

Dormancy in Cryptococcus neoformans: 60 years of accumulating evidence

Alexandre Alanio

▶ To cite this version:

Alexandre Alanio. Dormancy in Cryptococcus neoformans: 60 years of accumulating evidence. Journal of Clinical Investigation, 2020, pp.136223. 10.1172/jci136223. pasteur-02864599

HAL Id: pasteur-02864599 https://pasteur.hal.science/pasteur-02864599v1

Submitted on 11 Jun2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

1	
2	
3	Dormancy in Cryptococcus neoformans: sixty years of accumulating
4	evidence
5	
6	
7	
8	Alexandre Alanio ^{1,2,3}
9	
10	¹ Laboratoire de Parasitologie-Mycologie, Groupe Hospitalier Saint-Louis-Lariboisière-
11	Fernand-Widal, Assistance Publique-Hôpitaux de Paris (AP-HP), 75475 Paris, France
12	² Molecular Mycology Unit, Centre National de la Recherche Scientifique (CNRS) Unité
13	Mixte de Recherche 2000 (CNRS UMR2000), National Reference Center for Invasive
14	Mycoses and Antifungals (NRCMA), Institut Pasteur, 75724 Paris Cedex 15, France
15	³ Université de Paris, 75006 Paris, France
16	
17	Correspondance: Dr Alexandre Alanio, MD, PhD ; alexandre.alanio@pasteur.fr; Tel: +33-
18	1406-13255; Fax: +33-1456-88420
19	
20	
21	
_	

22

23 Abstract (125 words)

24

25 Cryptococcus neoformans is an opportunistic yeast that is present worldwide and interacts with various organisms. In humans, it is responsible for cryptococcosis, a 26 27 deadly invasive fungal infection which represents around 220,000 cases per year worldwide. Starting from the natural history of the disease in humans, there is 28 29 accumulating evidence on the capacity of this organism to enter dormancy. In response 30 to the harsh host environment, the yeast is able to adapt dramatically and escape the 31 vigilance of the host's immune cell to survive. Indeed, the yeast exposed to the host 32 takes on pleiotropic phenotypes, enabling the generation of populations in heterogenous 33 states, including dormancy, to eventually survive at low metabolic cost and revive in 34 favorable conditions. The concept of dormancy has been validated in C. neoformans 35 from both epidemiological and genotyping data, and more recently from the biological point of view with the characterization of dormancy through the description of viable but 36 non-culturable cells. 37

39 *Cryptococcus neoformans* is basidiomycetous opportunistic yeast that is widely present 40 in the environment. It causes human cryptococcosis, which mainly affects 41 immunocompromised patients and presents as a meningoencephalitis (1) that is lethal 42 without treatment. Clinical presentation is often diagnosed late because clinical 43 symptoms are initially mild with a sub-acute to chronic evolution (2).

44 Humans are exposed to *C. neoformans* from the environment. In nature, this fungus can survive the predation of various organisms ranging from protozoans to metazoans 45 46 through ready-made virulence traits (3). C. neoformans interacts closely with uni- or 47 multi-cellular organisms (2-4) and with cells dedicated to innate immune responses in metazoans (macrophages, dendritic cells, natural killer lymphocytes) with various 48 49 propensity to be phagocytosed and killed (4-6). C. neoformans is a facultative 50 intracellular pathogen (7). Interaction of C. neoformans with host cells can lead to phagocytosis, yeast replication within the phagolysosome, and is sometimes associated 51 52 with host cell lysis or with non-lytic exocytosis or cell-to-cell transfer and eventually killing 53 of the yeast (8–13). These phases have been well studied in different models of interaction 54 with host cells but mainly within macrophages. Indeed, intracellular persistence and 55 multiplication in immune cells provide advantages to the fungus by allowing escape from 56 the immune response and later dissemination through epithelial barriers (14, 15).

Characteristics of the infection depend on both hosts and microbial factors. Fungal 57 58 factors described as virulence factors influence the outcome of infection, according to 59 data obtained in the mouse model of cryptococcosis (16), but also in vitro (17) and in humans (8, 18). Microbial adaptation to the hosts is complex and has been studied 60 globally in lungs using histopathology (7) and global transcriptome analysis upon amoeba 61 (19) and macrophage ingestion (20) or upon early infection of mice and rabbits (21, 22). 62 63 Quiescence or dormancy is one of such adaptation that appears successful for enhancing the fungus's ability to survive, persist, reactivate, and then disseminate (23). 64

About sixty years of research focusing on how *C. neoformans* is able to cause infection in humans is available in literature, leading to the recent biological demonstration of dormancy in this organism. This review aims to summarize these sixty years of research, starting from the knowledge of human infection and ending with the characterization of dormancy biologically. This review is assembled to make the reader understand how this knowledge has been integrated to lead to more recent findings on the biology of *C. neoformans* characterizing dormancy, focusing on (i) The description of the natural history of *C. neoformans* infection in humans; (ii) The concept of dormancy in Fungi; (iii) Dormancy in *C. neoformans* in vivo and (iv) in vivo. To finish, a section is dedicated to discuss the (vi) relevance of the biological findings regarding human infection and to bring unsolved questions that can be the bases of future work in the field.

76 **1. Natural history of cryptococcosis in humans**

77

78 Cryptococcosis is one of the most frequent fungal invasive infection in humans 79 worldwide (24). The vast majority of patients with cryptococcosis are HIV-positive 80 patients, mostly those with CD4 T cells <100 cells per µL. Nevertheless, in western countries, the number of cryptococcosis cases recorded in HIV-negative individuals 81 becomes higher than that in HIV-positive patients (25). Immunocompromised HIV-82 negative patients at risk of cryptococcosis are mainly solid organ transplant recipients, 83 84 patients with systemic autoimmune disease, and those with hematological malignancies 85 (26).

The natural history of cryptococcosis is described following two main routes. The first one, although rare, occurs after exposure to *C. neoformans* while immunocompromised, leading to rapidly progressive cryptococcosis; the second one is reminiscent of tuberculosis, with a phase of latency with reactivation and dissemination. This second route appears to be the main mechanism of infection and so will be further developed in this review.

92

93 1.a. First route of infection: ready-made for disease

94

95 Confronted with the need to survive in nature and to survive different hosts in different 96 environments, *C. neoformans* has selected ready-made virulence traits (3). From a 97 deterministic point of view, *C. neoformans* population also needs diversity to survive 98 predators harboring different killing propensities. The plasticity of the *C. neoformans* 99 genome could lead to this diversity (27). *C. neoformans* and *C. gattii* are haploid 100 organisms that can be found as diploid organisms both in nature and in hosts (28, 29). 101 Generation of hybrids is possible between the varieties *grubii* (serotype A) and 102 *neoformans* (serotype D) (29, 30) but also between *C. neoformans* and *C. gattii*, again 103 illustrating this plasticity (31).

104 C. neoformans has long been associated with pigeon droppings (32). Indeed, pigeon fanciers are known to have higher anti-C. neoformans antibodies than control individuals 105 106 (33). The presence of *C. neoformans* in human dwellings was a risk factor (odds ratio = 107 2.05) for the development of cryptococcosis in HIV-positive patients from Brazil (34). C. 108 gattii has also been found in indoor environments in Brazil, although links with human cryptococcosis have not been demonstrated (35). Several cases of cryptococcosis have 109 110 been reported in immunosuppressed patients in contact with birds (pigeons, parrot, 111 cockatoo, cockatiel) (36–39). The presence of C. neoformans in the feces of some animals 112 have been also observed in zoo animals (Guizotia abyssinica, Palm Cockatoo, Military 113 Macaw, Gray Parrot) (40). Nosocomial cases of cryptococcosis acquired in various 114 hospital settings have also been suspected (41, 42). Transmission of C. neoformans 115 through transplanted deep organs from a contaminated donor has occurred (43, 44) with, 116 in some cases, the demonstration of the same strain in different patients transplanted 117 with organs from the same donor (45).

Primary cryptococcosis initiates with lung involvement and then disseminates from the lung in immunocompromised hosts. Primary pulmonary cryptocococcosis is observed in immunocompetent and immunocompromised hosts. It can be recognized within a broad range of presentation, from isolated asymptomatic nodules that can mimic cancer lesions to more disseminated lesions of the lung with respiratory failure (46–48). Primary cutaneous cryptococcosis is also a clinical entity that happens after environmental inoculation in immunocompetent or immunocompromised hosts (49, 50).

- 125
- 126

127 1.b. Second route of infection: ready-made for latency

128

The majority of the infections arise from a natural history of the infection following 3 steps: primary infection in childhood and immune control, followed by a silent phase of latency that can last for years, and finally, reactivation and dissemination that are responsible for the symptoms of the disease mainly occurring upon immunosuppression.

133

134 Early environmental exposure

Inhalation of aerosolized particles from soil (desiccated yeasts or basidiospores) is 135 thought to be the major route of infection in humans (51). Primary infection with C. 136 neoformans occurs mainly in immunocompetent children as demonstrated by serological 137 studies with unrecognized (asymptomatic) infection as the main clinical presentation. The 138 139 proportion of children immunized against C. neoformans increases with age. Acquisition 140 of cryptococcal antibodies begins very early (1 year) with minimal reactivity of the sera. After 5 years, 70% of children react with C. neoformans antigens (52). However, the 141 142 acquisition of anti-cryptococcal humoral immunity varies among geographic areas. 143 Cryptococcal antibodies are very common in Bronx children but not in another New York 144 areas (Dutchess County), nor are they common in Manila (The Philippines), another 145 densely populated urban area (53). Environmental exposure may depend on climatic and environmental factors (temperature, humidity, pigeon density), but also on human 146 sociological factors (habitat conditions, financial resources). These findings support 147 148 epidemiological data revealing that cryptococcosis in immunocompromised individuals 149 is more prevalent in some areas of the world, especially in Africa (24, 54).

150 Of note, *C. gattii* exposure and primoinfection does not follow the same epidemiological 151 trends than *C. neoformans* based on studies realized in endemic areas in animals and 152 humans (55, 56).

153

154 Latency

155 Serologic evidence of early cryptococcal immunity in immunocompetent hosts without 156 recognized infection seems paradoxical considering the very low frequency of 157 cryptococcosis in immunocompetent hosts. However, immune control of the yeasts by 158 immunocompetent hosts following primoinfection is possible, with latency of the disease 159 or complete clearance of the fungus as a consequence. Immunocompetent adults 160 frequently exposed to *C. neoformans* had positive skin test but did not develop clinical 161 disease (57). Autopsy studies have raised the hypothesis that pulmonary granulomas 162 could be the site for persistence because C. neoformans is observed in sub-pleural 163 nodules and draining lymph nodes in immunocompetent and immunocompromised 164 hosts (58). Indeed, several reports showing that C. neoformans lymphadenitis have been exclusively found and isolated from lymph node can be found in literature, thus arguing 165 166 that initial immune control of the yeasts operates in lymph nodes (59-65). From recent 167 and old reports, the lymph nodes associated with lymphadenitis correspond to granuloma composed of epitheloïd cells, Giant cells, and necrosis surrounded by a T cell 168 169 infiltrate together with yeasts (64, 66, 67).

170 Analysis of clinical isolates of *C. neoformans* var. grubii recovered in France from patients 171 born in Africa (who moved to France with a median of 110 months elapsing before 172 isolation of the yeast in France), revealed that yeast genotypes from these patients 173 clustered together, distinct from the yeast genotypes recovered from patients born in 174 Europe (68). This study is the main epidemiological evidence for this latency stage of the 175 disease. This latency can be translated into the capacity of dormancy of the yeasts, which 176 appears to be the more plausible explanation from the point of view of the biology of the 177 organism. The same conclusion is also drawn from a serologic survey of solid organs transplant recipients (immunocompromised hosts). Interestingly, sera obtained before 178 179 and after transplantation from transplanted patients with cryptococcosis was compared 180 to control transplanted patients without history of cryptococcosis. Among patients with 181 cryptococcosis, half exhibited antibody reactivity against C. neoformans only after 182 transplantation. This suggests that this half of the patients were exposed and developed 183 the disease after transplantation during immunosuppression. But for the other half of the 184 patients, antibody reactivity against *C. neoformans* was found before transplantation; 185 these patients' early development of cryptococcosis after transplantation suggests that 186 reactivation and dissemination occur rapidly after transplantation from a preexisting isolate in transplant recipient, thus validating again the latency phenomenon (69).
Additionally, report of *C. gattii* infections in patients who travelled to endemic areas years
or months prior to the Vancouver Island *C. gattii* outbreak provides more evidence for
latency (70).

191

192 <u>Reactivation</u>

193 The first manifestation of reactivation is observed in HIV-infected individuals in whom 194 asymptomatic cryptococcal antigenemia is detected (71-73). Viable yeasts are not 195 recovered from clinical sample at this step but treatment is mandatory to prevent symptoms and dissemination (74, 75). Pulmonary cryptococcosis is a well-described 196 197 clinical entity that can evolve differentially depending on the immune status of the hosts. 198 In immunocompetent hosts, C. neoformans does not usually disseminate, whereas the 199 possibility of dissemination in immunocompromised patients is high. It is likely that 200 dissemination occurs after reactivation of lung-persistent yeasts, crossing the lung epithelial barrier and disseminating through capillary blood (76, 77). However, abnormal 201 202 chest X-ray or CT scan was observed in 39% of HIV-positive patients and 55% of HIV-203 negative patients at diagnosis, although dissemination represented 60.6% and 38.5% of 204 the cases, respectively (1). However, pulmonary symptoms are not the main clinical 205 manifestation of cryptococcosis in immunocompromised patients. Indeed, most are 206 diagnosed at the stage of dissemination or meningoencephalitis (1). Cryptococcosis is 207 characterized by a high frequency of central nervous system (CNS) involvement with 208 positive cerebrospinal fluid (CSF) and dissemination through blood. Cryptococcosis is more severe in male HIV-positive patients and those infected with C. neoformans 209 210 serotype A (1). Acute cryptococcal meningoencephalitis (CM) is always fatal without antifungal therapy (78). Treatment of CM requires an antifungal therapy induction based 211 212 on amphotericin B and flucytosine (79). Based on recent large clinical trials in African 213 settings, one-week amphotericin B combined with oral flucytosine followed by high-dose 214 fluconazole is now recognized as the reference therapy (75, 80). Mycological failure after 2 weeks of induction is recognized as a factor of bad prognosis, which requires 215 216 continuation of the induction therapy (79). Mycological failure is independently

217 associated with initial dissemination, high serum antigenemia (>1:512), and lack of initial 218 flucytosine treatment (1, 80, 81). The three-month mortality rate during the management of acute cryptococcal meningoencephalitis approximates 15-20% in western countries 219 220 despite adequate treatment and management. It is still not clear whether this mortality 221 rate is due to individual's immune status, genetic factors (82), fungal determinants, or a 222 combination of these. Nevertheless, two reports clearly identified that fungal 223 determinants specific to the strain are responsible for a given phenotype of interaction 224 with host cells (high phagocytosis, high intracellular proliferation) that is associated with 225 mortality in patients (8, 83)

- 226
- 227

2. The concept of dormancy in Fungi

228

All microorganisms are exposed to periodic constraint conditions and react by inhibiting 229 230 their growth, entering into a non-replicative state called quiescence or dormancy (84, 85). Three main strategies can be delineated in these conditions. The first is the "bust and 231 232 bloom" strategy (85), where the microorganism population will grow rapidly with growth 233 maximization, but upon nutrient exhaustion, the majority of the individuals will die, with 234 only few cells surviving. These residual cells will resume growth rapidly upon exposure 235 to nutrient (86). The second strategy is "guiescence," where the bulk of the population 236 exposed to nutrient-limited environment will arrest or slow growth to enter a viable, non-237 replicating state for a long time. This can last month or years for Mycobacterium 238 tuberculosis (87). These cells keep a baseline and specific metabolic capacity, maintain 239 their membrane potential, and do not undergo major morphological change (88). The third strategy is called "true dormancy," with sporulation as the purest form, in which an 240 241 asymmetrical replication leads to the formation of a metabolically inactive spore (89). The 242 spore harbors specific morphology but shares some biological features with guiescent 243 cells.

244 Quiescence in S. cerevisiae has been studied for a long time. Recently, a strain of S. cerevisiae has been found "alive" in bottles of beer and Champagne from the 18th 245 246 century found in a shipwreck in the Baltic sea suggesting this phenomenon can last for 247 years in specific conditions. Quiescent yeasts are mainly obtained from cultures grown 248 to saturation in glucose-rich media (stationary phase) where all nutrients have been 249 consumed. Different phases have been described, including (i) a first phase of glycogen 250 production upon rarefaction of glucose (at about 50%) (90), followed by (ii) the regulation 251 of trehalose before and after glucose exhaustion. Then, (iii) the yeasts undergo a phase 252 of diauxic shift (following glucose depletion) where growth is slow and metabolism is 253 adapted to limitation of nutrients, relying on respiratory growth of non-fermentable 254 sugars such as ethanol or acetate with switch towards respiration, fatty acid pathway, 255 and glyoxylate cycle pathway and, as a consequence, increased formation of antioxidant 256 defenses (scavenging of ROS) (91). The yeast population obtained in stationary phase is 257 described as a heterogenous population including guiescent cells (composed of 258 daughter and young mother cells) but also non-quiescent cells, which lose their ability to 259 accumulate ROS, exhibit genomic instability, and become senescent or apoptotic (92) 260 In C. neoformans, growth arrest in G1 or G2 period has been demonstrated in stationary 261 phase (93). No specific morphological differences in the mitochondrial apparatus was observed in logarithmic versus stationary phase (Figure 1) (94). No comprehensive 262 263 analysis on the metabolism of C. neoformans in stationary phase compared to 264 logarithmic phase existed until recently, as part of the investigation of a specific 265 phenotype observed upon exposure to drastic conditions (95).

266

267 **3. Dormancy in** *C. neoformans* in vivo

268

269 The body of evidence for dormancy elaborates on various parameters, mainly including 270 viability, which should not be based on culturability, reactivation upon specific stimuli, 271 and specific biological activity. Viability requires the use and adaptation of tools available 272 to test viability/death in mammalian cells (23). For a long time, viability and its corollary 273 (killing or death of C. neoformans) was investigated using CFU counting (96). Other means to assess the viability/death of yeast have now been developed, including the use 274 275 of intercalating dyes such as propidium iodide that is able to diffuse and stain the DNA 276 of the yeasts only if the extracellular membrane lose integrity (97) This method allows 277 assessment of viability or death by using flow cytometry. Other intercalating dyes can be 278 used with the same principle (23, 98). These methods assume that a dead yeast cell will lose membrane integrity, which is potentially not necessary at first. Apoptosis should also 279 280 be checked in the context to determine if the cell is oriented towards cell-death or will 281 remain viable. The existence of apoptosis in fungi is debated (99), but evidence exists for 282 the presence of caspase-like proteins in C. neoformans (100) that could act as effectors 283 of mechanisms related to caspase-dependent cell death. Nevertheless, apoptosis in 284 Fungi cannot be directly equated to what is known in mammalian cells (99, 101).

285 In C. neoformans' stationary phase, it was shown that only a small proportion of the population was unable to grow and considered dead (23). From *in vivo* experiments and 286 287 interaction with macrophages, it has been shown that yeast cells were able to keep their 288 round shape and capsule, although dead as shown with different means (23). These dead 289 *C. neoformans* yeast cells have been called DropCn due to the presence of a large central 290 vesicle inside the cell. The cell wall was shown to be thicker that stationary phase yeasts 291 (Figure 1). In those dead cells, the intracellular content is collapsed around vesicles 292 including remaining membranes (stained with MDY64), nucleic acids (stained with 293 SYTO85), but with no organized nucleus (negative DAPI staining) and no mitochondria 294 (negative Mitotracker staining) (23). These cells were able to retain a CMFDA staining 295 (glutathione staining) in their remaining capsule and cell wall, which was supposed to be 296 intracellular, producing fluorescence artefacts, allowing detection despite being dead. To 297 prevent such bias, multispectral imaging flow cytometry was used, allowing observation 298 of the fluorescence within the cells to assess location (23). Apart from those dead cells, this study highlighted that heterogenous population of yeasts were generated in the lung 299 300 of infected mice and upon macrophage interaction. Indeed, the view of the existence of 301 homogenous population of yeasts in specific conditions turns out to be inadequate and 302 raise the question of the accuracy of studies dealing with global analysis of the global 303 population of yeasts recovered in specific settings. Nevertheless, global transcription 304 analyses supported the idea of fungal adaptation to hostile environments such as the 305 macrophage phagolysosome (19, 20, 102), inside amoeba (19), in the lung during murine

infection (21), in the central nervous system of rabbit (22), or in human cerebrospinal fluid(103).

Heterogenous populations generated during murine infection included (i) active yeasts 308 309 able to bud and multiply, (ii) dead yeasts, and also (iii) a population of more dormant 310 yeasts. These dormant yeasts were less prone to grow as compared to stationary phase, 311 which is already considered as a state where almost of the yeasts are quiescent. This 312 explain why these cells have been called dormant instead of guiescent cells (23). These cells had also a decreased response to stress (low glutathione production), increased 313 mitochondrial expression, increased autophagy, and decreased gluconeogenesis-314 associated transcriptional activity (23). 315

316 **4. Dormancy in** *C. neoformans* studied in vitro

317

318 In the previous study using the mouse model, as few as 10⁴ dormant yeast cells were 319 able to be generated after pooling several mice lungs, which is obviously insufficient to 320 study basic biological processes allowing the characterization of dormancy. Therefore, 321 the authors worked on an in vitro model to be able to generate a high number of dormant 322 yeast cells. Recently, the authors released and studied the standardized conditions 323 allowing the generation of yeast cells harboring a phenotype close to that dormant cells 324 generated in the lungs of infected mice (95). These conditions are based on a 325 combination of conditions (low oxygen and limited nutrients) inspired from the Wayne 326 and the Loebel models (two well-documented conditions enabling the generation of 327 quiescent *M. tuberculosis*) (104). After stationary phase in YPD and exposure to anaerobiosis and nutrient starvation during 8 days, the authors observed that 95% of 328 329 yeast cells were viable, with few dead cells. They demonstrated that cells were not 330 apoptotic upon TUNEL staining. Overtime, these yeast cells showed a decreased 331 culturability on YPD agar plates, ending with about 1% of the cells still able to grow on agar at Day 8 of incubation. The phenotype observed in the in vivo subpopulation was 332 333 resumed with delayed growth (increased latency) and low stress response (95). In total, 334 the population obtained was homogenously composed of cells characterized as Viable

335 but non-Culturable cells (VBNC) (Figure 1), phenotype well-known in many bacteria and 336 first described in 1982 in *Escherichia coli* (105). Among fungi, this phenotype has been described in S. cerevisiae (106), Candida stellata (107) and in Brettanomyces bruxellensis 337 grown in wine synthetic medium and induced by sulfur dioxide (108). C. neoformans 338 339 VBNCs were induced by hypoxia and nutrient deprivation and a proportion of them were able to be reactivated by the vitamin B5 (panthotenic acid) with a doubling number of 340 341 culturable cells (Figure 2). Of note, it has been shown is specific model in *E. coli* that the VBNCs were potentially unable to reactivate (109). Pantothenic acid is known to play a 342 role in the process of division (cell cycle) and in the quorum-sensing phenomenon (110) 343 The use of diluted medium (which is poor in nutrients) to try to reactivate VBNCs was 344 345 attempted, reflecting the observation that rich medium can be deleterious and induce death (111). Diluted medium did not lead to reactivation of more cells than rich medium, 346 347 but rather, the cells that did reactivate exhibited faster growth and an increased doubling time compared to rich media. This cell phenotype induced by diluted medium has been 348 349 called rewiring (95). Finally, based on large omics studies, the authors of this study were 350 able to show that C. neoformans VBNCs harbored a decreased and specific metabolic 351 activity based on phenotypic microarrays, transcriptome, secretome, and proteome 352 analysis (95). Specifically, the fatty acid pathway was required for the maintenance and 353 the viability of the VBNCs, and quorum sensing and mTor pathways seemed to play an 354 important role in generating and/or maintaining the phenotype. Interestingly, acetyl CoA 355 is a key precursor for both fatty acids and pantothenic acid, suggesting that regulation of acetyl CoA is a major factor for the generation of VBNCs (112). Based on these 356 357 findings, a basic model of the evolution of *C. neoformans* yeast cells from logarithmic 358 phase to dormancy and dead cells can be summarized as depicted in Figure 2.

An analysis of the bulk population of yeasts maintained 8-days in nutrient deprivation and anaerobiosis to generate VBNCs identified another sub-population of yeasts that are still able and ready to grow on agar-rich medium. This population can be considered as persister cells. Persister cells have been described in a population of bacteria exposed to fungicidal antibiotics as the small proportion of bacteria able to tolerate spontaneously and stochastically lytic drugs via different mechanisms (113, 114). It has been shown that 365 persisters and VBNCs can coexist in specific model of study in the bacteria Vibrio 366 vulnificus (115). Persister cells have been described in *C. albicans* biofilm (116) and seem to play a role in recurrent infection in human oral candidiasis associated to natural biofilm 367 (117). In VBNC-inducing conditions of *C. neoformans*, one can consider that remaining 368 369 cells able to enter grow rapidly after long exposure of harsh conditions could be related 370 to such persister cells, since a specific metabolism seems to occur compared to that of the VBNCs, which need a specific stimulus to grow again. This needs to be studied in 371 372 details in future research.

373

374 **5. Relevance and unsolved questions**

375

376 The recent study highlighting the capacity of *C. neoformans* to switch to VBNCs can be 377 viewed as a model to explore dormancy and metabolism in this organism and in 378 pathogenic fungi in general. Indeed, this phenotype have still not been evidenced per se in human infection yet, but experimental conditions and the number of yeasts needed to 379 380 obtain the demonstration is clearly not compatible with what can be recovered from the 381 CSF of a patient with CM. The question of observing yeasts in the CSF of patients after 382 7 days of induction therapy with a negative culture on regular medium obviously raise 383 the question whether these yeasts are VBNCs. Being involved in clinical diagnosis, I have 384 observed that the morphology of these non-culturable yeasts is abnormal and close to 385 that observed in murine infection and called Drop Cn (see above) (8). Indeed, dead yeasts are known to persist and keep their intact shape, although different stainings can help 386 387 differentiating them from regular and living yeasts (8). VBNCs, or at least part of the VBNC 388 population, have proven to be reactivable by pantothenic acid, part of the demonstration that these cells are VBNC. The mechanism behind the specific reactivation is yet to be 389 390 elucidated, but the fact that pantothenic acid (Vitamin B5) is a precursor of coenzyme A, 391 which is an essential compound that participates in the metabolism of fatty acids, 392 carbohydrates, and proteins through the formation of various active thioesters and 393 promotes virulence and growth (118). Indeed, fatty acid have been shown to be critical 394 in VBNCs (95). VBNCs obtained in *C. neoformans* can be considered as similar to that 395 obtained from bacteria or parasites as the definition relies on viability, culturability, and 396 reactivation upon specific stimuli. Nevertheless, the condition allowing the generation of 397 VBNCs and the stimuli that reactivate the population are different in different organisms. 398 Among organisms, many common conditions of induction rely on stresses including 399 starvation, low oxygen, low temperature, desiccation, or a combination of these. On the other hand, resuscitation conditions are extremely variable, such as increased nutrient 400 401 availability, temperature modifications, addition of chemicals, or addition of hosts factors, 402 depending on the organism (119).

403 The biology of dormancy in C. neoformans is a budding field and yet there are many more 404 questions than answers. We still lack data on the effect of antifungal drugs on dormant 405 yeasts cells because the experimental setting allowing demonstration of the effect or the 406 absence of effect is not easy to implement in dormant cells that are intrinsically not 407 cultivable. Indeed, turning on dormancy with some VBNC inductors that remain to be 408 discovered would definitely aid in treating acute infection. On the other hand, inducing 409 VBNCs could also be the cause of relapse by producing insensitivity to current antifungal 410 strategies. These factors need to be addressed. Moreover, we have no data yet on the possible extension of the VBNC phenotype to clinical isolates of C. neoformans type VNI 411 412 and to other phylogenetic lineages or species. There is a chance that all clinical isolates could have varying propensities to generate VBNCs, and so the impact on infection could 413 be variable, as already shown for phenotypes of interaction with macrophages (8, 83). 414 415 We are currently exploring the effect of the host on the induction and maintenance of 416 VBNCs with regards to the level of activation of primary monocytes. One important 417 aspect we are also currently exploring is the impact of VBNC metabolism on the physiology of the macrophages. Both studies aim to understand the interplay between 418 419 host and fungal metabolisms, opening the way to discover specific pathways that could 420 be modulated to push the system in one or the other direction (more killing or less 421 proliferation of yeasts).

- 422
- 423
- 424 **6. Conclusions**

425 To summarize, C. neoformans is able to adapt fantastically to various environments, 426 some of which are very drastic, such as 8 days in complete anaerobiosis and without 427 extracellular nutrients available. C. neoformans uses strategies to resist these conditions. It is first perfectly able to enter quiescence in nutrient starvation conditions (stationary 428 429 phase) or to be pushed into dormancy under additional anaerobiosis exposure. In vivo, 430 one can imagine that VBNCs/dormant yeasts are most likely hidden in the innate immune 431 cells for years before being able to reactivate and multiply either in the body of 432 immunocompromised patients but also in the environment. This makes C. neoformans 433 the first relevant pathogenic organism in which to study fungal dormancy and its role in 434 the pathogenesis in humans.

435

436 **Acknowledgments:**

I would like to thank Françoise Dromer for her helpful comments and support and
Stéphane Bretagne and Arturo Casadevall for their continuous support. No funding
sources supported this work. In memory of my father for whom the question of "in
between life and death" is not relevant anymore.

441

442 **Competing interests:** The author has declared that no competing interests exist.

Financial declaration: AA received a grant (102k€) from the ANR in 2019 as a partner in
 the French-Germany call to fight against antimicrobial resistance. AA owns a patent

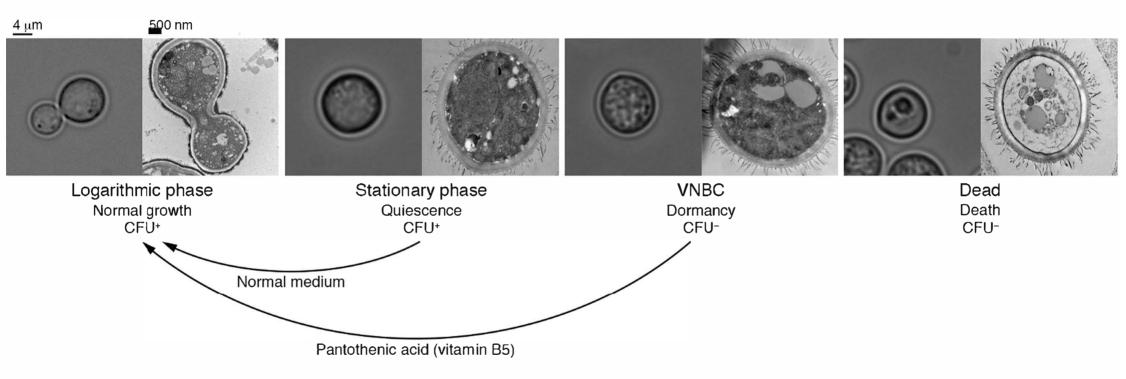
- 445 outside of the scope of this review on Pneumocystis diagnosis
- 446

- **Figures legends:**

Figure 1: Morphology of quiescent (Stationary phase), dormant and dead *C. neoformans* yeasts. The reference strain H99 was used in all conditions. STAT:
stationary phase (Yeast Peptone Dextrose, YPD, 22 hours with agitation 150 rpm) (17);
VBNC (after incubation 8 days in anaerobiosis and nutrient deprivation) (97); DEAD:
morphology of dead cells called DropCn including one or two vacuoles (17).

Figure 2: Schematic representation of the evolution of *C. neoformans* phenotypes upon incubation in nutrient deprivation and anaerobiosis. Yeasts cells under agitation and in glucose rich medium (YPD) are actively multiplying in logarithmic phase (LOG). Quiescent yeasts are culturable (Stationary phase, STAT) and does not need specific stimuli to grow in normal glucose rich (YPD) medium. Dormant yeasts (VBNC) are not culturable spontaneously and need a trigger stimulus for reactivation (addition of pantothenic acid). Dead yeasts (DEAD) are non-reversely unable to grow again (from 17, 97).





466 **Reference**

467

1. Dromer F, Mathoulin-Pélissier S, Launay O, Lortholary O. Determinants of Disease
Presentation and Outcome during Cryptococcosis: The CryptoA/D Study. *PLoS Med*2007;4(2):e21.

- 471 2. Perfect JR, Lang SD, Durack DT. Chronic cryptococcal meningitis: a new experimental model 472 in rabbits. *The American journal of pathology* 1980;101(1):177 194.
- 473 3. Casadevall A, Steenbergen JN, Nosanchuk JD. 'Ready made' virulence and 'dual use'
 474 virulence factors in pathogenic environmental fungi the *Cryptococcus neoformans* paradigm.
 475 *Curr Opin Microbiol* 2003;6(4):332–337.
- 476 4. Wozniak KL, Levitz SM. *Cryptococcus neoformans* Enters the Endolysosomal Pathway of 477 Dendritic Cells and Is Killed by Lysosomal Components. *Infect Immun* 2008;76(10):4764–4771.
- 478 5. Murphy JW, Zhou A, Wong SC. Direct interactions of human natural killer cells with
 479 *Cryptococcus neoformans* inhibit granulocyte-macrophage colony-stimulating factor and tumor
 480 necrosis factor alpha production. *Infection and Immunity* 1997;65(11):4564 4571.
- 481 6. Fries BC, Taborda CP, Serfass E, Casadevall A. Phenotypic switching of *Cryptococcus* 482 *neoformans* occurs in vivo and influences the outcome of infection. *J Clin Invest* 483 2001;108(11):1639–1648.
- 7. Feldmesser M, Kress Y, Novikoff P, Casadevall A. *Cryptococcus neoformans* is a facultative
 intracellular pathogen in murine pulmonary infection. *Infection and Immunity* 2000;68(7):4225
 4237.
- 487 8. Alanio A, Desnos-Ollivier M, Dromer F. Dynamics of *Cryptococcus neoformans*-Macrophage
 488 Interactions Reveal that Fungal Background Influences Outcome during Cryptococcal
 489 Meningoencephalitis in Humans. *mBio* 2011;2(4):e00158-11.
- 490 9. Alvarez M, Casadevall A. Cell-to-cell spread and massive vacuole formation after 491 *Cryptococcus neoformans* infection of murine macrophages. *BMC Immunol* 2007;8(1):16.
- 492 10. Alvarez M, Casadevall A. Phagosome Extrusion and Host-Cell Survival after *Cryptococcus* 493 *neoformans* Phagocytosis by Macrophages. *Curr Biol* 2006;16(21):2161 2165.
- 494 11. Johnston SA, May RC. The Human Fungal Pathogen *Cryptococcus neoformans* Escapes
 495 Macrophages by a Phagosome Emptying Mechanism That Is Inhibited by Arp2/3 Complex496 Mediated Actin Polymerisation. *PLoS Pathog* 2010;6(8):e1001041.
- 497 12. Dragotakes Q, Fu MS, Casadevall A. Dragotcytosis: Elucidation of the Mechanism for
 498 *Cryptococcus neoformans* Macrophage-to-Macrophage Transfer. *J Immunol* 2019;202(9):2661–
 499 2670.
- 500 13. Levitz SM et al. *Cryptococcus neoformans* resides in an acidic phagolysosome of human 501 macrophages. *Infect Immun* 1999;67(2):885–90.

- 502 14. Sorrell TC et al. Cryptococcal transmigration across a model brain blood-barrier: evidence of
- the Trojan horse mechanism and differences between *Cryptococcus neoformans* var. *grubii* strain
- 504 H99 and *Cryptococcus gattii* strain R265. *Microbes Infect* 2016;18(1):57–67.
- 505 15. Charlier C et al. Evidence of a Role for Monocytes in Dissemination and Brain Invasion by 506 *Cryptococcus neoformans. Infect Immun* 2009;77(1):120–127.
- 507 16. Chang YC, Kwon-Chung KJ. Complementation of a capsule-deficient mutation of 508 *Cryptococcus neoformans* restores its virulence. *Molecular and cellular biology* 1994;14(7):4912 509 4919.
- 510 17. Ma H et al. The fatal fungal outbreak on Vancouver Island is characterized by enhanced 511 intracellular parasitism driven by mitochondrial regulation. *Proc National Acad Sci* 512 2009;106(31):12980–12985.
- 18. Mansour MK, Vyas JM, Levitz SM. Dynamic Virulence: Real-Time Assessment of Intracellular
 Pathogenesis Links *Cryptococcus neoformans* Phenotype with Clinical Outcome. *mBio*2011;2(5):e00217-11.
- 516 19. Derengowski L da S et al. The Transcriptional Response of *Cryptococcus neoformans* to
 517 Ingestion by Acanthamoeba castellanii and Macrophages Provides Insights into the Evolutionary
 518 Adaptation to the Mammalian Host. *Eukaryot Cell* 2013;12(5):761–774.
- 519 20. Fan W, Kraus PR, Boily M-J, Heitman J. *Cryptococcus neoformans* Gene Expression during
 520 Murine Macrophage Infection. *Eukaryot Cell* 2005;4(8):1420–1433.
- 521 21. Hu G, Cheng P, Sham A, Perfect JR, Kronstad JW. Metabolic adaptation in *Cryptococcus* 522 *neoformans* during early murine pulmonary infection. *Mol Microbiol* 2008;69(6):1456–1475.
- 523 22. Steen BR et al. *Cryptococcus neoformans* Gene Expression during Experimental 524 Cryptococcal Meningitis. *Eukaryot Cell* 2003;2(6):1336–1349.
- 525 23. Alanio A, Vernel-Pauillac F, Sturny-Leclère A, Dromer F. *Cryptococcus neoformans* Host 526 Adaptation: Toward Biological Evidence of Dormancy. *mBio* 2015;6(2):e02580-14.
- 527 24. Rajasingham R et al. Global burden of disease of HIV-associated cryptococcal meningitis: an
 528 updated analysis. *Lancet Infect Dis* 2017;17(8):873–881.
- 529 25. Bitar D et al. Population-Based Analysis of Invasive Fungal Infections, France, 2001–2010.
 530 *Emerg Infect Dis* 2014;20(7):1163–1169.
- 531 26. Marr KA et al. A Multicenter, Longitudinal Cohort Study of Cryptococcosis in Human
 532 Immunodeficiency Virus-negative People in the United States. *Clin Infect Dis* 2019;70(2):252–261
- 27. Rhodes J et al. Tracing Genetic Exchange and Biogeography of *Cryptococcus neoformans*var. *grubii* at the Global Population Level. *Genetics* 2017;207(1):327–346.
- 535 28. Desnos-Ollivier M et al. Mixed Infections and In Vivo Evolution in the Human Fungal Pathogen 536 *Cryptococcus neoformans. mBio* 2010;1(1):e00091-10.

- 537 29. Xu J, Mitchell TG. Comparative gene genealogical analyses of strains of serotype AD identify 538 recombination in populations of serotypes A and D in the human pathogenic yeast *Cryptococcus*
- 539 *neoformans* [Internet]. *Microbiology* 2003;149:2147 2154.
- 540 30. Xu J, Vilgalys R, Mitchell TG. Multiple gene genealogies reveal recent dispersion and 541 hybridization in the human pathogenic fungus *Cryptococcus neoformans*. *Molecular ecology* 542 2000;9(10):1471 1481.
- 543 31. Bovers M et al. Unique hybrids between the fungal pathogens *Cryptococcus neoformans* and 544 *Cryptococcus gattii. Fems Yeast Res* 2006;6(4):599 607.
- 545 32. Emmons CW. Isolation of *Cryptococcus neoformans* from soil. *Journal of Bacteriology* 546 1951;62(6):685 690.
- 547 33. Fink JN, Barboriak JJ, Kaufman L. Cryptococcal antibodies in pigeon breeders' disease. *The* 548 *Journal of allergy* 1968;41(5):297 301.
- 549 34. Passoni LFC, Wanke B,Nishikawa MM, Lazera MS. *Cryptococcus neoformans* isolated from 550 human dwellings in Rio de Janeiro, Brazil: an analysis of the domestic environment of AIDS 551 patients with and without cryptococcosis. *Med Mycol* 1998;36(5):305–311.
- 552 35. Brito-Santos F et al. Environmental Isolation of *Cryptococcus gattii* VGII from Indoor Dust 553 from Typical Wooden Houses in the Deep Amazonas of the Rio Negro Basin. *PLoS One* 554 2015;10(2):e0115866.
- 36. Nosanchuk JD et al. Evidence of zoonotic transmission of *Cryptococcus neoformans* from a
 pet cockatoo to an immunocompromised patient. *Annals of internal medicine* 2000;132(3):205
 208.
- 558 37. Fessel WJ. Cryptococcal meningitis after unusual exposures to birds. *New Engl J Medicine* 1993;328(18):1354 1355.
- 38. Shrestha RK, Stoller JK, Honari G, Procop GW, Gordon SM. Pneumonia due to *Cryptococcus neoformans* in a patient receiving infliximab: possible zoonotic transmission from a pet cockatiel.
 Respiratory care 2004;49(6):606 608.
- 563 39. Fraison J-B et al. Pulmonary cryptococcosis in a patient with Crohn's disease treated with 564 prednisone, azathioprine and adalimumab: Exposure to chicken manure as a source of 565 contamination. *J Crohn's Colitis* 2013;7(1):e11–e14.
- 40. Staib F, Schulz-Dieterich J. *Cryptococcus neoformans* in fecal matter of birds kept in cagesControl of *Cr. neoformans* habitats. *Zentralblatt Für Bakteriologie Mikrobiologie Und Hyg 1 Abt*Orig B Hyg 1984;179(2):179–86.
- 41. Wang C-Y, Wu H-D, Hsueh P-R. Nosocomial Transmission of Cryptococcosis. *New Engl J Med* 2005;352(12):1271–1272.
- 42. Ingram CW, Haywood HB, Morris VM, Allen RL, Perfect JR. Cryptococcal VentricularPeritoneal Shunt Infection: Clinical and Epidemiological Evaluation of Two Closely Associated
 Cases. *Infect Cont Hosp Ep* 1993;14(12):719–722.

- 43. Kanj SS et al. Fungal Infections in Lung and Heart-Lung Transplant Recipients: Report of 9 Cases and Review of the Literature. *Medicine* 1996;75(3):142–156.
- 576 44. Ooi BS, Chen BTM, LimCH, Khoo OT, Chan KT. Survival of a patient transplanted with a 577 kidney infected with *Cryptococcus neoformans*. *Transplantation* 1971;11(4):428.
- 578 45. Baddley JW et al. Transmission of *Cryptococcus neoformans* by Organ Transplantation. *Clin* 579 *Infect Dis* 2011;52(4):e94-8.
- 46. Ye F et al. Retrospective Analysis of 76 Immunocompetent Patients with Primary Pulmonary
 Cryptococcosis. *Lung* 2012;190(3):339–346.
- 582 47. Zeng Y et al. Clinicopathologic and Ultrastructural Study of Non-HIV-related Primary 583 Pulmonary Cryptococcosis in China: Report of 43 Cases. *Ultrastruct Pathol* 2011;35(1):19–25.
- 48. Setianingrum F, Rautemaa-Richardson R, Denning DW. Pulmonary cryptococcosis: A review
 of pathobiology and clinical aspects. *Med Mycol* 2018;57(2):133–150.
- 586 49. Christianson JC, Engber W, Andes D. Primary cutaneous cryptococcosis in 587 immunocompetent and immunocompromised hosts. *Med Mycol* 2003;41(3):177–188.
- 588 50. Neuville S et al. Primary Cutaneous Cryptococcosis: A Distinct Clinical Entity. *Clin Infect Dis* 2003;36(3):337–347.
- 590 51. Giles SS, Dagenais TRT, Botts MR, Keller NP, Hull CM. Elucidating the pathogenesis of 591 spores from the human fungal pathogen *Cryptococcus neoformans*. *Infect Immun* 592 2009;77(8):3491 3500.
- 593 52. Goldman DL et al. Serologic evidence for *Cryptococcus neoformans* infection in early childhood. *Pediatrics* 2001;107(5):E66.
- 53. Davis J et al. Serologic evidence for regional differences in pediatric cryptococcal infection.
 Pediatric Infect Dis J 2007;26(6):549–551.
- 597 54. Park BJ et al. Estimation of the current global burden of cryptococcal meningitis among 598 persons living with HIV/AIDS. *Aids* 2009;23(4):525–530.
- 599 55. Hurtado JC et al. Mortality due to *Cryptococcus neoformans* and *Cryptococcus gattii* in low-600 income settings: an autopsy study. *Sci Rep-uk* 2019;9(1):7493.
- 56. Galanis E, MacDougall L, Kidd S, Morshed M. Epidemiology of *Cryptococcus gattii*, British
 Columbia, Canada, 1999–2007. *Emerg Infect Dis* 2010;16(2):251–257.
- 603 57. Atkinson AJ, Bennett JE. Experience with a new skin test antigen prepared from 604 *Cryptococcus neoformans. The American review of respiratory disease* 1968;97(4):637 643.
- 58. Baker RD. The primary pulmonary lymph node complex of cryptococcosis. *American journal* of clinical pathology 1976;65(1):83 92.

- 59. Bao F, Tan H, Liu W, Li Y, Li H. Pediatric Cryptococcal Lymphadenitis in the Absence of AIDS: Case Report and Literature Review. *Case Reports Pediatrics* 2013;2013:1–4.
- 609 60. Gurung J, Lyngdoh WV, Khyriem AB. Isolated cervical cryptococcal lymphadenitis without 610 meningitis in an immunocompetent human immunodeficiency virus-negative child: a rare case 611 report. *Jmm Case Reports* 2014;1(3).
- 612 61. Jha DK, Jha AK, Singh RK. An atypical initial presentation of AIDS as cryptococcal 613 lymphadenitis. *Oxf Medical Case Reports* 2018;2018(11).
- 614 62. Chauhan S. A Rare Case of Primary Supraclavicular Lymphadenitis Due to *Cryptococcus* 615 *neoformans* in an HIV Infected Patient. *J Clin Diagnostic Res* 2014;8(2) 137-138
- 616 63. Dogbey P, Golden M, Ngo N. Cryptococcal lymphadenitis: an unusual initial presentation of 617 HIV infection. *Bmj Case Reports* 2013;2013:bcr2013010316.
- 618 64. Kawamoto K et al. Clinicopathological features of cryptococcal lymphadenitis and a review 619 of literature. *J Clin Exp Hematop* 2017;17011.
- 65. Putignani L et al. Cryptococcal Lymphadenitis as a Manifestation of Immune Reconstitution
 Inflammatory Syndrome in an HIV-Positive Patient: A Case Report and Review of the Literature. *Int J Immunopath Ph* 2008;21(3):751–756.
- 623 66. Jagadha V, Andavolu RH, Huang CT. Granulomatous Inflammation in the Acquired Immune 624 Deficiency Syndrome. *Am J Clin Pathol* 1985;84(5):598–602.
- 625 67. Hill JO. CD4+ T cells cause multinucleated giant cells to form around *Cryptococcus* 626 *neoformans* and confine the yeast within the primary site of infection in the respiratory tract. *J* 627 *Exp Medicine* 1992;175(6):1685–1695.
- 628 68. Garcia-Hermoso D, Janbon G, Dromer F. Epidemiological evidence for dormant 629 *Cryptococcus neoformans* infection. *Journal of Clinical Microbiology* 1999;37(10):3204 3209.
- 630 69. Saha DC et al. Serologic Evidence for Reactivation of Cryptococcosis in Solid-Organ 631 Transplant Recipients. *Clin Vaccine Immunol* 2007;14(12):1550–1554.
- 70. Dromer F, Ronin O, Dupont B. Isolation of *Cryptococcus neoformans* var. *gattii* from an Asian
 patient in France: evidence for dormant infection in healthy subjects. *Journal of medical and veterinary mycology* 1992;30(5):395 397.
- 635 71. Liechty CA et al. Asymptomatic serum cryptococcal antigenemia and early mortality during
 636 antiretroviral therapy in rural Uganda. *Trop Med Int Health* 2007;12(8):929–935.
- 72. Andama AO et al. Prevalence and outcomes of cryptococcal antigenemia in HIV-seropositive
 patients hospitalized for suspected tuberculosis in Uganda. *Jaids J Acquir Immune Defic Syndromes* 2013;63(2):189 194.
- 640 73. Micol R et al. Prevalence, Determinants of Positivity, and Clinical Utility of Cryptococcal
 641 Antigenemia in Cambodian HIV-Infected Patients. *Jaids J Acquir Immune Defic Syndromes*642 2007;45(5):555–559.

- 643 74. Temfack E et al. Cryptococcal Antigen Screening in Asymptomatic HIV-Infected Antiretroviral
- Naïve Patients in Cameroon and Evaluation of the New Semi-Quantitative Biosynex CryptoPS
- 645 Test. Front Microbiol 2018;09:409.
- 646 75. Chammard TB, Temfack E, Lortholary O, Alanio A. Diagnostic and therapeutic strategies in 647 cryptococcosis: impact on outcome. *Memórias Instituto Oswaldo Cruz* 2018;113(7):e180050.
- 648 76. Shi M et al. Real-time imaging of trapping and urease-dependent transmigration of 649 *Cryptococcus neoformans* in mouse brain. *J Clin Invest* 2010;120(5):1683–1693.
- 650 77. Shi M, Colarusso P, Calaruso P, Mody CH. Real-time in vivo imaging of fungal migration to 651 the central nervous system. *Cell Microbiol* 2012;14(12):1819–1827.
- 652 78. French N et al. Cryptococcal infection in a cohort of HIV-1-infected Ugandan adults. *Aids* 2002;16(7):1031–1038.
- 79. Perfect JR et al. Clinical Practice Guidelines for the Management of Cryptococcal Disease:
 2010 Update by the Infectious Diseases Society of America 2010;50(3):291–322
- 80. Molloy SF et al. Antifungal Combinations for Treatment of Cryptococcal Meningitis in Africa. *New Engl J Medicine* 2018;378(11):1004–1017.
- 658 81. Day JN et al. Combination Antifungal Therapy for Cryptococcal Meningitis. *New Engl J Med* 659 2013;368(14):1291–1302.
- 660 82. Ou X-T et al. Genotypes Coding for Mannose-Binding Lectin Deficiency Correlated With 661 Cryptococcal Meningitis in HIV-Uninfected Chinese Patients. *J Infect Dis* 2011;203(11):1686– 662 1691.
- 663 83. Sabiiti W et al. Efficient phagocytosis and laccase activity affect the outcome of HIV-664 associated cryptococcosis. *J Clin Invest* 2014;124(5):2000–2008.
- 84. Gray JV et al. "Sleeping beauty": quiescence in Saccharomyces cerevisiae. Microbiol Mol Biol *R* 2004;68(2):187 206.
- 667 85. Rittershaus ESC, Baek S-H, Sassetti CM. The Normalcy of Dormancy: Common Themes in 668 Microbial Quiescence. *Cell Host Microbe* 2013;13(6):643–651.
- 86. Finkel SE. Long-term survival during stationary phase: evolution and the GASP phenotype. *Nat Rev Microbiol* 2006;4(2):113–120.
- 671 87. Corper HJ, Cohn ML. The viability and virulence of old cultures of tubercle bacilli. *Tubercle* 672 1951;32(11):232–237.
- 673 88. Gengenbacher M, Rao SPS, Pethe K, Dick T. Nutrient-starved, non-replicating 674 *Mycobacterium tuberculosis* requires respiration, ATP synthase and isocitrate lyase for 675 maintenance of ATP homeostasis and viability. *Microbiology*+ 2010;156(1):81–87.
- 89. Setlow P. Germination of Spores of *Bacillus* Species: What We Know and Do Not Know. J *Bacteriol* 2014;196(7):1297–1305.

- 678 90. Lillie SH, Pringle JR. Reserve carbohydrate metabolism in *Saccharomyces cerevisiae*: 679 responses to nutrient limitation. *J Bacteriol* 1980;143(3):1384–94.
- 680 91. Jamieson DJ. Oxidative stress responses of the yeast *Saccharomyces cerevisiae*. *Yeast* 681 1998;14(16):1511–1527.
- 682 92. Davidson GS et al. The proteomics of quiescent and nonquiescent cell differentiation in yeast
 683 stationary-phase cultures. *Mol Biol Cell* 2011;22(7):988–998.
- 93. Takeo K, Tanaka R, Miyaji M, Nishimura K. Unbudded G 2 as well as Gj arrest in the stationary
 phase of the basidiomycetous yeast *Cryptococcus neoformans*. *Fems Microbiol Lett* 1995;129(2–
 3):231–235.
- 687 94. Mocluzuki T, Tanaka S, Saito Y, Watanabe S. Mitochondrial Kinetics During Mitosis in 688 *Cryptococcus neoformans* an Ultrastructural Study. *Mycoses* 1989;32(1):7–13.
- 689 95. Hommel B et al. *Cryptococcus neoformans* resists to drastic conditions by switching to viable 690 but non-culturable cell phenotype. *PLoS Pathog* 2019;15(7):e1007945.
- 691 96. Diamond RD, Root RK, Bennett JE. Factors influencing killing of *Cryptococcus neoformans* 692 by human leukocytes in vitro. *The Journal of Infectious Diseases* 1972;125(4):367 376.
- 693 97. Green L, Petersen B, Steimel L, Haeber P, Current W. Rapid determination of antifungal 694 activity by flow cytometry. *J Clin Microbiol* 1994;32(4):1088–91.
- 98. Nicola AM, Robertson EJ, Albuquerque P, Derengowski L da S, Casadevall A. Nonlytic
 Exocytosis of *Cryptococcus neoformans* from Macrophages Occurs In Vivo and Is Influenced by
 Phagosomal pH. *mBio* 2011;2(4):e00167-11.
- 698 99. Hardwick JM. Do Fungi Undergo Apoptosis-Like Programmed Cell Death?. *mBio* 2018;9(4):e00948-18.
- 100. Semighini CP, Averette AF, Perfect JR, Heitman J. Deletion of *Cryptococcus neoformans*AIF Ortholog Promotes Chromosome Aneuploidy and Fluconazole-Resistance in a
 Metacaspase-Independent Manner. *PLoS Pathog* 2011;7(11):e1002364.
- 101. Aouacheria A et al. Comment on "Sterilizing immunity in the lung relies on targeting fungal
 apoptosis-like programmed cell death". *Science* 2018;360(6395):eaar6910.
- 102. Goulart L et al. *Cryptococcus neoformans* and *Cryptococcus gattii* genes preferentially
 expressed during rat macrophage infection. *Med Mycol* 2010;48(7):932–941.
- 103. Chen Y et al. The *Cryptococcus neoformans* Transcriptome at the Site of Human Meningitis.
 mBio 2014;5(1):e01087-13.
- 104. Gengenbacher M, Kaufmann SHE. *Mycobacterium tuberculosis*: success through
 dormancy. *Fems Microbiol Rev* 2012;36(3):514–532.
- 105. Xu H-S et al. Survival and viability of nonculturable *Escherichia coli* and *Vibrio cholerae* in the estuarine and marine environment. *Microbial Ecol* 1982;8(4):313–323.

- 106. Salma M, Rousseaux S, Grand AS-L, Divol B, Alexandre H. Characterization of the Viable but Nonculturable (VBNC) State in *Saccharomyces cerevisiae*. *PLoS One* 2013;8(10):e77600.
- 107. Mills DA, Johannsen EA, Cocolin L. Yeast Diversity and Persistence in *Botrytis*-Affected
 Wine Fermentations. *Appl Environ Microb* 2002;68(10):4884–4893.
- 108. Toit WJD, Pretorius IS, Lonvaud-Funel A. The effect of sulphur dioxide and oxygen on the
 viability and culturability of a strain of *Acetobacter pasteurianus* and a strain of *Brettanomyces bruxellensis* isolated from wine. *J Appl Microbiol* 2005;98(4):862–871.
- 109. Liew KL, Jee JM, Yap I, Yong PVC. In Vitro Analysis of Metabolites Secreted during Infection
 of Lung Epithelial Cells by *Cryptococcus neoformans*. *PLoS One* 2016;11(4):e0153356.
- 110. Albuquerque P et al. Quorum Sensing-Mediated, Cell Density-Dependent Regulation of
 Growth and Virulence in *Cryptococcus neoformans*. *mBio* 2014;5(1):e00986-13.
- 111. Nofal M, Zhang K, Han S, Rabinowitz JD. mTOR Inhibition Restores Amino Acid Balance in
 Cells Dependent on Catabolism of Extracellular Protein. *Mol Cell* 2017;67(6):936-946.e5.
- 112. Martinez DL, Tsuchiya Y, Gout I. Coenzyme A biosynthetic machinery in mammalian cells.
 Biochem Soc T 2014;42(4):1112–1117.
- 113. Lewis K. Persister cells, dormancy and infectious disease. *Nat Rev Microbiol* 2007;5(1):nrmicro1557.
- 114. Kim J-S, Wood TK. Persistent Persister Misperceptions. *Front Microbiol* 2016;07:2134.
- 115. Ayrapetyan M, Williams TC, Baxter R, Oliver JD. Viable but Nonculturable and Persister Cells
 Coexist Stochastically and Are Induced by Human Serum. *Infect Immun* 2015;83(11):4194 4203.
- 116. LaFleur MD, Kumamoto CA, Lewis K. Candida albicans Biofilms Produce AntifungalTolerant Persister Cells v. Antimicrob Agents Ch 2006;50(11):3839–3846.
- 117. LaFleur MD, Qi Q, Lewis K. Patients with Long-Term Oral Carriage Harbor High-Persister
 Mutants of Candida albicans. *Antimicrob Agents Ch* 2010;54(1):39–44.
- 118. Spry C, Kirk K, Saliba KJ. Coenzyme A biosynthesis: an antimicrobial drug target. *Fems Microbiol Rev* 2008;32(1):56–106.
- 119. Zhao X, Zhong J, Wei C, Lin C-W, Ding T. Current Perspectives on Viable but Non-culturable
 State in Foodborne Pathogens. *Front Microbiol* 2017;8:580.

741