SUPPLEMENTARY INFORMATION

ORF4 of the temperate archaeal virus SNJ1 governs the lysis-lysogeny switch and superinfection immunity

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Supplementary Figure 1. The titer of SNJ1 produced by the J7-1 strain with/without MMC treatment.

J7-1 was cultured in Halo-2 medium for 24 h and then treated with or without MMC for 30 min. Cells were washed with fresh Halo-2 medium to remove MMC and then cultivated for another 24 h. Supernatants were saved as virus stocks and titers were determined by double-layer plaque assay. Three independent experiments were performed, and error bars indicated standard deviations.



Supplementary Figure 2. Schematic map of the *E. coli-Natrinema* shuttle vector pYC-S.

pYC-S was obtained by ligation of *SacI* digested pUC19-mev into *SacI* digested pHH205 (proviral genome of SNJ1). Scheme of SNJ1 genome and pUC19-mev plasmid were shown. The putative ORFs of SNJ1 and pUC19-mev were indicated by arrows.



b



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CJ7

Supplementary Figure 3. Western blot of ORF4-GFP and functional test in superinfection immunity.

a. Samples from CJ7/pFJ6-PphaRP-GFP, CJ7/pFJ6-PphaRP-ORF4-GFP-1 and CJ7/pFJ6-PphaRP-ORF4-GFP-2 were collected at late-exponential phase and crude extracts were loaded for SDS-PAGE, lane 1 through lane 3 respectively. Anti-GFP antibodies was used for Western blot assay of GFP and ORF4-GFP fusion protein. b. Expression of ORF4-GFP fusion protein in CJ7 did not show resistance to SNJ1 infection. Ten-fold serial dilutions of SNJ1 viruses were spotted onto the lawns of CJ7, CJ7/pFJ6-PphaRP-ORF4-GFP-1 and CJ7/pFJ6-PphaRP-ORF4-GFP-2. Plates were incubated at 37°C for 48 h and photographed. The maximum dilution for observed plaques was highlighted by asterisk.

Supplementary Table 1: Strains used in this study

Strains	Description	Source and reference
Natrinema sp. J7-1 ^a	Lysogenic strain of SNJ1	1-3
Natrinema sp. CJ7	SNJ1 cured strain of J7-1	2,3
<i>Natrinema</i> sp. CJ7-F	$CJ7\Delta pyrF$ (orotidine 5'-phosphate decarboxylase, NJ7G_2091), uracil auxotrophic strain	4
Escherichia coli DH5α	For plasmids construction $supE44 \Delta lacU169(\phi 80 lacZ\Delta M15) hsdR17$ recA1 endA1 gyrA96 thi-1 relA1	CCTCC ^b
Escherichia coli JM110	For plasmids demethylation $dam \ dcm \ supE44 \ hsdR17 \ thi \ leu \ rpsL \ lacY \ galK$ $galT \ ara \ tonA \ thr \ tsx \ \Delta(lac-proAB) \ F'(traD36)$ $proAB^+lacI^q \ lacZ \ \Delta M15)$	CCTCC

a: GenBank Acc. No. AJVG00000000.1. b: CCTCC, China Center for Type Culture Collection.

Supplementary Table 2: Plasmids used in this study

Plasmids	Description	Source and
		reference
pUC19-mev	pUC19 with mevinolin resistance marker (mevR).	2
pYC-S	An E. coli-Natrinema shuttle vector based on SNJ1 genome,	2
	pUC19-mev was digested with SacI and inserted into SNJ1	
	proviral genome.	
pYC-S-4M	pYC-S with start codon mutated of <i>orf4</i> .	This study
pYC-S Δ1-575	pYC-S with 1-575 deleted within SNJ1.	This study
pYC-1	pUC19-Mev with the insertion of 5.9 kb SacI-BamHI 575-6,482	2
	fragment of SNJ1, used for pYC-S Δ 1-575 construction	
pYC-SHS	An E. coli-Natrinema shuttle vector based on SNJ1 replication	2
1	and regulation elements. pUC19-mev was digested with SacI-	
	HindIII and linked with SacI-HindIII digested SHS fragment (1-	
	6,482) of SNJ1.	
pYC-SHS-4M	pYC-SHS with start codon mutated of <i>orf4</i> .	2
pFJ6-MCS	An <i>E. coli-Natrinema</i> shuttle vector based on replication element	4
1	of CJ7 chromosome, containing pUC19, pyrF marker and	
	multiple cloning site.	
pFJ6-1-656	pFJ6-MCS with <i>MunI-SphI</i> fragment containing 1-656 of SNJ1	This study
pFJ6-1-656-	pFJ6-1-656 with frame-shift mutation of <i>orf3</i>	This study
3FSM		
pFJ6-1-656-	pFJ6-1-656 with frame-shift mutation of orf4	This study
4FSM		-
pFJ6-1-656-	pFJ6-1-656 with frame-shift mutation of orf3 and orf4	This study
3FSM 4FSM		
pYCJ-H	pUC19-Mev with the insertion of 4.5 kb SacI-SmaI 1-4,481	2
1	fragment of SNJ1 and heat-shock protein 70 promoter (Hpro)	
	from Haloferax volcanii DS52, used for pFJ6-Hpro construction	
pFJ6-Hpro	pFJ6-MCS with MunI-NotI fragment containing Hpro	This study
pFJ6-Hpro-orf4	pFJ6-Hpro with <i>Not</i> I-SphI fragment containing orf4 (499-293)	This study
pFJ6-Hpro-orf4-	pFJ6-Hpro- <i>orf4</i> containing frame-shift mutation of <i>orf4</i>	This study
4FSM		-
pFJ6-Hpro- <i>orf4</i> ^x	pFJ6-Hpro with NotI-SphI fragment containing a series of	This study
	truncation of <i>orf4</i> . ($x = 1-50$, 1-33, 1-24, 1-16, 1-8, 34-68)	
pFJ6-PphaRP-	pFJ6-MCS with AflII-SphI fragment containing promoter of the	This study
GFP	phaRP operon from Haloferax mediterranei and GFP	
pFJ6-PphaRP-	pFJ6-MCS with AflII-SphI fragment containing promoter of the	This study
ORF4-GFP-1	phaRP operon from Haloferax mediterranei and ORF4-GFP	
	fusion protein linked by A(EAAAK)*3A polypeptide.	
pFJ6-PphaRP-	pFJ6-MCS with AflII-SphI fragment containing promoter of the	This study
ORF4-GFP-2	phaRP operon from Haloferax mediterranei and ORF4-GFP	-
	fusion protein linked by (GGGGS)*3 polypeptide.	

Supplementary Table 3: Primers used in this study

Primers	sequences (5'-3')	Description
HJ-F	TGAATCAAGATGCCGCCGTG	primers used
		to check the
		genome
		integrity of
HJ-K	CGULICIGALCCACICG	the mutant
		viruses
		generated
		from pYC-S
pYC-S-	GTACCCTCTAGTCAAGGCCTTAAGTGAGTCGTATTACGGA	primers used
AflII-F	СТ	for pYC-S-
pYC-S-	CGCTAGCTAGGAGGTTTCCCATGGTTCCCGGCCGTCGGC	4M
NcoI-R	CCG	construction
4M-F	GTGCCATCTGAATCCTCTTACC	
4M-R	GGTAAGAGGATTCAGATGGCAC	1
SacI-1-F	AAAGAGCTCGCGATCGTTGACGTCGTC	primers used
Neel	TTCCCATCCTCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	for pYC-
1554 D		SHS-4M
1334-K		construction
SphI-1-F	AAAGCATGCGAGCTCGCGATCGTTGAC	primers used
		for pFJ6-1-
		656, pFJ6-1-
MunI-	AAACAATTGACACGTAGTATCGTATTATTGTTAG	656-3FSM,
656-		pFJ6-1-656-
ORF4-R		4FSM and
		pFJ6-1-656-
		3FSM 4FSM
		construction
ORF3-	GTCTAAGAGGCCATCGGTCA	primers used
FSM-F		for frame-
ORF3-	TGACCGATGGCCTCTTAGAC	shift
FSM-R		mutation of
		orf3
ORF4	TCAATATCAAGGCCATCTGCATCCT	primers used
FSM-F		for frame-
ORF4	GATGCAGATGGCCTTGATATTGATC	shift
FSM-R		mutation of
		orf4
MunI-	AAACAATTGTCTTCGCTGCCGCCGTTC	primers used
Hpro-F		for pFJ6-
NotI-	AAAGCGGCCGCCTTGCCCGCAAATACCGT	Hpro
Hpro-R		construction

NotI-	AATGCGGCCGCAATGCAGATGGCACTTGAT	primers used
ORF4-		for pFJ6-
499-F		Hpro- <i>orf4</i> ,
SphI-	AAAGCATGCCTACTCTTCGTCGGTACC	pFJ6-Hpro-
ORF4-		orf4-4FSM
293-R		and pFJ6-
		Hpro- <i>orf4</i> ^x
		construction
NotI-	AATGCGGCCGCAATGCAGATGGCCTTGATATTG	primer used
ORF4-		for pFJ6-
FSM-F		Hpro- <i>orf4</i> -
		4FSM
		construction
SphI-	TAAGCATGCCTAAACCATCTCAAGTTCGTCGTCTT	primers used
ORF4-		for pFJ6-
150-R		Hpro- <i>orf4</i> ^x
SphI-	TAAGCATGCCTAGGGCGGGATCGAGAGGATATTGCTA	construction
ORF4-		
99-R		
SphI-	AAAGCATGCTCACCCAGAGGACCGGATTGCGCGCT	
ORF4-		
72-R		
SphI-	AAAGCATGCTCATGCTTTCACCGTCGATTCATTCG	
ORF4-		
48-R		
SphI-	AAAGCATGCTCAATCAATATCAAGTGCCATCT	
ORF4-	GCAT	
24-R		
NotI-	AAAGCGGCCGCATGCAGATGATGCAGTTGA	
ORF4-		
400-F		
radA-F	ATCCACGTCGCGAAGGCGTTCA	primers used
radA-R	CTGCTTGTTGAGTTTCTGCTGTCGGTC	in qPCR
ORF14-F	GACAGATGTGAATAGCGGCGAAAG	assays
ORF14-	AGAAAGCGACGGAGACGATGGA	
R		
vector-F	GGGGATAACGCAGGAAAGAACA	
vector-R	GGCAGGGTCGGAACAGGAG	
ORF7-F	CGTTGAGGCGACGAAGCTCGGTGA	primers used
ORF11-	CCTCTTTCGTCAGCGTGTCCAAGCTCT	in southern
R		blot assay
ORF4-	GACGAACCCTTGCAATTGCTTAAGTAATCTCGTGGTATCT	primers used
GFP-1-F	СТ	for pFJ6-
ORF4-	ATCAAGTGCCATCTGCATTCAGGATCCCATCTCCTA	PphaRP-GFP

GFP-1-R		and	pFJ6-	
ORF4-	TAGGAGATGGGATCCTGAATGCAGATGGCACTTGAT	PphaR	P-	
GFP-2-F		ORF4-	GFP-	
ORF4-	GGCCTTGGCGGCGGCTTCCTTGGCGGCGGCTTCCTTGGCG	1/2		
GFP-2-R	GCGGCTTCGGCCTCTTCGTCGGTACCGGC	constru	uction	
ORF4-	GCCGAAGCCGCCGCCAAGGAAGCCGCCGCCAAGGAAGC			
GFP-3-F	CGCCGCCAAGGCCATGAGTAAAGGAGAAGAACTTTTCAC			
	TGGA			
ORF4-	ACCATGATTACGCCAAGCTTGCATGCTTATTTGTATAGTTC			
GFP-3-R	ATCCATGCCATGTGT			
ORF4-	GGCGGCGGTGGGTCGGGCGGCGGCGGCGGCGGCGG			
GFP-4-F	TGGGTCGATGAGTAAAGGAGAAGAACTTT			
ORF4-	AAAGTTCTTCTCCTTTACTCATCGACCCACCGCCGCCCGA			
GFP-4-R	CCCACCGCCGCCCGACCCACCGCCGCCCTCTTCGTCGGT			
	ACCGGCC			

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