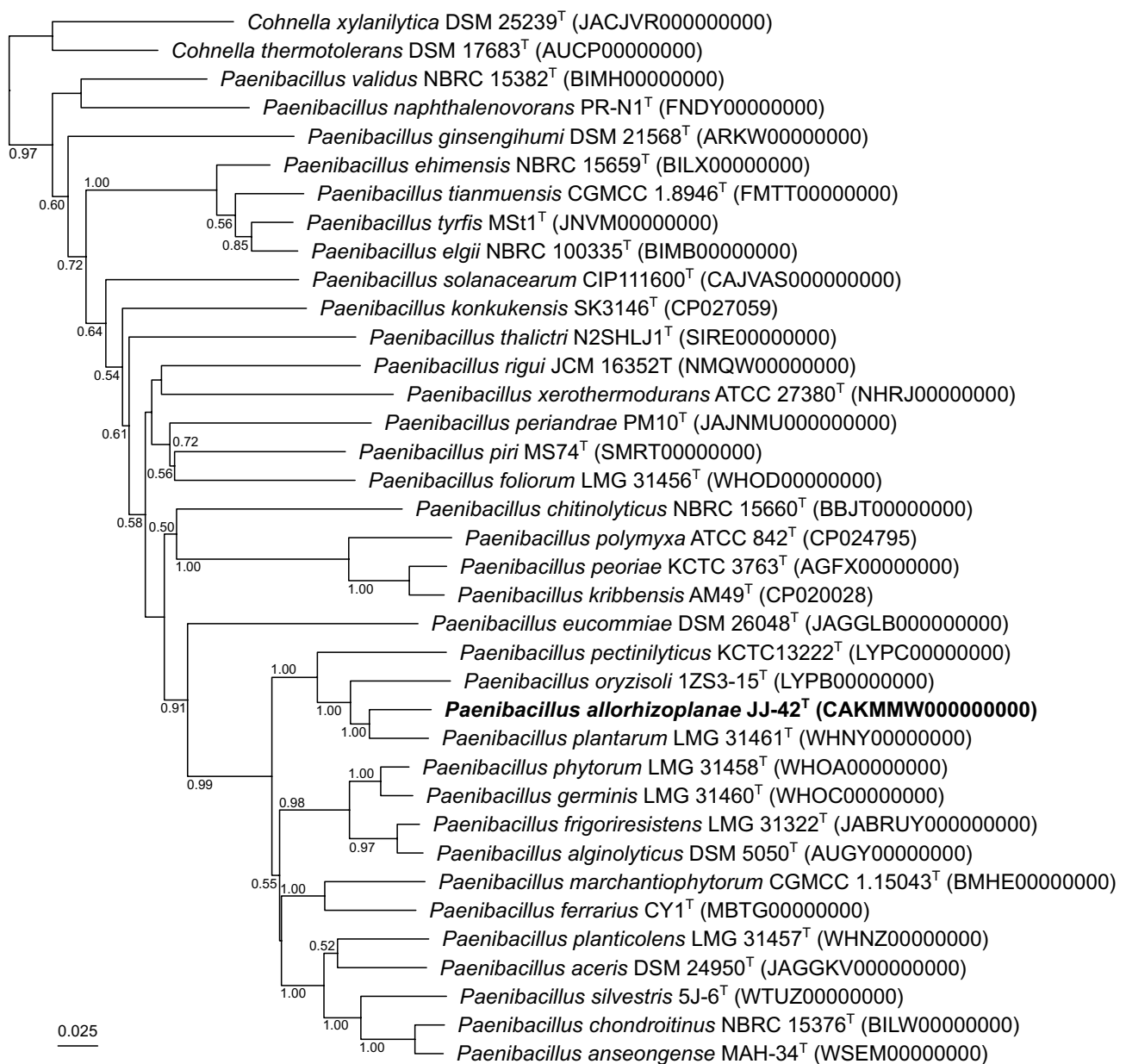


Fig. 2 Whole-genome-based tree showing the phylogenetic placement of strain JJ-42^T among type strains of closely related *Paenibacillus* species. This minimum evolution tree was inferred using JolyTree (<https://gitlab.pasteur.fr/GIPhy/JolyTree>). Two publicly available *Cohnella* type strain genomes were used as an outgroup.

The genome sequence accession is specified between parentheses next after each taxon name. Branch supports were assessed by the rate of elementary quartets, as estimated by JolyTree (only supports > 0.5 were specified). Bar, 0.025 nucleotide substitutions per site

Conclusion

Based on the summary of genotypic and phenotypic, we describe a novel species of the genus *Paenibacillus*, for which the name *Paenibacillus allorhizoplanae* is proposed with JJ-42^T as proposed type strain.

Description of *Paenibacillus allorhizoplanae* sp. nov.

Paenibacillus allorhizoplanae (Gr. masc. adj. *allos*, other; N.L. gen. n. *rhizoplanae*, a specific epithet ('of the rhizoplane'); N.L. gen. n. *allorhizoplanae*, related, but distinct from *Paenibacillus rhizoplanae*).

Table 1 Differential characteristics of strain JJ-42^T and phylogenetically related species of the genus *Paenibacillus*

Characteristic	1	2	3	4	5	6	7
Source	Rhizoplane of a <i>Zea mays</i> root	Gut of <i>Diestrammena apicalis</i> (Korea)	Root of <i>Sinopodophyl-lum hexandrum</i> (China)	Soil (China)	Soil (China)	Soil (U.S.A)	Soil (U.S.A)
Nitrate reduction	–	+	+	–	–	+	+
H ₂ S production	–	–	–	–	–	–	+
Urease	–	+	–	+	–	–	–
Growth at/in:							
4 °C	–	–	–	+	+	–	–
37 °C	+	+	–	+	+	+	+
1% NaCl	+	–	+	–	+	–	+
Hydrolysis of:							
Aesculin	+	+	–	+	+	+	+
Starch	–	–	–	+	–	–	–
Assimilation of (API 20NE):							
<i>N</i> -Acetylglucosamine	+	–	+	+	–	–	+
L-Arabinose	–	–	–	+	–	–	+
D-Glucose	–	–	+	+	–	–	+
Maltose	+	–	+	+	+	–	+
D-Mannitol	–	–	–	+	–	–	+
D-Mannose	–	–	–	+	–	–	–
Potassium gluconate	+	–	–	+	–	–	+
Enzyme activities (API ZYM)							
Acid phosphatase	–	–	–	+	+	–	+
α -Chymotrypsin	–	–	–	+	–	+	–
Cystine arylamidase	–	+	–	+	w	+	–
α -Galactosidase	–	–	–	+	–	+	–
α Glucosidase	–	–	w	+	+	–	w
β -Glucosidase	+	+	+	+	–	–	–
Trypsin	–	–	–	+	–	–	–
Valine arylamidase	+	–	–	+	+	–	–

Strains: 1, JJ-42^T; 2, *Paenibacillus pectinilyticus* KCTC 13222^T; 3, *Paenibacillus qinlingensis* TEGT-2^T; 4, *Paenibacillus frigorigresistens* CCTCC AB 2011150^T; 5, *Paenibacillus ferrarius* CCTCC AB 2013369^T; 6, *Paenibacillus alginolyticus* NBRC 15375^T; 7, *Paenibacillus chondroitinus* DSM 5051^T. Data for taxa 1 and 2 from this study; all other data are from Xin et al. (2017). +, Positive; –, negative; w, weakly positive

*After 48 h of incubation

Cells (with rounded ends) stain Gram-positive. No chains or filaments could be observed after growth on TSA at 28 °C for 48 h. Cells were 2.0–3.0 µm in length and 0.8–1.0 µm in width) and showed no motility. Oval endospores are formed in a subterminal position in swollen sporangia. No other cell inclusions could be detected. Colonies grown on tryptone soy agar (after 48 h of incubation on TSA) are circular, convex, and beige with a shiny appearance and an average diameter of 2–3 mm. The growth at 28 °C on R2A (Oxoid), NB (Oxoid), TSA, Malt agar (Merck), PYE, CASO agar (Carl Roth), K7, M65, DEV agar (DEV, Merck), NA (Becton Dickinson), LB (Sigma-Aldrich), and Columbia agar with sheep blood

(Oxoid). No growth on MA (Becton Dickinson) and MacConkey agar (Oxoid).

Good growth on blood agar between 25 and 45 °C (optimal growth at 36 °C), growth occurred between 15 and 45 °C, but not at 4, 10, 50 °C, and above. Optimal pH for growth in R2A broth at 28 °C is pH 7–8; growth occurs between pH 4.5 and 10.5. Optimal growth in R2A broth at 28 °C without and in the presence of 1% NaCl, growth occurs between 0 and 3% NaCl, but not at 4% or above. Both, pH- and salinity-dependent growth tested in R2A broth at 28 °C. Tests for catalase and oxidase activities are negative. According to API 20 NE, positive for the activity of β -glucosidase (esculin hydrolysis), PNPG

beta-galactosidase, and the assimilation of potassium gluconate, and negative for the activity of nitrate reduction, indol production, D-glucose fermentation, arginine dihydrolase, urease, gelatin hydrolysis, and the assimilation of D-glucose, L-arabinose, D-mannose, D-mannitol, N-acetylglucosamine, D-maltose, capric acid, adipic acid, malic acid, trisodium citrate, and phenylacetic acid.

According to API ZYM, positive for leucine arylamidase, acid phosphatase, naphthol-AS-BI-phosphohydrolase, and β -galactosidase, and negative for alkaline phosphatase, esterase (C4), esterase lipase (C8), lipase (C14), valine arylamidase, cystine arylamidase, trypsin, α -chymotrypsin, α -galactosidase, β -glucuronidase, α -glucosidase, β -glucosidase, N-acetyl- β -glucosaminidase, α -mannosidase, and α -fucosidase.

Some sugars or sugar-related compounds were utilized: L-arabinose (weak), D-galactose, D-gluconate, glucose (weak), D-melibiose, ribose, D-trehalose, and D-maltitol are utilized as sole sources of carbon.

Arbutin, D-cellobiose, D-fructose, i-inositol, D-mannose, D-maltose, salicin, sucrose, D-sorbitol, D-xylose, acetate, N-acetyl-D-glucosamine, cis-aconitate, trans-aconitate, adipate, D-adonitol, 4-aminobutyrate, azelate, citrate, itaconate, malate, mesaconate, 2-oxoglutarate, propionate, putrescine, pyruvate and L-rhamnose are not utilized as sole carbon source.

The quinone system contains only menaquinone MK-7. The polar lipid profile is composed of the major lipids diphenylglycerol, phosphatidylglycerol, phosphatidylethanolamine, three unidentified aminophospholipids (APL1-3) and one unidentified polar lipid (L).

The major fatty acids are anteiso $C_{15:0}$ and iso $C_{16:0}$. The genomic DNA G + C content is 45.07 mol% (based on the genome sequence).

The type strain JJ-42^T (= LMG 32089^T = CCM 9085^T = DSM 111786^T = CIP 111891^T) was isolated from the root surface of a field-grown corn plant in Dunbar, Nebraska USA.

The genome sequence of the type strain is available under accession number CAKMMW000000000 and the 16S rRNA gene sequence under OP023150.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00203-022-03225-w>.

Acknowledgements We thank Gundula Will, Maria Sowinsky and Katja Grebing for excellent technical assistance and Prof. Aharon Oren for his help with the etymology of the specific epithet. We also acknowledge the help of the HPC Core Facility of the Institut Pasteur for this work.

Author contributions P.K. conceived and designed the study and provided fatty acid data. P.K. S.G. A.C. J.M. A.L. and D.C. wrote the manuscript. S.G. A.L. and P.K. conducted the experiments. A.L. analyzed

the lipids and quinones. A.C. and L. L. performed the genomic and phylogenomic analyses.

Funding Open Access funding enabled and organized by Projekt DEAL. No funding was received.

Availability of data and material The new generated sequences were uploaded to the GenBank database at the National Center for Biotechnology Information (NCBI) and are available. The complete genome sequence of strain JJ-42^T has been deposited under the GenBank/EMBL/DBJ accession numbers CAKMMW000000000 and the 16S rRNA gene sequence under OP023150.

Declarations

Conflict of interest The authors have no relevant financial or non-financial interests to disclose. The authors declare no competing interests.

Ethical approval This article does not contain any studies with human participants or animals performed by any of the authors.

Consent for publication The manuscript is submitted with the consent of all authors.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Ash C, Priest FG, Collins MD (1993) Molecular identification of rRNA group 3 bacilli (Ash, Farrow, Wallbanks and Collins) using a PCR probe test. *Antonie Van Leeuwenhoek* 64:253–260. <https://doi.org/10.1007/bf00873085>
- Beneduzi A, Costa PB, Parma M et al (2010) *Paenibacillus riograndensis* sp. nov., a nitrogen-fixing species isolated from the rhizosphere of *Triticum aestivum*. *Int J Syst Evol Microbiol* 60:128–133. <https://doi.org/10.1099/ijs.0.011973-0>
- Blin K, Shaw S, Kloosterman AM et al (2021) antiSMASH 6.0: improving cluster detection and comparison capabilities. *Nucleic Acids Res* 49(W1,2):W29–W35. <https://doi.org/10.1093/nar/gkab335>
- Brosius J, Palmer ML, Kennedy PJ et al (1978) Complete nucleotide sequence of a 16S ribosomal-RNA gene from *Escherichia coli*. *PNAS* 75:4801–4805. <https://doi.org/10.1073/pnas.75.10.4801>
- Carro L, Flores-Félix JD, Cerda-Castillo E et al (2013) *Paenibacillus endophyticus* sp. nov., isolated from nodules of *Cicer arietinum*. *Int J Syst Evol Microbiol* 63:4433–4438. <https://doi.org/10.1099/ijs.0.050310-0>
- Cherif-Silini H, Thissera B, Chenari Bouket A et al (2019) Durum wheat stress tolerance induced by endophyte *Pantoea agglomerans*

- with genes contributing to plant functions and secondary metabolite arsenal. *Int J Mol Sci* 20:3989. <https://doi.org/10.3390/ijms20163989>
- Crisuolo A (2019) A fast alignment-free bioinformatics procedure to infer accurate distance-based phylogenetic trees from genome assemblies. *Res Ideas Outcomes* 5:e36178. <https://doi.org/10.3897/rio.5.e36178>
- Crisuolo A (2020) On the transformation of MinHash-based uncorrected distances into proper evolutionary distances for phylogenetic inference. *F1000Research* 9:1309. <https://doi.org/10.12688/f1000research.26930.1>
- Elo S, Suominen I, Kämpfer P et al (2001) *Paenibacillus borealis* sp. nov., a nitrogen-fixing species isolated from spruce forest humus in Finland. *Int J Syst Evol Microbiol* 51:535–545
- Felsenstein J (1985) Confidence limits of phylogenies: an approach using the bootstrap. *Evolution* 39:783–791. <https://doi.org/10.1111/j.1558-5646.1985.tb00420.x>
- Felsenstein J (2005) PHYLIP (Phylogeny Inference Package) version 3.6. Distributed by the author. Department of Genome Sciences, University of Washington, Seattle
- Gao JL, Lv FY, Wang XM et al (2015) *Paenibacillus wenxiniae* sp. nov., a nifH gene-harboring endophytic bacterium isolated from maize. *Antonie Van Leeuwenhoek* 108:1015–1022. <https://doi.org/10.1007/s10482-015-0554-8>
- Gerhardt P, Murray RGE, Wood W et al (1994) *Methods for General and Molecular Bacteriology*. American Society for Microbiology, Washington
- Grady EN, MacDonald J, Liu L et al (2016) Current knowledge and perspectives of *Paenibacillus*: a review. *Microb Cell Fact* 15:203. <https://doi.org/10.1186/s12934-016-0603-7>
- Han T-Y, Tong X-M, Wang Y-W et al (2015) *Paenibacillus populi* sp. nov., a novel bacterium isolated from the rhizosphere of *Populus alba*. *Antonie Van Leeuwenhoek* 108:659–666. <https://doi.org/10.1007/s10482-015-0521-4>
- Hong YY, Ma YC, Zhou YG et al (2009) *Paenibacillus sonchi* sp. nov., a nitrogen-fixing species isolated from the rhizosphere of *Sonchus oleraceus*. *Int J Syst Evol Microbiol* 59:2656–2661. <https://doi.org/10.1099/ijms.0.009308-0>
- Kämpfer P (1990) Evaluation of the Titertek-Enterobac-Automated system (TTE-AS) for identification of *Enterobacteriaceae*. *Zentralbakteriol* 273:164–172. [https://doi.org/10.1016/s0934-8840\(11\)80244-6](https://doi.org/10.1016/s0934-8840(11)80244-6)
- Kämpfer P, Kroppenstedt RM (1996) Numerical analysis of fatty acid patterns of coryneform bacteria and related taxa. *Can J Microbiol* 42:989–1005. <https://doi.org/10.1139/m96-128>
- Kämpfer P, Steiof M, Dott W (1991) Microbiological characterisation of a fuel-oil contaminated site including numerical identification of heterotrophic water and soil bacteria. *Microbiol Ecol* 21:227–243. <https://doi.org/10.1007/bf02539156>
- Kämpfer P, Busse HJ, Kloepper JW et al (2016) *Paenibacillus cucumis* sp. nov., isolated from a cucumber plant. *Int J Syst Evol Microbiol* 66:2599–2603
- Kämpfer P, Busse HJ, McInroy JA et al (2017a) *Paenibacillus nebraskensis* sp. nov., isolated from the root surface of field-grown maize. *Int J Syst Evol Microbiol* 67:4956–4961. <https://doi.org/10.1099/ijsem.0.002357>
- Kämpfer P, Busse HJ, McInroy JA et al (2017b) *Paenibacillus rhizoplanae* sp. nov., isolated from the rhizosphere of *Zea mays*. *Int J Syst Evol Microbiol* 67:1058–1063. <https://doi.org/10.1099/ijsem.0.001779>
- Kämpfer P, Busse HJ, McInroy JA et al (2021) *Paenibacillus allorhizosphaerae* sp. Nov., from soil of the rhizosphere of *Zea mays*. *Int J Syst Evol Microbiol*. <https://doi.org/10.1099/ijsem.0.005051>
- Kim BC, Lee KH, Kim MN et al (2009a) *Paenibacillus pini* sp. nov., a cellulolytic bacterium isolated from the rhizosphere of pine tree. *J Microbiol* 47:699–704. <https://doi.org/10.1007/s12275-009-0343-z>
- Kim BC, Lee KH, Kim MN et al (2009b) *Paenibacillus pinihumi* sp. nov., a cellulolytic bacterium isolated from the rhizosphere of *Pinus densiflora*. *J Microbiol* 47:530–535. <https://doi.org/10.1007/s12275-009-0270-z>
- Kittiwongwattana C, Thawai C (2015) *Paenibacillus lemnae* sp. nov., an endophytic bacterium of duckweed (*Lemna aequinoctialis*). *Int J Syst Evol Microbiol* 65:107–112. <https://doi.org/10.1099/ijms.0.067876-0>
- Lai WA, Hameed A, Lin SY et al (2015) *Paenibacillus medicaginis* sp. nov. a chitinolytic endophyte isolated from the root nodule of alfalfa (*Medicago sativa* L.). *Int J Syst Evol Microbiol* 65:3853–3860. <https://doi.org/10.1099/ijsem.0.000505>
- Lane DJ (1991) 16S/23S rRNA sequencing. In: Stackebrandt E, Goodfellow M (eds) *Nucleic acid techniques in bacterial systematics*. Wiley, Chichester, United Kingdom, pp 115–175
- Liu Y, Zhai L, Wang R et al (2015) *Paenibacillus zae* sp. nov., isolated from maize (*Zea mays* L.) seeds. *Int J Syst Evol Microbiol* 65:4533–4538. <https://doi.org/10.1099/ijsem.0.000608>
- Ludwig W, Strunk O, Westram R et al (2004) ARB: a software environment for sequence data. *Nucleic Acid Res* 32:1363–1371. <https://doi.org/10.1093/nar/gkh293>
- Ma Y, Xia Z, Liu X et al (2007) *Paenibacillus sabinae* sp. nov., a nitrogen-fixing species isolated from the rhizosphere soils of shrubs. *Int J Syst Evol Microbiol* 57:6–11. <https://doi.org/10.1099/ijms.0.64519-0>
- Minnikin DE, O'Donnell AG, Goodfellow M et al (1984) An integrated procedure for the extraction of bacterial isoprenoid quinones and polar lipids. *J Microbiol Method* 2:233–241. [https://doi.org/10.1016/0167-7012\(84\)90018-6](https://doi.org/10.1016/0167-7012(84)90018-6)
- Moaledj K (1986) Comparison of Gram-staining and alternate methods, KOH test and aminopeptidase activity in aquatic bacteria: their application to numerical taxonomy. *J Microbiol Methods* 5:303–310. [https://doi.org/10.1016/0167-7012\(86\)90056-4](https://doi.org/10.1016/0167-7012(86)90056-4)
- Rivas R, Mateos PF, Martínez-Molina E et al (2005) *Paenibacillus phyllosphaerae* sp. nov., a xylanolytic bacterium isolated from the phyllosphere of *Phoenix dactylifera*. *Int J Syst Evol Microbiol* 55:743–746. <https://doi.org/10.1099/ijms.0.63323-0>
- Rivas R, García-Fraile P, Mateos PF et al (2006) *Paenibacillus cellululosilyticus* sp. nov., a cellulolytic and xylanolytic bacterium isolated from the bract phyllosphere of *Phoenix dactylifera*. *Int J Syst Evol Microbiol* 56:2777–2781. <https://doi.org/10.1099/ijms.0.64480-0>
- Schauss T, Busse HJ, Golke J et al (2015) *Empedobacter stercoris* sp. nov., isolated from an input sample of a biogas plant. *Int J Syst Evol Microbiol* 65:3746–3753. <https://doi.org/10.1099/ijsem.0.000486>
- Son J-S, Kang H-U, Ghim S-Y (2014) *Paenibacillus dongdonensis* sp. nov., isolated from rhizospheric soil of *Elymus tsukushiensis*. *Int J Syst Evol Microbiol* 64:2865–2870. <https://doi.org/10.1099/ijms.0.061077-0>
- Stamatakis A (2006) RAxML-VI-HP: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22:2688–2690. <https://doi.org/10.1093/bioinformatics/btl446>
- Tamura K, Stecher G, Kumar S (2021) MEGA11: molecular evolutionary genetics analysis Version 11. *Mol Biol Evol* 38(7):3022–3027. <https://doi.org/10.1093/molbev/msab120>
- Wang DS, Jiang YY, Wie XM et al (2014) *Paenibacillus quercus* sp. nov., isolated from rhizosphere of *Quercus aliena* var. *acuteserrata*. *Antonie Van Leeuwenhoek* 105:1173–1178. <https://doi.org/10.1007/s10482-014-0178-4>
- Wiertz R, Schulz SC, Müller U et al (2013) *Corynebacterium frankenforstense* sp. nov. and *Corynebacterium lactis* sp. nov.,

- isolated from raw cow milk. *Int J Syst Evol Microbiol* 63:4495–4501. <https://doi.org/10.1099/ijms.0.050757-0>
- Xin K, Li M, Chen C et al (2017) *Paenibacillus qinlingensis* sp. nov., an indole-3-acetic acid-producing bacterium isolated from roots of *Sinopodophyllum hexandrum* (Royle) Ying. *Int J Syst Evol Microbiol* 67:589–595. <https://doi.org/10.1099/ijsem.0.001666>
- Yarza P, Richter M, Peplies J et al (2008) The all-species living tree project: A 16S rRNA-based phylogenetic tree of all sequenced type strains. *Syst Appl Microbiol* 31:241–250. <https://doi.org/10.1016/j.syapm.2008.07.001>
- Zhang J, Wang ZT, Yu HM et al (2013) *Paenibacillus catalpae* sp. nov., isolated from the rhizosphere soil of *Catalpa speciosa*. *Int J Syst Evol Microbiol* 63:1776–1781. <https://doi.org/10.1099/ijms.0.040659-0>
- Zhang L, Gao JS, Zhang S et al (2015) *Paenibacillus rhizoryzae* sp. nov., isolated from rice rhizosphere. *Int J Syst Evol Microbiol* 65:3053–3059. <https://doi.org/10.1099/ijms.0.000376>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.