

The healthy human virome: from virus–host symbiosis to disease

Eugene V Koonin, Valerian Dolja, Mart Krupovic

▶ To cite this version:

Eugene V Koonin, Valerian Dolja, Mart Krupovic. The healthy human virome: from virus-host symbiosis to disease. Current Opinion in Virology, 2021, This review comes from a themed issue on The virome in health and disease, 47, pp.86-94. 10.1016/j.coviro.2021.02.002. pasteur-03165430

HAL Id: pasteur-03165430 https://pasteur.hal.science/pasteur-03165430

Submitted on 10 Mar 2021 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

1	The healthy human virome: from virus-host symbiosis to disease
2	
3	Eugene V. Koonin, ^{1,#} Valerian V. Dolja, ² Mart Krupovic ³
4	
5	¹ National Center for Biotechnology Information, National Library of Medicine, National Institutes
6	of Health, Bethesda, Maryland, USA
7	² Department of Botany and Plant Pathology, Oregon State University, Corvallis, Oregon, USA
8	³ Archaeal Virology Unit, Institut Pasteur, Paris, France
9	
10	[#] For correspondence: <u>koonin@ncbi.nlm.nih.gov</u>
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24 25	
25	
20 27	
28	

29 Abstract

Viruses are ubiquitous, essential components of any ecosystem, and of multicellular organism 30 31 holobionts. Numerous viruses cause acute infection, killing the host or being cleared by immune system. In many other cases, viruses coexist with the host as symbionts, either temporarily or for 32 33 the duration of the host's life. Apparently, virus-host relationships span the entire range from aggressive parasitism to mutualism. Here we attempt to delineate the healthy human virome, that 34 35 is, the entirety of viruses that are present in a healthy human body. The bulk of the healthy virome 36 consists of bacteriophages infecting bacteria in the intestine and other locations. However, a 37 variety of viruses, such as anelloviruses and herpesviruses, and the numerous endogenous retroviruses, persist by replicating in human cells, and these are our primary focus. Crucially, the 38 39 boundary between symbiotic and pathogenic viruses is fluid such that members of the healthy 40 virome can become pathogens under changing conditions. 41

42

43

44 Introduction

45 Billions of years of perennial and ubiquitous co-evolution between viruses and cells have produced 46 a broad spectrum of virus-host interaction regimes, ranging from aggressive antagonism to 47 commensalism, whereby viruses coexist with their hosts without harming them, at least, in the 48 short term, and even to mutualism when a virus is beneficial and can be essential to the host [1-4]. The relationship between a particular virus and its host can be rarely, if ever, defined by a single 49 regime. Rather, the mode of virus-host interaction is a function of multiple factors, including the 50 51 environmental conditions, host and virus population structure, the immunological status of the host 52 and many more.

53

Virus-host relationships that do not result in the demise of the host cell have been described across the virosphere [3,4]. Hence the concept of the 'normal' or 'healthy' virome, in principle, is applicable to any organism, including humans. Indeed, although most of the best characterized human viruses cause acute infections and are associated with diseases, a healthy human organism is host to a much greater variety of viruses [5-7]. The substantial majority of these infect bacteria that inhabit the human intestine, but a number of viruses actually reproduce in human cells without causing disease, at least, in the short term.

61

In this brief review, we systematically discuss the healthy human virome and emphasize that the boundary between "normal" and pathogenic (that is, causally associated with clinically manifested disease) viromes is blurred. Indeed, the same virus can be either a symbiont (with either no perceptible fitness effect on the host, that is, effectively, a commensal, or with a beneficial effect) or a pathogen depending on the conditions such as the health status and developmental stage of the host. We further stress that the current knowledge of symbiotic viruses lags far behind that of the pathogenic viruses.

69

70 The healthy human phageome

The human intestine, by far the richest microbial habitat in the body, contains about 10¹⁴ bacterial and archaeal cells at any given moment [8]. As many microbial communities, the human microbiome hosts a broad variety of viruses, in which tailed bacteriophages (class *Caudoviricetes* within the realm *Duplodnaviria*) comprise the overwhelming majority [8,9], albeit with a considerable contribution by ssDNA phages of the families *Microviridae* [10-12] and *Inoviridae* [13] (realm *Monodnaviria*). Systematic metagenomics surveys of the human phageome identify 77 thousands of phages but show that relatively few are common components of the healthy gut 78 phageome. Thus, one of the most detailed metagenomics analyses resulted in the identification of only 23 phages that were shared by >50% of the tested individuals [9]. Strikingly, the most 79 80 prevalent human-associated phage (and most prominent component of the healthy human virome 81 altogether), the crAssphage, has been discovered only through metagenomics [14]. This initial 82 finding has been followed by the characterization of an expansive group of crAss-like phages, also 83 by metagenome analysis [15,16]. Subsequently, some crAss-like phages have been grown in 84 cultures of the respective host bacteria, members of the phylum Bacteroidetes [17,18]. Given the 85 difficulty of growing the human intestine-associated bacteria-phage systems in the laboratory, the 86 actual size and diversity of the healthy human phageome remains to be discovered. Importantly, 87 substantial changes in the human phageome have been associated with various diseases conditions, 88 including infection with human and simian immunodeficiency viruses, and in some cases, the 89 phageome perturbation was, apparently, decoupled from the changes in the microbiome [19-21].

90

91 Symbiotic viruses replicating in human cells

Over many decades, diverse viruses have been isolated from healthy humans more or less by chance. In the last few years, systematic surveys of the healthy human virome became possible thanks to the advances of metagenomics [6,7]. Below we briefly discuss the viruses that have been shown, either by traditional virology approaches or by metagenomics (Figure 1a), to be commonly present in healthy humans without causing apparent disease, following the main divisions of the current virus taxonomy [22,23]. The key aspects of the association of these viruses with the human organism are summarized in Figure 1.

99

100 Realm Riboviria

101 Kingdom Orthornavira

102 The realm *Riboviria* consists of viruses with RNA genomes as well as viruses with DNA genomes 103 that employ reverse transcription in their replication cycles. The kingdom *Orthornavira* 104 encompasses RNA viruses that share homologous RNA-dependent RNA polymerases. Many RNA 105 viruses have been isolated or detected by metatranscriptomics in healthy humans but few can be 106 confidently identified as symbionts replicating in human cells (Figure 1a).

107

Perhaps, the most notable apparent human symbionts among the orthornaviruses are pegiviruses
(family *Flaviviridae*) [24,25]. The prototypical pegivirus has been initially tentatively identified

as hepatitis G virus [26], but no association with hepatitis has been subsequently confirmed [27]. 110 111 In the family tree of flavivirids, pegiviruses tightly cluster with hepaciviruses which include 112 hepatitis C virus (HCV), a major human pathogen [24,25]. However, unlike HCV, pegiviruses 113 have not been linked to any pathology. Pegivirus infection in humans is common, with the 114 incidence of about 5%, and pegiviruses readily grow in human cell cultures, leaving no doubt that 115 these are bona fide human viruses [25]. Notably, pegivirus infection appears to be associated with 116 benign clinical outcome in AIDS patients [28,29] indicating that these apparent viral symbionts of 117 humans could benefit the host via protection from other viruses.

118

119 A recent, intriguing addition to the human virome are statoviruses (for STool-Associated TOmbus-120 like viruses) that were originally identified in multiple metagenomes from humans, macaques, cows and mice [30], followed by detection in nasal-throat swabs of humans with acute respiratory 121 122 disease [31]. Although the actual hosts of statoviruses remain unknown, RdRP phylogeny, where statovirus RdRPs cluster with a variety of unclassified tombus-like viruses from invertebrate 123 124 holobionts (that is, a host together with the entirety of associated symbionts and parasites) [32], 125 suggests that statoviruses are associated with either mammalian diet (e.g., plants) or, more likely, 126 some protist symbionts or parasites.

127

Another widespread group of putative members of the healthy human virome are members of *Picobirnaviridae*. Picobirnaviruses are commonly detected in mammalian, including human, intestines and have not been convincingly linked to any diseases although are suspected to be associated with diarrhea [33,34]. Picobirnaviruses have never been grown in cell cultures, and their true hosts remain unknown. The presence of highly conserved ribosome-binding sites (Shine-Dalgarno sequences), which are a hallmark of prokaryotic mRNAs, has led to the suggestion that picobirnaviruses infect bacteria in mammalian microbiomes [35].

135

In addition, certain food-derived plant viruses are present in the human gastrointestinal tract and in feces, sometimes in substantial amounts. The list of such viruses is topped by pepper mild mottle virus (PMMoV), a tobamovirus commonly found in pepper, pepper-derived products and their consumers all over the world [36]. Remarkably, PMMoV retains infectivity after passing through the human alimentary tract. Due to its stability and wide presence in human feces, PMMoV is used as a surrogate marker of fecal contamination of water [37]. However, given that there is no evidence of plant virus replication in vertebrate cells, any substantial role of plant viruses in humanhealth is an extremely remote possibility.

144

145 Kingdom *Pararnavira*

146 This virus kingdom includes viruses that employ reverse transcription in their replication cycles. Over 3,000 of human endogenous retroviruses (HERVs) are integrated into the host genome, 147 comprising about 8% of human DNA [38]. Accordingly, the HERVs play important and diverse 148 149 roles in human biology that cannot be discussed in this brief review in any detail (for recent reviews, see [39-42]). Most of the HERVs appear to descend from ancient integration events so 150 151 that the virus genes are disrupted and rearranged. The legacy of these ancient HERVs are genes 152 encoding virus structural proteins (Gag and Env) that have been recruited for a variety of 153 physiological functions [43], the best known being syncitins, the placental trophoblast receptors 154 [39]. However, some members of the youngest group of HERVs, known as HERV-K or HML2, that are thought to have invaded the human genome less than a million years ago form virus 155 particles, particularly, in early embryogenesis [44,45]. The HERV-K viruses, at least, are bona 156 157 fide members of the healthy human virome. Many of the other HERVs are expressed as well and 158 are implicated in a variety of functions including modulation of innate immunity, even though 159 functional virus proteins are usually not produced $[\underline{46}]$. In particular, it has been reported that expression of one of HERV-K suppresses the spread of invasive melanoma [47]. However, 160 potential associations between HERVs and various diseases have been reported as well. Thus, the 161 relationship between the HERVs and the human host is a typical symbiosis, with both beneficial 162 163 and potential deleterious effects on the host (Figure 1b).

164

165 Realm Monodnaviria

Realm Monodnaviria includes prokaryotic and eukaryotic viruses which encode homologous 166 167 replication initiation endonucleases of the HUH superfamily or their inactivated derivatives, as in the case of polyomaviruses and papillomaviruses [22,48]. In addition to phages of the 168 Microviridae and Inoviridae families mentioned above, several other representatives of 169 Monodnaviria have been repeatedly identified as part of the healthy human virome. These include 170 171 members of the families Parvoviridae, Genomoviridae, Smacoviridae, Papillomaviridae and 172 *Polyomaviridae* (Figure 1a). The actual hosts for human-associated genomoviruses [49] and smacoviruses [50] remain unknown but these viruses likely infect human-associated microbes 173 174 rather than humans directly.

175

176 *Parvoviruses*

177 Parvoviruses from several genera have been detected in various samples from apparently healthy 178 humans, including human bocaviruses (HBoV; genus *Bocaparvovirus*), adeno-associated viruses 179 (AAVs; genus Dependoparvovirus), human parvovirus 4 (PARV4; genus Tetraparvovirus), parvovirus B19 (B19V; genus Erythroparvovirus) and several protoparvoviruses (genus 180 *Protoparvovirus*) [51-54]. These viruses display highly variable cell tropism and pathogenicity. 181 182 For instance, AAVs and PARV4 can infect cells from multiple tissue types and are not known to cause any disease. By contrast, HBoV is most commonly found in the respiratory and 183 184 gastrointestinal tracts as well as in blood [55,56], and is associated with acute respiratory 185 symptoms, especially in children [57]. However, the direct role of HBoV as a pathogen remains unclear. B19V invades red blood cell precursors in the bone marrow and is commonly considered 186 187 as a human pathogen causing a wide range of pathological conditions, including the fifth disease 188 in children, persistent anemia in immunocompromised patients, transient aplastic crises, hydrops 189 fetalis in pregnant women, and arthropathy [52]. Yet, in healthy adults, B19V infections are largely 190 asymptomatic [58], with the prevalence of up to 25% in healthy human skin biopsies [59]. Thus, 191 B19V is a conditional component of the "healthy" human virome that can turn into a pathogen in 192 response to various factors (Figure 1b).

193

194 Polyomaviruses and Papillomaviruses

195 Monodnaviria includes class Papovaviricetes that consists of Polyomaviridae and Papillomaviridae, two families of viruses with small (5-8 Kbps), circular dsDNA genomes 196 197 [60,61]. Papovaviruses are thought to have evolved from parvoviruses [48], with polyomaviruses 198 emerging in invertebrates and co-evolving with animals for at least half a billion years [62]. In 199 humans, polyomaviruses lead a low-profile life styles characterized by low propagation levels, 200 evasion of clearance by the immune system and asymptomatic infections in immunocompetent 201 individuals [60]. Apparently, human polyomaviruses have evolved mechanisms to limit their own 202 reproduction levels in order to establish persistent infections [63]. A poster child human 203 polyomavirus is John Cunningham virus (JCV) that establishes life-long latent infections [64]. The 204 seroprevalence of JCV and other human polyomaviruses in adults can reach 90% or higher, with 205 common coinfection by several polyomaviruses that are transmitted through direct contacts between humans or through contaminated objects. Contrary to their name and the oncogenic 206 207 potential of the large T (tumor) antigen (the early virus protein involved in genome replication) in experimental settings, most of the human polyomaviruses are not oncogenic. The Merkel cell
polyomavirus is the only one that has been convincingly identified as the etiological agent of the
eponymous carcinoma, a skin cancer caused by malignant transformation of the skin
neuroendocrine cells apparently facilitated by virus genome integration into the host chromosomes
[60,65].

213

Papillomaviruses are the closest, albeit apparently somewhat 'younger' relatives of 214 215 polyomaviruses that likely emerged in vertebrates ~350 mya, but their life style is distinct. The majority of the ~200 known human papillomaviruses (HPVs) belong to Beta and Gamma types 216 (Betapapillomavirus and Gammapapillomavirus genera, respectively), and cause unapparent 217 productive infections or low-grade disease of the skin or mucosal epithelium (e.g., warts and 218 condylomas) that are normally cleared by the immune system [66,67]. The virus replication cycle 219 is tightly linked to epithelial differentiation and is orchestrated by a regulatory network that 220 221 involves coordination between the cell cycle, virus DNA replication and transcription, and RNA 222 splicing [67,68]. Most of the infections by the 'high-risk', Alpha HPV types (HPV16 and HPV18 being most prevalent; genus Alphapapillomavirus) in women result in more prolonged cervical 223 224 infections, 80-90% of which are asymptomatic and are eventually cleared by the immune system 225 [66,67]. However, a small fraction of such infections, primarily due to defects in the immune 226 response, progress to the formation of persistent papillomas and, in a minority of cases, to cervical 227 cancer. This small fraction of HPV infections, nevertheless, accounts for close to 100% of cervical 228 cancers that affect over 500,000 women annually [69]. On much rarer occasions, the high-risk 229 HPV also cause other types of carcinomas [70]. Carcinogenesis is a dead end for HPV because 230 transformed cells produce no infectious virus. Therefore, despite the carcinogenic potential of 231 high-risk HPVs, by and large, these common components of human virome should be considered 232 symbionts, as demonstrated by the protection from skin cancer caused by immunity to ubiquitous 233 skin-infecting HPVs [71] (Figure 1b).

234

235 Anelloviruses

Members of the family *Anelloviridae* are among the most enigmatic components of the healthy human virome, both in terms of their evolutionary origin and the impact on human health. Anelloviruses have small icosahedral virions that encapsidate tiny (3 to 3,5 kb) circular ssDNA genomes [72], but unlike the ssDNA viruses in the realm *Monodnaviria*, do not encode recognizable homologs of the rolling circle replication endonuclease. Moreover, no homologs 241 outside the family have been detected for any of the anellovirus proteins. Hence, anelloviruses are 242 currently not included in the realm *Monodnaviria* and their provenance remains unclear [22,48]. The entire human population is believed to be infected with anelloviruses, and there is no 243 244 convincing evidence of viral clearance from infected individuals [72,73]. The infections occur at 245 an early age and so far have not been convincingly associated with any disease. The virus load 246 appears to be controlled by the immune system because virus levels increase with the level of host 247 immunosuppression [74,75]. Asymptomatic anellovirus infections are also common in other 248 mammals [73,76-78], suggesting extensive coevolution of anelloviruses with mammalian hosts. 249 Although the potential impact of anelloviruses on human health remains a matter of debate, it has 250 been suggested that they positively influence human physiology by shaping the immunity during 251 early development [72]. Thus, anelloviruses might be the most 'friendly', genuinely symbiotic 252 component of the human virome.

253

254 Realm Duplodnaviria

255 The realm Duplodnaviria includes viruses with dsDNA genomes that are encapsidated in 256 icosahedral capsids consisting of a distinct type of capsid protein, displaying the HK97 fold (after 257 the first phage for which the capsid protein structure was solved), with the help of the 258 corresponding variety of packaging ATPase, known as the terminase [22]. This realm includes the 259 bulk of the viruses associated with the human microbiome, namely, the numerous tailed 260 bacteriophages, as well as a major component of the virome associated with human cells, the herpesviruses. Some of the human herpesviruses (Herpesviridae) infect a variety of cell types and 261 262 cause life-long latent infections [79]. Of the 9 human herpesviruses identified so far, herpes simplex virus 1 (HSV1), human cytomegalovirus (HCMV), Epstein-Barr virus (EBV) and 263 264 varicella zoster virus (VZV) are highly prevalent in the human population. Depending on the geographic location, socioeconomic status and, in the case of VZV, vaccination levels, the 265 266 incidence of these viruses reaches up to 96% [80-82]. The health impact of these viruses (if any) 267 depends on the age and immune system status of the infected person, with ethnicity, gender and 268 genotype being additional significant contributors.

269

The most stealthy of the human herpes viruses are apparently EBV and HCMV. If acquired in childhood, both of these viruses typically cause asymptomatic infections, mainly, in Blymphocytes in the case of EBV [83] or in several cell types in the case of HCMV [79]. Such silent infections, however, even if not manifested in disease, cause a range of effects at the molecular, 274 cellular, tissue and organism levels. In the case of EBV, the gene expression pattern of infected B-275 cells is reprogrammed [84] and the proportion of plasma cells in blood appears to be increased [7]. 276 Major immunity stimulation effects, a likely result of a balance reached over long virus-host coevolution process, are well established for HCMV. In particular, ~10% of the CD4⁺ and CD8⁺ T 277 278 cells in latently infected, otherwise healthy adults are HCMV-specific. Likewise, HCMV causes 279 expansion of adaptive CD57⁺ natural killer (NK) cells targeting virus-infected cells [85]. These 280 powerful arms of the immune system, however, fail to clear the virus due to its armament of 281 immunoevasins, HCMV-encoded proteins involved in modulation of host immunity [86]. 282 Extensive immunity stimulation in HCMV-seropositive individuals appears to enhance 283 responsiveness and protection against heterologous viruses rather than compromise host immunity 284 [86]. These potentially positive effects notwithstanding, damaging consequences of herpesvirus 285 infections appear to be fairly common and vary from mild illness to a variety of life-threatening 286 conditions. Thus, HCMV congenital (in utero) infections that occur at up to 2% of childbirths 287 frequently cause neurodevelopmental defects or leukemia [85]. At the other life extremity, in 288 seniors, HCMV seropositivity is associated with increased risks of cancer, cardiovascular disease, 289 and ultimately, mortality.

290

291 The delicate, life-long EBV-host balance arising from most childhood infections is also fragile: if 292 infection occurs in even non-immunocompromised young adults, it results in infectious 293 mononucleosis ('kissing disease') [87]. Similarly, EBV is etiologically linked to Hodgkin's and 294 Burkitt's lymphomas and nasopharyngeal carcinoma which are common in Southern Chinese and 295 Eskimo people, as well as to a host of other diseases in immunocompromised individuals [83]. A 296 different pathology pattern is characteristic of VZV infections that cause varicella (chickenpox) in 297 children followed by prolonged latency that, in about 15% cases, leads to virus reactivation in 298 elderly people with weakened immune control, causing zoster (postherpetic neuralgia) [88].

299

The HSV1 infections exhibit another distinct pattern of virus-host interactions. This virus is normally acquired in childhood causing mainly oral infections; its seroprevalence varies from 70% in developed nations to 100% in developing nations [81,89]. Upon infecting epithelial cells, HSV1 is transmitted to axons and establishes life-long latency in dorsal root ganglia that is periodically manifested in reactivation leading to recurrent acute infections or asymptomatic virus shedding. Similar to other human herpesviruses, HSV1 efficiently evades antiviral innate immune responses mediated by Toll-like and other pathogen recognition receptors. The underlying mechanisms of HSV1 immunoevasion involve multiple virus proteins targeting diverse innate immunity signaling
pathways [90].

309

Thus, human herpesviruses display a striking variety of cell tropisms and infection patterns that 310 311 emerged over long-term virus-human co-evolution and co-adaptation. Some herpesviruses, in 312 particular HCMV and EBV, can reach a perfect balance with the human host and often persist for 313 the host's lifetime without causing any pathology. However, because of the complexity of virus-314 host interactions that involve a variety of genetic, environmental and socioeconomic factors, this 315 balance is fragile and can be broken in many situations resulting in morbidity or even mortality 316 (Figure 1b). Therefore, herpesvirus-host co-existence that involves the majority of the human 317 population blurs the very concept of a 'healthy human virome'.

318

319 Realm Varidnaviria

320 This realm includes a broad variety of viruses with dsDNA genomes and icosahedral capsids built 321 of protein unrelated to the capsid proteins of duplodnaviruses [22]. Unlike the members of 322 Duplodnaviria, varidnaviruses are minor components of the healthy human virome, at best. 323 Several intriguing reports have appeared on the detection of large and giant viruses of the class 324 *Nucleocytoviricota* in healthy humans [91]. Perhaps, the most notable of these findings is the 325 detection in human blood of several members of Marseilleviridae one of which has been reported 326 to grow in T lymphocytes [92]. Additionally, several viruses from both *Marseilleviridae* and 327 Mimiviridae have been isolated from human stools or detected in human-associated metagenomes 328 [93]. Furthermore, the presence of mimiviruses in peripheral blood mononuclear cells of patients 329 with atypical pneumonia has been reported, and a role for these viruses in the pneumonia 330 pathogenesis has been suggested [94]. Unexpectedly, DNA of Acanthocystis turfacea chlorella 331 virus 1, a member of *Phycodnaviridae*, has been detected in nearly half of the tested oropharyngeal samples from healthy humans, and also has been reported to persist in mouse macrophages [95]. 332 333 However, so far, many of the reports on the presence of members of Nucleocytoviricota in human 334 samples and, especially, their ability to replicate in human cells have been disavowed in follow-335 up studies [96]. Thus, the status of these viruses as components of the healthy human virome 336 except, perhaps, as occasional contaminants, remains dubious.

337

338 Conclusions

339 The healthy virome of any organism, and especially humans, clearly, is an important component

340 of the holobiont that makes a major contribution to the health status of the host. However, the very 341 concept of a healthy virome is nebulous and fluid because it is virtually impossible to ascertain 342 that any virus would not cause disease under any conditions. A strong case in point are the 343 herpesviruses that are nearly ubiquitous in the human population, remaining symbionts in most 344 individuals most of the time, but consistently cause disease, in some cases, devastating, in 345 immunocompromised individuals. Conversely, it appears plausible that any virus can become 346 beneficial to the host through protection from other viruses, general stimulation of immunity as in 347 the striking case of β -HPV protecting human hosts from skin cancer, or recruitment of virus genes 348 for host functions. The numerous HERVs integrated in the human genome can be considered the 349 paradigm of virus-host symbiosis. Generally, there is no doubt that many viruses evolved multiple 350 mechanisms to manipulate the host innate and adaptive immunity pathways, ensuring virus 351 persistence and controlling the damage to the host, as most conspicuously exemplified by the latent 352 herpesviruses that are virtually ubiquitous in the human population.

353

354 The healthy virome is obviously heterogeneous and consists of 3 distinct components (Figure 1a): 355 i) viruses that systematically enter the human organism, primarily, with food, but do not replicate 356 in humans, ii) viruses infecting prokaryotes and, possibly, unicellular eukaryotes that comprise the 357 healthy human microbiome, and iii) viruses that actually replicate and persist in human cells. With 358 the advances of metagenomics, the human "microbiovirome" has become a subject of intense 359 studies that continue bringing discoveries of new bacteriophage groups. In contrast, the "true" 360 healthy human virome is poorly understood, with many questionable sightings of diverse viruses 361 but little solid evidence on persistence mechanisms. On the whole, and in contrast to the diseaseassociated virome, the healthy human virome appears to be dominated by DNA viruses, in 362 363 particular, anelloviruses and herpesviruses, that are substantially more common than RNA viruses 364 in healthy humans. A thorough investigation of this component of the healthy virome can be 365 expected to enhance our understanding of virus-host interactions and have major implications for 366 human health.

367

368 Acknowledgements

E.V.K. is supported by the Intramural Research Program of the National Institutes of Health
(National Library of Medicine). M.K was supported by l'Agence Nationale de la Recherche grant
ANR-20-CE20-0009.

372 **References**

422

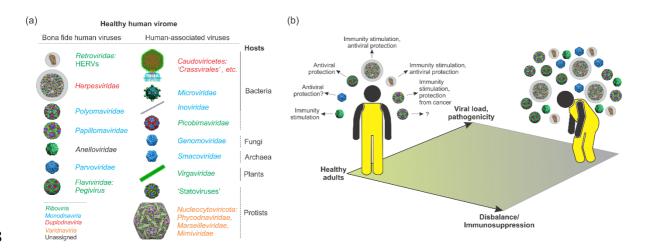
- 373 1. Koonin EV, Dolja VV: A virocentric perspective on the evolution of life. *Curr Opin Virol* 2013, 3:546-557.
- 375 2. Forterre P, Prangishvili D: The major role of viruses in cellular evolution: facts and hypotheses.
 376 *Curr Opin Virol* 2013, 3:558-565.
- 377 3. Roossinck MJ, Bazan ER: Symbiosis: Viruses as Intimate Partners. *Annu Rev Virol* 2017, 4:123-139.
- 4. Koonin EV, Makarova KS, Wolf YI, Krupovic M: Evolutionary entanglement of mobile genetic
 elements and host defence systems: guns for hire. *Nat Rev Genet* 2020, 21:119-131.
- 381 5. Rascovan N, Duraisamy R, Desnues C: Metagenomics and the Human Virome in Asymptomatic
 382 Individuals. *Annu Rev Microbiol* 2016, 70:125-141.
- 383 6. Adiliaghdam F, Jeffrey KL: Illuminating the human virome in health and disease. *Genome Med*384 2020, 12:66.
- **7. Kumata R, Ito J, Takahashi K, Suzuki T, Sato K: A tissue level atlas of the healthy human virome.
 BMC Biol 2020, 18:55.
- 387 The latest broad-scale metagenomics analysis of the healthy human virome.
- *8. Shkoporov AN, Clooney AG, Sutton TDS, Ryan FJ, Daly KM, Nolan JA, McDonnell SA,
 Khokhlova EV, Draper LA, Forde A, et al.: The Human Gut Virome Is Highly Diverse, Stable,
 and Individual Specific. *Cell Host Microbe* 2019, 26:527-541 e525.
- A detailed investigation of the diversity and short scale dynamics of the component of the humanhealthy virome associated with the gut microbiome.
- 393 9. Manrique P, Bolduc B, Walk ST, van der Oost J, de Vos WM, Young MJ: Healthy human gut phageome. *Proc Natl Acad Sci U S A* 2016, 113:10400-10405.
- 395 10. Wang H, Ling Y, Shan T, Yang S, Xu H, Deng X, Delwart E, Zhang W: Gut virome of mammals
 396 and birds reveals high genetic diversity of the family Microviridae. *Virus Evol* 2019, 5:vez013.
- 397 11. Krupovic M, Forterre P: Microviridae goes temperate: microvirus-related proviruses reside in the
 398 genomes of Bacteroidetes. *PLoS One* 2011, 6:e19893.
- 399 12. Minot S, Bryson A, Chehoud C, Wu GD, Lewis JD, Bushman FD: Rapid evolution of the human gut virome. *Proc Natl Acad Sci U S A* 2013, 110:12450-12455.
- *13. Roux S, Krupovic M, Daly RA, Borges AL, Nayfach S, Schulz F, Sharrar A, Matheus Carnevali
 PB, Cheng JF, Ivanova NN, et al.: Cryptic inoviruses revealed as pervasive in bacteria and archaea
 across Earth's biomes. *Nat Microbiol* 2019, 4:1895-1906.
- An in depth search for inoviruses integrated in bacterial genomes, including those of the human gut
 microbiota, using the most powerful computational approaches. The results greatly expand the
 diversity of inovorises and demonstrated their unsuspected high abundance.
- 407 14. Dutilh BE, Cassman N, McNair K, Sanchez SE, Silva GG, Boling L, Barr JJ, Speth DR, Seguritan
 408 V, Aziz RK, et al.: A highly abundant bacteriophage discovered in the unknown sequences of
 409 human faecal metagenomes. *Nat Commun* 2014, 5:4498.
- 410 15. Yutin N, Makarova KS, Gussow AB, Krupovic M, Segall A, Edwards RA, Koonin EV: Discovery
 411 of an expansive bacteriophage family that includes the most abundant viruses from the human gut.
 412 *Nat Microbiol* 2018, 3:38-46.
- *16. Yutin N, Benler S, Shmakov SA, Wolf YI, Tolstoy I, Rayko M, Antipov D, Pevzner PA,
 KooninE.V.: Analysis of metagenome-assembled viral genomes from the human gut reveals
 diverse putative CrAss-like phages with unique genomic features *Nature Communications* 2021,
 in press.
- A comprehensive analysis of virus genomes from the human gut dramatically expanding the diversity
 of crAss-like phages and demonstrating that this group of phages dominates the human
 "microbiovirome". Some subgroups of crAss-like phages are shown to possess unique gene
 organizations, with unprecedented densities of self-splicing introns and intein, and alternative
 genetic codes.
- 423 17. Guerin E, Shkoporov A, Stockdale SR, Clooney AG, Ryan FJ, Sutton TDS, Draper LA, Gonzalez424 Tortuero E, Ross RP, Hill C: Biology and Taxonomy of crAss-like Bacteriophages, the Most
 425 Abundant Virus in the Human Gut. *Cell Host Microbe* 2018, 24:653-664 e656.

- 426 18. Drobysheva AV, Panafidina SA, Kolesnik MV, Klimuk EI, Minakhin L, Yakunina MV, Borukhov
 427 S, Nilsson E, Holmfeldt K, Yutin N, et al.: Structure and function of virion RNA polymerase of a
 428 crAss-like phage. *Nature* 2020.
- 429 19. Norman JM, Handley SA, Baldridge MT, Droit L, Liu CY, Keller BC, Kambal A, Monaco CL,
 430 Zhao G, Fleshner P, et al.: Disease-specific alterations in the enteric virome in inflammatory bowel
 431 disease. *Cell* 2015, 160:447-460.
- 432 20. Monaco CL, Gootenberg DB, Zhao G, Handley SA, Ghebremichael MS, Lim ES, Lankowski A,
 433 Baldridge MT, Wilen CB, Flagg M, et al.: Altered Virome and Bacterial Microbiome in Human
 434 Immunodeficiency Virus-Associated Acquired Immunodeficiency Syndrome. *Cell Host Microbe*435 2016, 19:311-322.
- 436 21. Handley SA, Desai C, Zhao G, Droit L, Monaco CL, Schroeder AC, Nkolola JP, Norman ME,
 437 Miller AD, Wang D, et al.: SIV Infection-Mediated Changes in Gastrointestinal Bacterial
 438 Microbiome and Virome Are Associated with Immunodeficiency and Prevented by Vaccination.
 439 *Cell Host Microbe* 2016, 19:323-335.
- **22. Koonin EV, Dolja VV, Krupovic M, Varsani A, Wolf YI, Yutin N, Zerbini FM, Kuhn JH: Global
 Organization and Proposed Megataxonomy of the Virus World. *Microbiol Mol Biol Rev* 2020, 84.
- 442 A comprehensive analysis of the evolutionary relationships among the major groups of viruses. A
 443 megataxonomy of viruses is proposed on the basis of these relationships and subsequently adopted
 444 by the International Committee for Taxonomy of Viruses (Ref. 23).
- 445 23. International Committee on Taxonomy of Viruses Executive C: The new scope of virus taxonomy:
 446 partitioning the virosphere into 15 hierarchical ranks. *Nat Microbiol* 2020, 5:668-674.
- 447 24. Stapleton JT, Foung S, Muerhoff AS, Bukh J, Simmonds P: The GB viruses: a review and proposed
 448 classification of GBV-A, GBV-C (HGV), and GBV-D in genus Pegivirus within the family
 449 Flaviviridae. J Gen Virol 2011, 92:233-246.
- 450 25. Hartlage AS, Cullen JM, Kapoor A: The Strange, Expanding World of Animal Hepaciviruses.
 451 *Annu Rev Virol* 2016, 3:53-75.
- 452 26. Linnen J, Wages J, Jr., Zhang-Keck ZY, Fry KE, Krawczynski KZ, Alter H, Koonin E, Gallagher
 453 M, Alter M, Hadziyannis S, et al.: Molecular cloning and disease association of hepatitis G virus: 454 a transfusion-transmissible agent. *Science* 1996, 271:505-508.
- 455 27. Alter MJ, Gallagher M, Morris TT, Moyer LA, Meeks EL, Krawczynski K, Kim JP, Margolis HS:
 456 Acute non-A-E hepatitis in the United States and the role of hepatitis G virus infection. Sentinel
 457 Counties Viral Hepatitis Study Team. *N Engl J Med* 1997, 336:741-746.
- 458 28. Schwarze-Zander C, Blackard JT, Rockstroh JK: Role of GB virus C in modulating HIV disease.
 459 *Expert Rev Anti Infect Ther* 2012, 10:563-572.
- 460 29. Greenhalgh S, Schmidt R, Day T: Fighting the Public Health Burden of AIDS With the Human
 461 Pegivirus. *Am J Epidemiol* 2019, 188:1586-1594.
- 30. Janowski AB, Krishnamurthy SR, Lim ES, Zhao G, Brenchley JM, Barouch DH, Thakwalakwa C,
 Manary MJ, Holtz LR, Wang D: Statoviruses, A novel taxon of RNA viruses present in the
 gastrointestinal tracts of diverse mammals. *Virology* 2017, 504:36-44.
- 31. Thi Kha Tu N, Thi Thu Hong N, Thi Han Ny N, My Phuc T, Thi Thanh Tam P, Doorn HRV, Dang
 Trung Nghia H, Thao Huong D, An Han D, Thi Thu Ha L, et al.: The Virome of Acute Respiratory
 Diseases in Individuals at Risk of Zoonotic Infections. *Viruses* 2020, 12.
- 468 32. Wolf YI, Kazlauskas D, Iranzo J, Lucia-Sanz A, Kuhn JH, Krupovic M, Dolja VV, Koonin EV:
 469 Origins and Evolution of the Global RNA Virome. *MBio* 2018, 9.
- 470 33. Ganesh B, Banyai K, Martella V, Jakab F, Masachessi G, Kobayashi N: Picobirnavirus infections:
 471 viral persistence and zoonotic potential. *Rev Med Virol* 2012, 22:245-256.
- 472 34. Ganesh B, Masachessi G, Mladenova Z: Animal picobirnavirus. *Virusdisease* 2014, 25:223-238.
- 473 *35. Krishnamurthy SR, Wang D: Extensive conservation of prokaryotic ribosomal binding sites in known and novel picobirnaviruses. *Virology* 2018, 516:108-114.
- 475 Unexpected observation of highly conserved Shinne-Dalgarno sequences in picobirnaviruses leading
 476 to the proposal that these viruses are parasites of prokaryotes.
- 477 36. Zhang T, Breitbart M, Lee WH, Run JQ, Wei CL, Soh SW, Hibberd ML, Liu ET, Rohwer F, Ruan 478 X: PNA viral community in human faces: provalance of plant pathogenic viruses. *PLoS Biol* 2006
- 478 Y: RNA viral community in human feces: prevalence of plant pathogenic viruses. *PLoS Biol* 2006,
 479 4:e3.

- **37. Symonds EM, Rosario K, Breitbart M: Pepper mild mottle virus: Agricultural menace turned
 effective tool for microbial water quality monitoring and assessing (waste)water treatment
 technologies. *PLoS Pathog* 2019, 15:e1007639.
- 483 Demonstration of the ubiquity and high abundance of a food-associated plant virus in the human gut.
- 38. Tokuyama M, Kong Y, Song E, Jayewickreme T, Kang I, Iwasaki A: ERVmap analysis reveals
 genome-wide transcription of human endogenous retroviruses. *Proc Natl Acad Sci U S A* 2018, 115:12565-12572.
- 487 39. Johnson WE: Origins and evolutionary consequences of ancient endogenous retroviruses. *Nat Rev* 488 *Microbiol* 2019, 17:355-370.
- The latest detailed overview of the human endogenous retroviruses. The dual role of retroviruses in animal evolution is emphasized: first, as driveers of defense systems evolution, and second, as reservoir of genetic material for exaptation.
- 492 40. Chiu ES, VandeWoude S: Endogenous Retroviruses Drive Resistance and Promotion of
 493 Exogenous Retroviral Homologs. *Annu Rev Anim Biosci* 2020.
- 494 41. Cullen H, Schorn AJ: Endogenous Retroviruses Walk a Fine Line between Priming and Silencing.
 495 *Viruses* 2020, 12.
- 496 *42. Alcazer V, Bonaventura P, Depil S: Human Endogenous Retroviruses (HERVs): Shaping the
 497 Innate Immune Response in Cancers. *Cancers (Basel)* 2020, 12.
- 498 A conceptual article discussing the stimulatory effect of endogenous retroviruses on innate immunity
 499 promoting tumor resistance.
- 43. Koonin EV, Krupovic M: The depths of virus exaptation. *Curr Opin Virol* 2018, 31:1-8.
- 44. Garcia-Montojo M, Doucet-O'Hare T, Henderson L, Nath A: Human endogenous retrovirus-K
 (HML-2): a comprehensive review. *Crit Rev Microbiol* 2018, 44:715-738.
- 503 45. Xue B, Sechi LA, Kelvin DJ: Human Endogenous Retrovirus K (HML-2) in Health and Disease.
 504 *Front Microbiol* 2020, 11:1690.
- 46. Grandi N, Tramontano E: Human Endogenous Retroviruses Are Ancient Acquired Elements Still
 Shaping Innate Immune Responses. *Front Immunol* 2018, 9:2039.
- 507 *47. Singh M, Cai H, Bunse M, Feschotte C, Izsvak Z: Human Endogenous Retrovirus K Rec forms
 508 a Regulatory Loop with MITF that Opposes the Progression of Melanoma to an Invasive Stage.
 509 *Viruses* 2020, 12.
- This work uncovers the specific mechanism of prevention of the progression of invasive melanoma by
 the expression of the endogenous viruses of the HERV-K group.
- 48. Kazlauskas D, Varsani A, Koonin EV, Krupovic M: Multiple origins of prokaryotic and eukaryotic
 single-stranded DNA viruses from bacterial and archaeal plasmids. *Nat Commun* 2019, 10:3425.
- 49. Krupovic M, Ghabrial SA, Jiang D, Varsani A: Genomoviridae: a new family of widespread singlestranded DNA viruses. *Arch Virol* 2016, 161:2633-2643.
- 516 50. Varsani A, Krupovic M: Smacoviridae: a new family of animal-associated single-stranded DNA viruses. *Arch Virol* 2018, 163:2005-2015.
- 51. Matthews PC, Sharp C, Simmonds P, Klenerman P: Human parvovirus 4 'PARV4' remains elusive
 despite a decade of study. *F1000Res* 2017, 6:82.
- 520 52. Qiu J, Soderlund-Venermo M, Young NS: Human Parvoviruses. *Clin Microbiol Rev* 2017, 30:43 521 113.
- 53. Soderlund-Venermo M: Emerging Human Parvoviruses: The Rocky Road to Fame. *Annu Rev Virol* 2019, 6:71-91.
- 524 54. Cotmore SF, Agbandje-McKenna M, Canuti M, Chiorini JA, Eis-Hubinger AM, Hughes J,
 525 Mietzsch M, Modha S, Ogliastro M, Penzes JJ, et al.: ICTV Virus Taxonomy Profile: Parvoviridae.
 526 J Gen Virol 2019, 100:367-368.
- 527 55. Allander T: Human bocavirus. *J Clin Virol* 2008, 41:29-33.
- 528 56. Guido M, Tumolo MR, Verri T, Romano A, Serio F, De Giorgi M, De Donno A, Bagordo F, Zizza
 529 A: Human bocavirus: Current knowledge and future challenges. *World J Gastroenterol* 2016, 22:8684-8697.
- 57. Ma X, Endo R, Ishiguro N, Ebihara T, Ishiko H, Ariga T, Kikuta H: Detection of human bocavirus
 in Japanese children with lower respiratory tract infections. *J Clin Microbiol* 2006, 44:1132-1134.
- 58. Woolf AD, Campion GV, Chishick A, Wise S, Cohen BJ, Klouda PT, Caul O, Dieppe PA: Clinical
 manifestations of human parvovirus B19 in adults. *Arch Intern Med* 1989, 149:1153-1156.

- 535 59. Bonvicini F, La Placa M, Manaresi E, Gallinella G, Gentilomi GA, Zerbini M, Musiani M:
 536 Parvovirus b19 DNA is commonly harboured in human skin. *Dermatology* 2010, 220:138-142.
- 60. Moens U, Krumbholz A, Ehlers B, Zell R, Johne R, Calvignac-Spencer S, Lauber C: Biology,
 evolution, and medical importance of polyomaviruses: An update. *Infect Genet Evol* 2017, 54:1838.
- 540 61. Spriggs CC, Laimins LA: Human Papillomavirus and the DNA Damage Response: Exploiting
 541 Host Repair Pathways for Viral Replication. *Viruses* 2017, 9.
- 542 62. Buck CB, Van Doorslaer K, Peretti A, Geoghegan EM, Tisza MJ, An P, Katz JP, Pipas JM,
 543 McBride AA, Camus AC, et al.: The Ancient Evolutionary History of Polyomaviruses. *PLoS*544 *Pathog* 2016, 12:e1005574.
- 545 63. Krump NA, Liu W, You J: Mechanisms of persistence by small DNA tumor viruses. *Curr Opin Virol* 2018, 32:71-79.
- 547 64. Cortese I, Reich DS, Nath A: Progressive multifocal leukoencephalopathy and the spectrum of JC
 548 virus-related disease. *Nat Rev Neurol* 2020.
- 549 65. Krump NA, You J: Molecular mechanisms of viral oncogenesis in humans. *Nat Rev Microbiol* 2018, 16:684-698.
- 66. Doorbar J, Egawa N, Griffin H, Kranjec C, Murakami I: Human papillomavirus molecular biology
 and disease association. *Rev Med Virol* 2015, 25 Suppl 1:2-23.
- 67. Graham SV: The human papillomavirus replication cycle, and its links to cancer progression: a comprehensive review. *Clin Sci (Lond)* 2017, 131:2201-2221.
- 555 68. Chiu YF, Sugden B: Plasmid Partitioning by Human Tumor Viruses. J Virol 2018, 92.
- 69. Serrano B, Brotons M, Bosch FX, Bruni L: Epidemiology and burden of HPV-related disease. *Best Pract Res Clin Obstet Gynaecol* 2018, 47:14-26.
- 558 70. Araldi RP, Sant'Ana TA, Modolo DG, de Melo TC, Spadacci-Morena DD, de Cassia Stocco R,
 559 Cerutti JM, de Souza EB: The human papillomavirus (HPV)-related cancer biology: An overview.
 560 *Biomed Pharmacother* 2018, 106:1537-1556.
- **71. Strickley JD, Messerschmidt JL, Awad ME, Li T, Hasegawa T, Ha DT, Nabeta HW, Bevins PA,
 Ngo KH, Asgari MM, et al.: Immunity to commensal papillomaviruses protects against skin
 cancer. *Nature* 2019, 575:519-522.
- 564 Squamous cell skin carcinoma has been associated with the replication of human b-HPV. This work
 565 shows that such correlation does not reflect causative role of b-HPV in cancer progression, but on
 566 the contrary, occurs because T call mediated immunity against b-HPV suppresses tumorigenesis.
- **72. Kaczorowska J, van der Hoek L: Human anelloviruses: diverse, omnipresent and commensal
 members of the virome. *FEMS Microbiol Rev* 2020, 44:305-313.
- The latest review on anelloviruses emphasizing the ubiquity of these viruses in humans, the lack ofconvincing disease association and the potential beneficial roles.
- 571 73. Webb B, Rakibuzzaman A, Ramamoorthy S: Torque teno viruses in health and disease. *Virus Res* 2020, 285:198013.
- 573 74. Focosi D, Antonelli G, Pistello M, Maggi F: Torquetenovirus: the human virome from bench to
 574 bedside. *Clin Microbiol Infect* 2016, 22:589-593.
- 575 75. Blatter JA, Sweet SC, Conrad C, Danziger-Isakov LA, Faro A, Goldfarb SB, Hayes D, Jr., Melicoff
 576 E, Schecter M, Storch G, et al.: Anellovirus loads are associated with outcomes in pediatric lung
 577 transplantation. *Pediatr Transplant* 2018, 22.
- 578 76. Fahsbender E, Burns JM, Kim S, Kraberger S, Frankfurter G, Eilers AA, Shero MR, Beltran R,
 579 Kirkham A, McCorkell R, et al.: Diverse and highly recombinant anelloviruses associated with
 580 Weddell seals in Antarctica. *Virus Evol* 2017, 3:vex017.
- 581 77. de Souza WM, Fumagalli MJ, de Araujo J, Sabino-Santos G, Jr., Maia FGM, Romeiro MF, Modha
 582 S, Nardi MS, Queiroz LH, Durigon EL, et al.: Discovery of novel anelloviruses in small mammals
 583 expands the host range and diversity of the Anelloviridae. *Virology* 2018, 514:9-17.
- 584 78. Crane A, Goebel ME, Kraberger S, Stone AC, Varsani A: Novel anelloviruses identified in buccal
 585 swabs of Antarctic fur seals. *Virus Genes* 2018, 54:719-723.
- 586 79. Connolly SA, Jardetzky TS, Longnecker R: The structural basis of herpesvirus entry. *Nat Rev* 587 *Microbiol* 2020.
- 588 80. Cohen JI: Epstein-Barr virus infection. *N Engl J Med* 2000, 343:481-492.
- 589 81. Gupta R, Warren T, Wald A: Genital herpes. *Lancet* 2007, 370:2127-2137.

- 590 82. Zuhair M, Smit GSA, Wallis G, Jabbar F, Smith C, Devleesschauwer B, Griffiths P: Estimation of
 591 the worldwide seroprevalence of cytomegalovirus: A systematic review and meta-analysis. *Rev*592 *Med Virol* 2019, 29:e2034.
- 593 83. Taylor GS, Long HM, Brooks JM, Rickinson AB, Hislop AD: The immunology of Epstein-Barr virus-induced disease. *Annu Rev Immunol* 2015, 33:787-821.
- 84. Moustafa A, Xie C, Kirkness E, Biggs W, Wong E, Turpaz Y, Bloom K, Delwart E, Nelson KE,
 Venter JC, et al.: The blood DNA virome in 8,000 humans. *PLoS Pathog* 2017, 13:e1006292.
- 597 85. Semmes EC, Hurst JH, Walsh KM, Permar SR: Cytomegalovirus as an immunomodulator across
 598 the lifespan. *Curr Opin Virol* 2020, 44:112-120.
- **86. Berry R, Watson GM, Jonjic S, Degli-Esposti MA, Rossjohn J: Modulation of innate and adaptive immunity by cytomegaloviruses. *Nat Rev Immunol* 2020, 20:113-127.
- A comprehensive discussion of immunoevasins, the diverse proteins encoded by human
 cytomegaloviruses involved in the evasion and modulation of the host immune system.
- 603 87. Cruz-Munoz ME, Fuentes-Panana EM: Beta and Gamma Human Herpesviruses: Agonistic and
 604 Antagonistic Interactions with the Host Immune System. *Front Microbiol* 2017, 8:2521.
- 88. Gershon AA, Breuer J, Cohen JI, Cohrs RJ, Gershon MD, Gilden D, Grose C, Hambleton S,
 Kennedy PG, Oxman MN, et al.: Varicella zoster virus infection. *Nat Rev Dis Primers* 2015, 1:15016.
- 608 89. Saleh D, Yarrarapu SNS, Sharma S: Herpes Simplex Type 1. In *StatPearls*. Edited by; 2020.
- 609 90. Zhu H, Zheng C: The Race between Host Antiviral Innate Immunity and the Immune Evasion
 610 Strategies of Herpes Simplex Virus 1. *Microbiol Mol Biol Rev* 2020, 84.
- 611 91. Colson P, Aherfi S, La Scola B, Raoult D: The role of giant viruses of amoebas in humans. *Curr*612 *Opin Microbiol* 2016, 31:199-208.
- 92. Popgeorgiev N, Boyer M, Fancello L, Monteil S, Robert C, Rivet R, Nappez C, Azza S, Chiaroni
 J, Raoult D, et al.: Marseillevirus-like virus recovered from blood donated by asymptomatic
 humans. J Infect Dis 2013, 208:1042-1050.
- 616 93. Colson P, Fancello L, Gimenez G, Armougom F, Desnues C, Fournous G, Yoosuf N, Million M,
 617 La Scola B, Raoult D: Evidence of the megavirome in humans. *J Clin Virol* 2013, 57:191-200.
- 618 94. Colson P, La Scola B, Raoult D: Giant viruses of amoebae as potential human pathogens.
 619 *Intervirology* 2013, 56:376-385.
- 95. Petro TM, Agarkova IV, Zhou Y, Yolken RH, Van Etten JL, Dunigan DD: Response of
 Mammalian Macrophages to Challenge with the Chlorovirus Acanthocystis turfacea Chlorella
 Virus 1. *J Virol* 2015, 89:12096-12107.
- 623 96. Abrahao J, Silva L, Oliveira D, Almeida G: Lack of evidence of mimivirus replication in human
 624 PBMCs. *Microbes Infect* 2018, 20:281-283.
- 625
- 626



628

629 Figure 1. The healthy human virome.

630 (a) Taxa of viruses found in healthy humans. Taxa of viruses replicating in human cells are shown on the left, whereas those of viruses infecting human-associated microbes or associated with food 631 632 sources are shown on the right (the actual or suspected hosts are listed next to the corresponding 633 taxa, with the uncertain assignments indicated with broken lines). Depicted virus taxa are represented with virion structures which were retrieved from VIPERdb (viperdb.scripps.edu) or 634 635 the Electron Microscopy Data Bank (https://www.ebi.ac.uk/pdbe/emdb/). When the structure of a virus representing a particular taxon was not available, a structurally related member was chosen 636 637 instead. Virus taxa are colored according to their realm affiliation (the key is provided at the bottom). 638

(b) Fluidity of the healthy human virome. The (potential) beneficial effects of the healthy human virome are indicated next to the corresponding virus structures. Question marks denote uncertainty. Upon changes in the health status/immunosuppression, viruses that cause asymptomatic infections or are beneficial in healthy individuals proliferate and can cause diseases including severe ones. The figure illustrates the general tendency of increased virus load under immunosuppression/disease conditions and should not be interpreted as a quantitative representation of the changes for any depicted group of viruses.

627