The presence of Pneumocystis jirovecii in critically ill patients with COVID-19

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Key words: COVID-19; pneumocystosis; Pneumocystis jirovecii

- 1 Letters to the Editor
- 2 Dear Editor,

3 We read with interest the recent review on co-infections in coronavirus disease-2019 (COVID-19) 4 patients¹ and believe that fungal co-infections as evaluated from selected studies are underestimated. 5 Severe acute respiratory syndrome coronavirus-2 (SARS-CoV2) is still spreading pandemically. Approximately 5-10% of COVID-19 patients may require intensive care unit (ICU) management and 6 7 30% may develop secondary pneumonia without identified etiology.² Hospital-acquired bacterial or 8 fungal superinfections, as described in critically ill patients with *Influenza* virus, can be suspected.³ Since 9 pneumocystosis is usually reported in patients with T-cell immunodepression,⁴ less attention has been 10 paid to Pneumocystis jirovecii in non-immunocompromised ICU patients although it accounts for 7% of 11 the co-infections reported in those admitted with Influenza.⁵ Interestingly, COVID-19 patients may 12 develop lymphocytopenia and acute respiratory distress syndrome (ARDS) requiring adjunctive steroids and/or immunomodulatory therapies, well-known susceptibility factors for developing pneumocystosis.⁵ 13 14 We designed this observational cohort study to investigate the prevalence of *P. jirovecii* acid nucleic 15 detection in respiratory specimens sampled to identify co-infections in COVID-19 patients in the ICU.

All consecutive patients admitted to the ICU between 2020/03/15 and 2020/05/01 with a positive SARS-CoV-2 PCR (Cobas[®] SARS-CoV-2 Test, Roche, France) and ≥ 1 respiratory sample (bronchoalveolar lavage (BAL), tracheal aspirate, sputum) sent to the mycology department. This study was part of the COVID-ICU and French COVID-19 cohort registries. Our institutional ethics committee approved the study (IDRCB, 2020-A00256-33; CPP, 11-20-20.02.04.68737). When possible, signed informed consent was obtained from the patients or the next of kin.

Whenever possible, bronchoalveolar lavage (BAL) was performed in the middle lobe by a trained
pneumologist in the ICU using at least 120 ml saline (yield, ~50%). Upon reception, specimens including
BAL fluids and dTT-treated aspirations (dTT 1X at 37°C for 15min) were centrifuged, suspended in
200µL of water and submitted to extraction (whole nucleic acids extraction) using the GeneLead-VIII
extractor-thermocyclerTM (Precision System Science, Japan). *P. jirovecii* reverse transcriptase quantitative
PCR (RTqPCR) was performed to amplify mtSSU and mtLSU RNA and DNA of *P. jirovecii* using the
new R-DiaPnJ kitTM (Diagenode, Belgium). Serum β-D-glucan was tested using the Fungitell kitTM (Cape

Cod Inc, US) as recommended by the manufacturer. Data are presented as median [25th-75th percentiles]
or percentages as appropriate. Comparisons were performed using Mann-Whitney or exact Fisher tests as
required. *P*-values ≤0.05 were considered as significant.

32 One hundred-and-eight successive HIV-negative COVID-19 patients (Male/Female sex ratio, 4.4; age, 33 62 years [56-68]) with the usual risk factors for severe COVID-19 presentation were included (Table 1). 34 All except three patients were intubated on admission. Thirty-four patients (31.4%) who developed 35 ARDS received at least one day of corticosteroids before BAL sampling. Respiratory samples included 80 36 BALs (74.1%), 22 tracheal aspirates (20.4%), 4 sputa (3.7%) and two bronchial aspiration fluids (1.9%). 37 In 10/108 patients (9.3%), P. jirovecii RTqPCR was positive. Median delay between sampling and ICU 38 admission was 2days [1-2]. The median quantitative cycle value was 32.6 [30.8-34.7]. Serum β -D-glucan 39 were measured in nine patients and was negative (>80pg/mL) in seven patients.

40 Clinical characteristics of the patients carrying P. jirovecii did not significantly differ from the other 41 patients except for lower plasma D-dimer (1,270ng/mL [750-2,390] vs 2,610ng/mL [1,405-4,700], 42 P=0.03) and more frequent lopinavir/ritonavir administration (40.0% vs 12.2%, P=0.04), while long-term 43 corticosteroid prescription tended to be more frequent (30.0% vs 8.2%, P=0.06). Of note, among our 10 44 P. jirovecii carriers, five concomitantly met the criteria for COVID-19-associated pulmonary 45 aspergillosis.⁶ Out of these patients, four (40%) received co-trimoxazole as prophylaxis (80/400mg once 46 daily) whereas six including four who rapidly improved did not. One co-trimoxazole-treated and two non-47 treated patients died while the seven remaining patients were discharged. Mortality was similar in both 48 groups.

We found an unexpectedly high proportion of critically ill COVID-19 patients detected with *P*. *jirovecii* (10/108 patients; 9.3%), similarly to previous findings in influenza patients (3/45; ~7%).⁵

The presence of *P. jirovecii* in the healthy adult population has been measured using oropharyngeal wash samples obtained by gargling and examined by conventional or nested PCR methods.⁷ However, experts agree that the reported prevalence (\sim 20%) has been overestimated due to technical issues such as contamination with amplicons responsible for false positives.⁴ In our center managing almost exclusively immunocompromised patients, prevalence of qPCR-positive respiratory specimens with fungal load as low as in our COVID-19 patients, is \sim 13% (unpublished data), as reported elsewhere.³ COVID-19 patients mostly exhibited marked lymphopenia and alterations in lymphocyte functions,⁸ likely explaining
the high-rate of *P. jirovecii* detection.

59 Since serum β -D-glucan is advocated in pneumocystosis diagnosis,⁴ we measured its concentrations in 60 four of our five P. jirovecii RTqPCR-positive patients and obtained low values (<120pg/mL) in 61 accordance with the low nucleic acids fungal loads in the lung alveoli.⁹ Of note, in two out of our nine 62 tested P. jirovecii RTqPCR-negative patients, higher B-D-glucan concentrations (450 and 500pg/ml) lead 63 to the diagnosis of pulmonary aspergillosis, another fungal infection of risk in COVID-19 patients.⁶ Although a recent meta-analysis questioned its sensitivity in non-HIV patients,¹⁰ β-D-glucan has been 64 65 widely used to rule out pneumocystosis because of its high negative predictive value. This finding may 66 support the hypothesis that our patients were carrying *P. jirovecii*, yet not being infected *per se*. Thus, 67 although interesting in the context of invasive fungal infections diagnosis, serum β-D-glucan should be 68 interpreted with caution when excluding the diagnosis of pneumocystosis.

Here, four out of ten *P. jirovecii* RTqPCR-positive patients received co-trimoxazole as prophylactic regimen, based on the treating physician's decision. Whether a positive result should be an indication to consider administering co-trimoxazole, at least at prophylactic dosage in COVID-19 patients remains questionable.

Our study limitations include the relatively small number of patients, the bi-center setting, and the short study period. However, to the best of our knowledge, this is the first study evaluating the prevalence of *P. jirovecii* in COVID-19 patients. Because we focused on critically ill COVID-19 patients, *P. jirovecii* prevalence in less severe patients remains to be determined.

In conclusion, an unexpectedly high proportion of *P. jirovecii*-positive pulmonary samples is observed
in critically ill COVID-19 patients. Based on our findings, we advocate systematically searching for *P. jirovecii* in deep respiratory specimens in these patients. We believe that this strategy may be useful in
limiting enhanced inflammation due to the presence of *P. jirovecii* in the lung and avoiding inter-patient *P. jirovecii* transmission.

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83 Declaration of Competing Interest

84 The authors declare that they have no competing interests.

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86	Ethics approval and consent to participate
87	This study was part of the French COVID-19 cohort registry conducted by the REACTing consortium
88	(REsearch and ACTion targeting emerging infectious diseases) and directed by INSERM (Institut national de la
89	santé et de la recherche médicale) and ISARIC (International Severe Acute Respiratory and Emerging Infection
90	Consortium). Our institutional ethics committee approved the study (N°, IDRCB, 2020-A00256-33; CPP, 11-20-
91	20.02.04.68737).
92	
93	Availability of data and materials
94	Drs. Alanio, Mégarbane and Bretagne conceived of and designed the study. Drs. Voicu, Azoulay and
95	Mégarbane managed the patients. Drs. Alanio, Dellière and Bretagne performed the microbiological analysis. All
96	authors acquired, analyzed the data and interpreted the results. Drs. Alanio, Mégarbane and Bretagne drafted the
97	manuscript. All authors participated to the critical revision of the manuscript for important intellectual content.
98	Dr. Alanio has full access to all data and takes responsibility for the data integrity and its analysis accuracy.
99	
100	Consent for publication
101	All the authors agree to publish.
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105	at the university hospital.
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- 137 *Microbiol Infect* 2020;S1198-743X(20)30301-3. doi:10.1016/j.cmi.2020.05.024

138 Table 1

Characteristics of hundred-and-eight critically ill COVID-19 patients according to *Pneumocystis jirovecii*detection in the respiratory samples. Data are presented as percentages or medians [25th-75th percentiles].
Comparisons were performed using Mann-Whitney (•) or Fisher exact tests (*), as appropriate.

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	Total (N=108)	No detection of <i>P. jirovecii</i> (N=98)	Detection of P. jirovecii (N=10)	Р
Male gender, N (%)	88 (81.5%)	80 (81.6%)	8 (80.0%)	1*
Age (years)	62 [56-68]	62 [56-68]	59 [46-68]	0.400
COVID-19 risk factors		<u> </u>		
Past hypertension, N (%)	64 (59.3%)	58 (59.2%)	6 (60.0%)	1*
Diabetes, N (%)	40 (37.0%)	37 (37.8%)	3 (30.0%)	0.74*
Obesity, N (%)	35 (32.4%)	32 (32.7%)	3 (30.0%)	1*
Coronary disease, N (%)	15 (13.9%)	14 (14.3%)	1 (10.0%)	1*
Body-mass index (kg/m ²)	28 [25-31]	28 [25-31]	28 [27-32]	0.610
Other remarkable comorbidities				
Asthma, N (%)	5 (4.6%)	4 (4.1%)	1 (10.0%)	0.39*
Chronic obstructive pulmonary disease, N (%)	2 (1.9%)	2 (2.0%)	0 (0.0%)	1*
Immunocompromised patient, N (%)	10 (9.3%)	10 (10.2%)	0 (0.0%)	0.59*
Long-term corticosteroids, N (%)	11 (10.2%)	8 (8.2%)	3 (30.0%)	0.06*
Biological data of interest on admission	<u> </u>			
PaO ₂ /FiO ₂ (mmHg)	137 [83-247]	134 [83-239]	177 [108-253]	0.600

Plasma D-dimer (ng/mL) 2,2395 [1,193- 4,635] 2,610 [1,405- 4,700] 1,270 [750- 2,390] 0.03 Serum lactate dehydrogenase (1U/L) 687 [540-901] 687 [568-903] 708 [436-893] 0.65 BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL polymorphonuclear cells 37 [26-81] 46 [26-81] 29 [18-32] 0.54 % BAL Lymphocytes 13 [6-32] 14 [5-34] 13 [10-23] 0.81 Specific anti-COVID-19 therapy, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 3 (3.2%) 3 (3.1%) 0 (0.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 4 (3.7%) 4 (4.1%) 2 (20.0%)					
Plasma D-dimer (ng/mL) 4,635] 4,700] 2,390] 0.03 Serum lactate dehydrogenase (1U/L) 687 [540-901] 687 [568-903] 708 [436-893] 0.65 Bronchoalveolar lavage characteristics	Serum creatinine (µmol/L)	80 [64-111]	80 [63-111]	80 [67-104]	0.990
Bronchoalveolar layage characteristics % BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL polymorphonuclear cells 37 [26-81] 46 [26-81] 29 [18-32] 0.54 % BAL Lymphocytes 13 [6-32] 14 [5-34] 13 [10-23] 0.81 Specific anti-COVID-19 therapy, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Lopinavir-ritonavir, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 4 (3.7%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 53 (49.1%) 15 (15.3%) 1 (10.0%) 1* Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%)	Plasma D-dimer (ng/mL)			_	0.030
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Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome	Eculizumab, N (%)	6 (5.6%)	4 (4.1%)	2 (20.0%)	0.10*
Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome	Tocilizumab, N (%)	4 (3.7%)	4 (4.1%)	0 (0.0%)	1*
Severity during hospitalization and outcome	Dexamethasone, N (%)	53 (49.1%)	46 (46.9%)	7 (70.0%)	0.19*
	Dexamethasone cumulative dose >100mg, N (%)	16 (14.8%)	15 (15.3%)	1 (10.0%)	1*
SAPS II on admission 37 [31-49] 38 [31-51] 34 [28-37] 0.16	Severity during hospitalization and outcome		<u> </u>	<u> </u>	
	SAPS II on admission	37 [31-49]	38 [31-51]	34 [28-37]	0.160
SOFA on admission 6 [3-8] 6 [3-8] 5 [2-7] 0.43	SOFA on admission	6 [3-8]	6 [3-8]	5 [2-7]	0.430

Lowest PaO ₂ /FiO ₂ (mmHg)	71 [58-89]	71 [59-89]	65 [53-102]	0.790
Vasopressors, N (%)	89 (82.4%)	81 (82.7%)	8 (80.0%)	1*
Renal replacement therapy, N (%)	38 (35.2%)	35 (35.7%)	3 (30.0%)	1*
ECMO, N (%)	10 (9.3%)	9 (9.2%)	1 (10.0%)	1*
SAPS II on admission	37 [31-49]	38 [31-51]	34 [28-37]	0.160
ICU length of stay (days)	20 [12-32]	20 [12-33]	10 [6-19]	0.030
Mortality, N (%)	47 (43.5%)	44 (44.9%)	3 (30.0%)	0.51 *

143 BAL, bronchoalveolar lavage; ECMO, extracorporeal membrane oxygenation; ICU, intensive care unit; SOFA, Sepsis-

144 related Organ Failure Assessment; SAPS II, Simplified Acute Physiology Score II.

1 Table 1

Characteristics of hundred-and-eight critically ill COVID-19 patients according to *Pneumocystis jirovecii* detection in the respiratory samples. Data are presented as percentages or medians [25th-75th percentiles].

- 4 Comparisons were performed using Mann-Whitney (•) or Fisher exact tests (*), as appropriate.
- 5

	Total	No detection	Detection of	
	Total	of <i>P. jirovecii</i>	P. jirovecii	Р
	(N=108)	(N=98)	(N=10)	
Male gender, N (%)	88 (81.5%)	80 (81.6%)	8 (80.0%)	1*
Age (years)	62 [56-68]	62 [56-68]	59 [46-68]	0.400
COVID-19 risk factors		<u> </u>		
Past hypertension, N (%)	64 (59.3%)	58 (59.2%)	6 (60.0%)	1*
Diabetes, N (%)	40 (37.0%)	37 (37.8%)	3 (30.0%)	0.74*
Obesity, N (%)	35 (32.4%)	32 (32.7%)	3 (30.0%)	1*
Coronary disease, N (%)	15 (13.9%)	14 (14.3%)	1 (10.0%)	1*
Body-mass index (kg/m ²)	28 [25-31]	28 [25-31]	28 [27-32]	0.610
Other remarkable comorbidities				
Asthma, N (%)	5 (4.6%)	4 (4.1%)	1 (10.0%)	0.39*
Chronic obstructive pulmonary disease, N (%)	2 (1.9%)	2 (2.0%)	0 (0.0%)	1*
Immunocompromised patient, N (%)	10 (9.3%)	10 (10.2%)	0 (0.0%)	0.59*
Long-term corticosteroids, N (%)	11 (10.2%)	8 (8.2%)	3 (30.0%)	0.06*
Biological data of interest on admission	<u> </u>			
PaO ₂ /FiO ₂ (mmHg)	137 [83-247]	134 [83-239]	177 [108-253]	0.600

Plasma D-dimer (ng/mL) 2,2395 [1,193- 4,635] 2,610 [1,405- 4,700] 1,270 [750- 2,390] 0.03 Serum lactate dehydrogenase (1U/L) 687 [540-901] 687 [568-903] 708 [436-893] 0.65 BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL polymorphonuclear cells 37 [26-81] 46 [26-81] 29 [18-32] 0.54 % BAL Lymphocytes 13 [6-32] 14 [5-34] 13 [10-23] 0.81 Specific anti-COVID-19 therapy, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 3 (3.2%) 3 (3.1%) 0 (0.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 4 (3.7%) 4 (4.1%) 2 (20.0%)					
Plasma D-dimer (ng/mL) 4,635] 4,700] 2,390] 0.03 Serum lactate dehydrogenase (1U/L) 687 [540-901] 687 [568-903] 708 [436-893] 0.65 Bronchoalveolar lavage characteristics	Serum creatinine (µmol/L)	80 [64-111]	80 [63-111]	80 [67-104]	0.990
Bronchoalveolar layage characteristics % BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL polymorphonuclear cells 37 [26-81] 46 [26-81] 29 [18-32] 0.54 % BAL Lymphocytes 13 [6-32] 14 [5-34] 13 [10-23] 0.81 Specific anti-COVID-19 therapy, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Lopinavir-ritonavir, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 4 (3.7%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 53 (49.1%) 15 (15.3%) 1 (10.0%) 1* Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%)	Plasma D-dimer (ng/mL)			_	0.030
% BAL macrophages 28 [15-46] 27 [14-42] 51 [49-55] 0.13 % BAL polymorphonuclear cells 37 [26-81] 46 [26-81] 29 [18-32] 0.54 % BAL Lymphocytes 13 [6-32] 14 [5-34] 13 [10-23] 0.81 Specific anti-COVID-19 therapy, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine + Azithromycin, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1*	Serum lactate dehydrogenase (IU/L)	687 [540-901]	687 [568-903]	708 [436-893]	0.650
** BAL polymorphonuclear cells 37 [26-81] 46 [26-81] 29 [18-32] 0.54 ** BAL Lymphocytes 13 [6-32] 14 [5-34] 13 [10-23] 0.81 Specific anti-COVID-19 therapy, N (%) Azithromycin, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine + Azithromycin, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1*	Bronchoalveolar lavage characteristics				
************************************	% BAL macrophages	28 [15-46]	27 [14-42]	51 [49-55]	0.130
Specific anti-COVID-19 therapy, N (%) Azithromycin, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine + Azithromycin, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Dexamethasone, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome 10 15 (15.3%) 1 (10.0%) 1*	% BAL polymorphonuclear cells	37 [26-81]	46 [26-81]	29 [18-32]	0.540
Azithromycin, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine + Azithromycin, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome 1000000000000000000000000000000000000	% BAL Lymphocytes	13 [6-32]	14 [5-34]	13 [10-23]	0.810
Hydroxychloroquine, N (%) 34 (31.5%) 30 (30.6%) 4 (40.0%) 0.72 Hydroxychloroquine + Azithromycin, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1*	Specific anti-COVID-19 therapy, N (%)				
Hydroxychloroquine + Azithromycin, N (%) 29 (26.9%) 26 (26.5%) 3 (30.0%) 1* Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1*	Azithromycin, N (%)	34 (31.5%)	30 (30.6%)	4 (40.0%)	0.72*
Lopinavir-ritonavir, N (%) 16 (14.8%) 12 (12.2%) 4 (40.0%) 0.04 Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1*	Hydroxychloroquine, N (%)	34 (31.5%)	30 (30.6%)	4 (40.0%)	0.72*
Polyvalent immunoglobulins, N (%) 3 (2.8%) 3 (3.1%) 0 (0.0%) 1* Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome 15 (15.3%) 1 (10.0%) 1*	Hydroxychloroquine + Azithromycin, N (%)	29 (26.9%)	26 (26.5%)	3 (30.0%)	1*
Sarilumab, N (%) 1 (0.9%) 1 (1.0%) 0 (0.0%) 1* Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome 15 (15.3%) 1 (10.0%) 1*	Lopinavir-ritonavir, N (%)	16 (14.8%)	12 (12.2%)	4 (40.0%)	0.04*
Eculizumab, N (%) 6 (5.6%) 4 (4.1%) 2 (20.0%) 0.10 Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome	Polyvalent immunoglobulins, N (%)	3 (2.8%)	3 (3.1%)	0 (0.0%)	1*
Tocilizumab, N (%) 4 (3.7%) 4 (4.1%) 0 (0.0%) 1* Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome 15 (15.3%) 1 (10.0%) 1*	Sarilumab, N (%)	1 (0.9%)	1 (1.0%)	0 (0.0%)	1*
Dexamethasone, N (%) 53 (49.1%) 46 (46.9%) 7 (70.0%) 0.19 Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome	Eculizumab, N (%)	6 (5.6%)	4 (4.1%)	2 (20.0%)	0.10*
Dexamethasone cumulative dose >100mg, N (%) 16 (14.8%) 15 (15.3%) 1 (10.0%) 1* Severity during hospitalization and outcome	Tocilizumab, N (%)	4 (3.7%)	4 (4.1%)	0 (0.0%)	1*
Severity during hospitalization and outcome	Dexamethasone, N (%)	53 (49.1%)	46 (46.9%)	7 (70.0%)	0.19*
	Dexamethasone cumulative dose >100mg, N (%)	16 (14.8%)	15 (15.3%)	1 (10.0%)	1*
SAPS II on admission 37 [31-49] 38 [31-51] 34 [28-37] 0.16	Severity during hospitalization and outcome		<u> </u>	<u> </u>	
	SAPS II on admission	37 [31-49]	38 [31-51]	34 [28-37]	0.160
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ICU length of stay (days)	20 [12-32]	20 [12-33]	10 [6-19]	0.030
Mortality, N (%)	47 (43.5%)	44 (44.9%)	3 (30.0%)	0.51 *
				1

6 BAL, bronchoalveolar lavage; ECMO, extracorporeal membrane oxygenation; ICU, intensive care unit; SOFA, Sepsis-

7 related Organ Failure Assessment; SAPS II, Simplified Acute Physiology Score II.