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Fatty acid metabolism complements glycolysis in the selective regulatory T cell expansion during tumor growth

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The tumor microenvironment restrains conventional T cell (Tconv) activation while facilitating the expansion of regulatory T cells (Tregs). Here we showed that Tregs advantage in the tumor milieu relies on supplemental energetic routes involving lipid metabolism. In murine models, tumor-infiltrating Tregs displayed intracellular lipid accumulation, which was attributable to an increased rate of fatty acid (FA) synthesis. Since relative advantage in glucose-uptake may fuel FA synthesis in intratumoral Tregs, we demonstrated that both glycolytic and oxidative metabolism concurred to Tregs expansion. We corroborated our data in human tumors showing that Tregs displayed a gene signature towards glycolysis and lipid synthesis. Our data support a model in which signals from the tumor microenvironment induce a circuitry of glycolysis, FA synthesis and oxidation that confers to Tregs a preferential proliferative advantage, whose targeting might represent a strategy for cancer treatment.

Treg | tumor microenvironment | glycolysis | fatty acid synthesis | ox40

Introduction

Regulatory T cells (Tregs), i.e., Foxp3-expressing CD4 T lymphocytes, are widely recognized as crucial controllers of immune responses, and the balance between Tregs and effector T cells determines the success of immune surveillance over cancer progression. Therefore, understanding the dynamics that dictate the relative Treg advantage over Tconvs is of utmost importance in the design of cancer immunotherapies (1). Both thymus-derived (tTregs) and peripherally induced (pTregs) subsets populate the tumor microenvironment (TME) (2): while the differentiation of pTregs (as well as of iTregs, in vitro induced) is favored in conditions of poor T cell proliferation, both pTregs and tTregs can actively proliferate in response to adequate signals (3, 4) and independently concur to generate the Treg pool in tumor-bearing hosts (5).

The T cell switch between quiescence and activation relies on a metabolic shift from oxidative to glycolytic pathways, respectively ensuring long-term survival and fuelling fast energy supply, biosynthesis and replication (6). Distinct cellular subsets display different metabolic signatures. Many pieces of evidence suggest that conventional T cells (Tconvs) require amplifying oxidative pathways, while blunting glycolysis, to undergo the terminal/restrained activation that ultimately favors their differentiation into iTreg/pTreg (7-9). First, mouse iTregs have been shown to preferentially use fatty acid oxidation (FAO) (7, 8, 10); however glycolysis is rather essential for human iTreg development upon suboptimal stimulation, since the "moonlighting" enzyme enolase-1 suppresses the transcription of the exon 2-containing FOXP3 splicing variant (FOXP3-E2, crucial determinant of the regulatory function), unless engaged in glycolysis (11).

The proliferation of pre-established Tregs may have completely distinct metabolic requirements, with respect to the conversion of Tconvs into iTregs/pTregs. Ex vivo, highly proliferative human Tregs show active mTOR and high levels of the glucose transporter Glut1; both glycolytic and FA-oxidative pathways concur to their expansion in vitro (12-14), and mTOR drives a lipogenic and cholesterogenic signature in Tregs, which crucially sustains their expansion in vivo (15). Overall, these data indicate that Tregs may arrange both glycolytic and oxidative routes and, similar to memory CD8 T cells (16), may also perform fatty acid synthesis (FAS). This may become especially true in contexts of limited nutrient availability such as the TME, where tumor cells subtract glucose to effector T cells thus blocking their glycolytic shift and anti-tumor activity (17, 18).

Here, we hypothesized that Tregs may overwhelm Tconvs in the hostile tumor context thanks to their peculiar ability to exploit supplemental metabolic routes and thus to optimize nutrient usage. We first documented that, in a mouse tumor model, tumor-infiltrating Tregs preferentially capture and utilize glucose not only to fuel glycolysis but also to synthesize and accumulate FAs, and Tregs relied on FAS and FAO to a higher extent than Tconvs in vitro. Genes involved in glycolysis and lipid biosynthesis are enriched in the signature of activated Tregs sorted from human liver cancer. Thus, exploiting a combination of different

Significance

Recent studies have established that metabolic restrains, such as glucose restriction, impair the activities of effector T cells in the tumor microenvironment. In the same context, a huge expansion of activated regulatory T cells in tumor tissues has been described in mice and humans, contributing to the suppression of protective anti-tumor immunity. Our data demonstrate that Tregs are committed to survive and proliferate in such a hostile milieu thanks to a metabolic advantage based on the combination of glycolysis and fatty acid synthesis and oxidation. This allows Treg to prevail over Tconv that rely primarily on the glycolytic pathway for their metabolic demands. The awareness of the metabolic dynamics of Treg in tumor could provide new means for cancer immunotherapy.

Reserved for Publication Footnotes

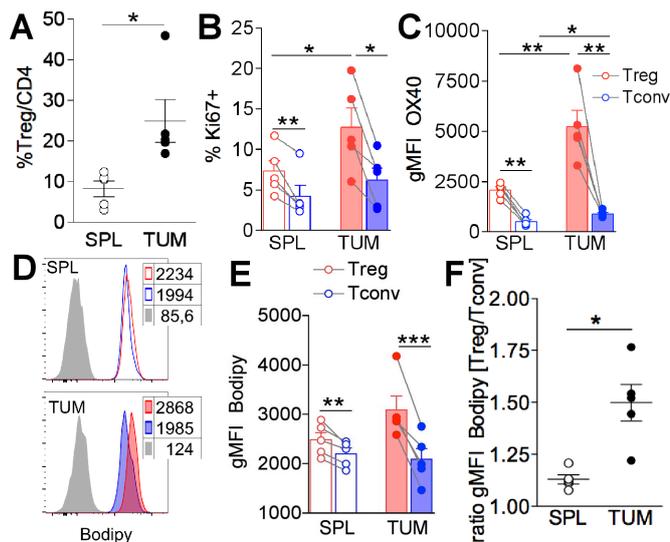


Fig. 1. Expanded and activated tumor-infiltrating Tregs display high intracellular lipid content. 5×10^5 MCA38 cells were injected s.c. into C57BL/6 mice, and flow cytometry analysis was performed on lymphocytes extracted from spleen (SPL) and tumor (TUM) after 14 days. **(A)** Frequency of CD25+Foxp3+ Tregs in gated CD4+ lymphocytes. **(B)** Frequency of cells expressing Ki67 and **(C)** geometric mean fluorescent intensity (gMFI) of OX40 in gated CD25+Foxp3+ Tregs (red) and CD25-Foxp3- Tconvs (blue) in SPL (empty bars) and TUM (filled bars) from tumor-bearing mice. **(D)** Representative histograms and **(E)** gMFI analysis of Bodipy incorporation in gated CD25+Foxp3+ Tregs (red) and CD25-Foxp3- Tconvs (blue) in SPL (empty peaks and bars) and TUM (solid peaks and bars) from tumor-bearing mice. Peaks in grey represent the fluorescence-minus-one (FMO) controls. **(F)** Ratio between Bodipy gMFI in Tregs and Tconvs from SPL and TUM. Error bars show mean \pm SEM, each dot corresponds to a single mouse. Data shown are from a representative out of two independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.005$, by Student t test, paired between Tregs and Tconvs in the same samples; unpaired, between populations of different samples.

metabolic routes may be a novel immune escape mechanism that conveys to Tregs a selective advantage in the TME.

Results

Proliferating Tregs in the TME accumulate intracellular lipids.

In the visceral adipose tissue (VAT), expanded Tregs accumulate intracellular lipids under the cell-intrinsic control of the adipogenic factor PPAR γ (19). Therefore, we first checked whether lipid accumulation also occurred in another context of Treg expansion, i.e., the TME. To this aim, CD4+Foxp3+ Tregs were analyzed by flow cytometry, in comparison with CD4+Foxp3- Tconvs, in cells extracted from tumor beds (TUM), or from spleens as control (SPL), of C57BL/6 wild-type mice, bearing nodules of the subcutaneously implanted colon carcinoma cell line MCA38 (the gating strategy is shown in SI Appendix, Fig. S1). Confirming previous studies in a variety of mouse tumor models and in human cancers (1), Treg frequency was significantly increased at the tumor site (Fig. 1A); compared to spleen cells and to Tconvs, TUM-Tregs were more proliferative, as pointed out by Ki67+ frequency (Fig. 1B), and expressed higher levels of the receptor OX40 (Fig. 1C), which we previously demonstrated to sustain Treg fitness in mouse models of homeostatic proliferation and colitis (20), and to promote the expansion of stable and suppressive Tregs in human cancers (21, 22).

When we stained the cells with Bodipy, a cell membrane-permeable fluorophore specific for neutral lipid stores, we could detect a higher intracellular lipid content in Tregs compared to Tconvs in both SPL and TUM (Fig. 1D-E and SI Appendix, Fig. S1B); however, this phenomenon affected especially TUM-Tregs, as evidenced by a significantly increased ratio of Bodipy levels,

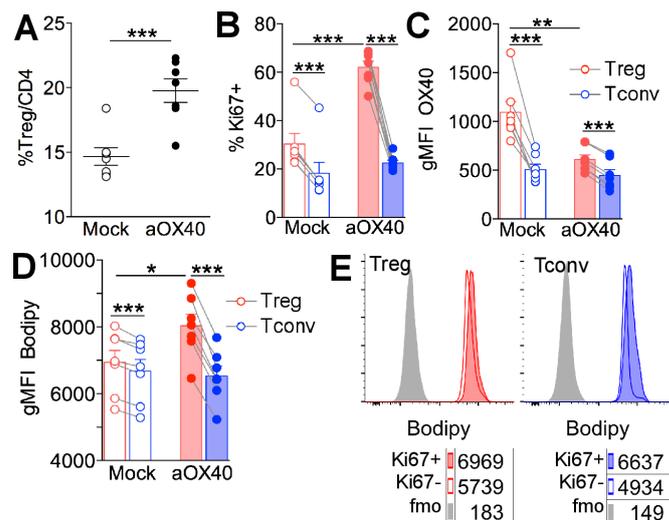


Fig. 2. OX40 triggering alone recapitulates Treg proliferation and FA accumulation in vivo. C57BL/6 naïve mice were injected i.p. with 500 μ g of anti-OX40 agonist mAb (OX86) or PBS as mock control. After 6 days, flow cytometry analysis was performed in splenocytes. **(A)** Frequency of CD25+Foxp3+ Tregs in gated CD4+ lymphocytes. **(B)** Frequency of cells expressing Ki67, **(C)** geometric mean fluorescent intensity (gMFI) of OX40, and **(D)** gMFI analysis of Bodipy incorporation, in gated CD25+Foxp3+ Tregs (red) and CD25-Foxp3- Tconvs (blue) in aOX40-treated compared to mock-treated mice. **(E)** Representative histograms showing Bodipy labeling in gated Ki67+ or Ki67- Tregs or Tconvs; numbers indicate Bodipy gMFI; peaks in grey represent the fluorescence-minus-one (FMO) controls. Error bars show mean \pm SEM, each dot corresponds to a single mouse; 6-7 mice were included in each experimental group. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.005$, by Student t test, paired between Tregs and Tconvs in the same samples; unpaired, between populations of different samples.

between Tregs and Tconvs, in TUM compared to SPL (Fig. 1F). Similar results were obtained in a distinct tumor model, i.e., the subcutaneously injected melanoma B16F10 (SI Appendix, Fig. S2).

To ascertain whether the preferential lipid accumulation in Tregs was an exclusive feature of tumor, or was shared with other conditions characterized by Treg expansion, we analyzed Treg frequency and intracellular lipid content in two distinct models of liver disease, namely the *Mdr2*^{-/-} mouse (developing from an early age inflammatory cholangitis that progresses into cirrhosis and cancer (23)), and the HCV-transgenic (HCVTg) mouse (spontaneously developing steatosis at advanced ages with no evidence of inflammation (24)). In young (3 weeks old) *Mdr2*^{-/-} compared to wild-type mice, hepatic Tregs displayed a higher frequency, and this was again associated with greater intracellular lipid content in relation to Tconvs (SI Appendix, Fig. S3A-C); conversely, hepatic Tregs were not expanded in aged (>10 months old) HCVTg compared to wild-type mice, and no preferential lipid accumulation was detected in Tregs (SI Appendix, Fig. S3D-F). Overall these data indicate that Tregs enlarge their intracellular lipid pool when prompted to expansion, irrespective of whether triggered by inflammation- or tumor-associated stimuli.

We have previously demonstrated that OX40 plays non-redundant roles in Treg expansion in a variety of contexts including cancer (20, 21); therefore, we sought to test whether OX40 signal could induce lipid accumulation in proliferating Tregs. To this aim, we injected an anti-OX40 agonist mAb, OX86, in naïve mice: according to published data (25), OX40 triggering induced a huge increase of Treg frequency (Fig. 2A). This expansion was likely due to the direct promotion of the selective proliferation of Tregs to the expense of Tconvs, as demonstrated by the increased percentage of Ki67+ Tregs, but not Ki67+ Tconvs (Fig. 2B),

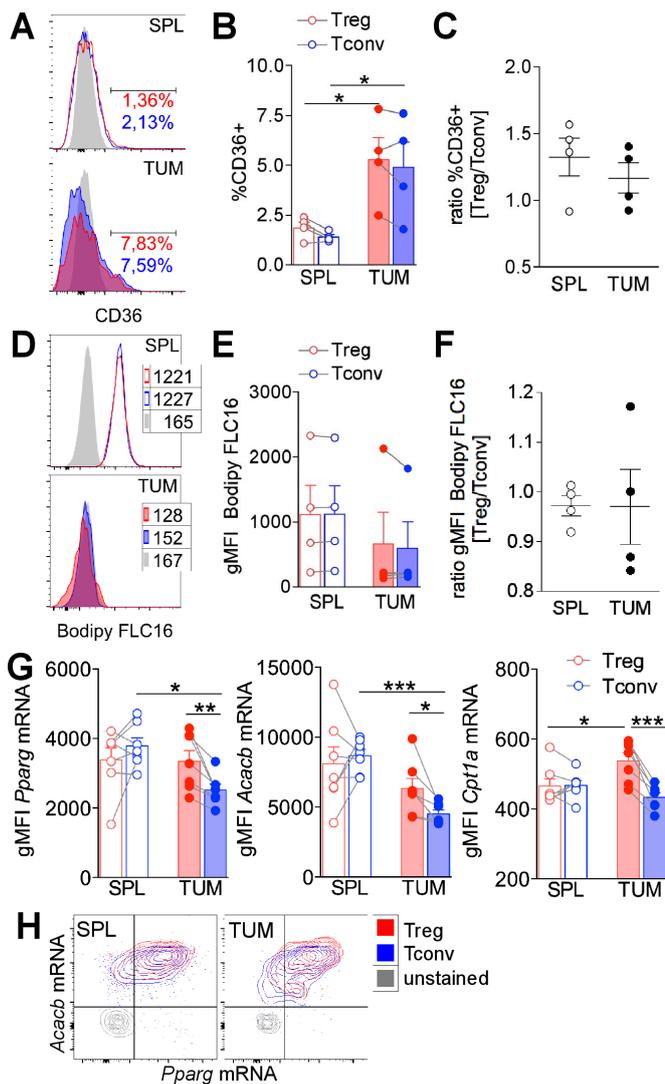


Fig. 3. Fatty acid synthesis, rather than uptake, shapes the lipid Treg pool and contributes to Treg proliferation. (A-F) Mice bearing tumors of 8x8 mm in size received an i.p. injection of the green fluorescent palmitate (Bodipy FL C16), 1 hour before spleen (SPL) and tumor (TUM) collection. Then Bodipy FL C16 uptake and CD36 expression were quantified by flow cytometry in CD25+Foxp3+ Tregs and CD25-Foxp3- Tconvs. (A) Representative histograms and (B) frequency of cells expressing CD36 in gated Tregs (red) and Tconvs (blue) in SPL (empty peaks and bars) and TUM (solid peaks and bars). (C) Ratio between frequency of CD36-expressing Tregs and Tconvs in SPL and TUM. (D) Representative histograms and (E) geometric mean fluorescence intensity (gMFI) of Bodipy FL C16 acquisition in gated Tregs (red) and Tconvs (blue) in SPL (empty peaks and bars) and TUM (solid peaks and bars). (F) Ratio between Bodipy FL C16 gMFI of Tregs and Tconvs in SPL and TUM tissue. Peaks in grey represent the fluorescence-minus-one (FMO) controls. (G-H) The expression level of *Pparg*, *Acacb* and *Cpt1a* mRNA was assessed in gated CD4+CD25+ Tregs and CD4+CD25- Tconvs by PrimeFlow RNA assay. (G) Geometric mean fluorescence intensity of each gene expression in Tregs (red) and Tconvs (blue) in SPL (empty bars) and TUM (filled bars). (H) Representative overlay contour plots of *Acacb* and *Pparg* co-expressing cells in the indicated samples. Each dot corresponds to a single mouse; data shown are from a representative out of four independent experiments each including 3-7 mice per group. Error bars show mean \pm SEM, * p < 0.05, ** p < 0.01, *** p < 0.005, by Student t test, paired between Tregs and Tconvs in the same sample; unpaired, between populations of different samples.

and also corroborated by the decreased OX40 staining ex vivo (with the same OX86 clone) of Tregs but not Tconvs of aOX40-treated mice (Fig. 2C). Notably, aOX40-induced Treg expansion

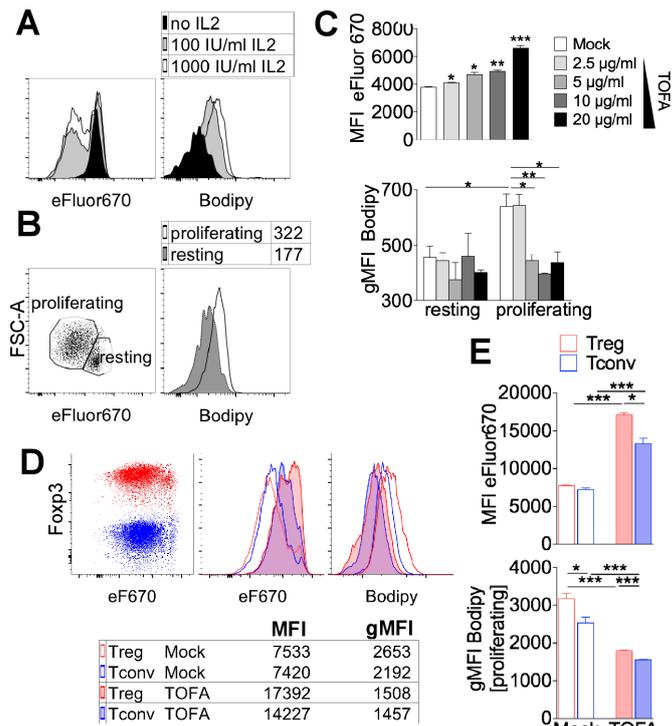


Fig. 4. Inhibition of fatty acid synthesis blocks proliferation and lipid accumulation preferentially in Tregs in vitro. (A-C) CD4+CD25+ Tregs, magnetically purified from splenocytes of naive C57BL/6 mice, were labeled with eFluor670 and polyclonally stimulated for 4 days, then stained with Bodipy and analyzed by flow cytometry. (A) Representative histograms of eFluor670 dilution (left) and Bodipy incorporation (right) in Tregs cultured with different IL-2 concentrations. (B) Representative plot of proliferating and resting Treg gating strategy (left) and representative histogram of Bodipy incorporation within each gate (right). (C) Mean fluorescence intensity (MFI) of eFluor670 in gated Tregs (left), and geometric mean fluorescence intensity (gMFI) of Bodipy incorporation in gated resting and proliferating Tregs (right), cultured with different TOFA concentrations. (D-E) Magnetically purified Tregs and Tconvs were co-cultured at 1:1 ratio with feeder cells, anti-CD3 and IL-2, with or without TOFA (5 μ g/ml). After 4 days, cells were stained with Bodipy and Foxp3, and flow cytometry analysis was performed in gated Tregs (Foxp3+, red) or Foxp3- (Tconvs, blue). (D) Representative overlay of Tregs and Tconvs in the indicated conditions; numbers indicate MFI of eFluor670 or gMFI of Bodipy. (E) Analysis of eFluor670 MFI or Bodipy gMFI (gated on proliferating cells) Tregs (red bars) or Tconvs (blue bars), either mock-treated (empty) or TOFA-treated (filled). Each condition was tested in triplicates; data shown are from a representative out of two independent experiments. Error bars show mean \pm SEM, * p < 0.05, ** p < 0.01, *** p < 0.005, by Student t test, unpaired.

was associated to a greater Bodipy staining of Tregs, which was even more pronounced in Ki67+ Tregs (Fig. 2D-E). These results strengthen the link between Treg proliferation and FA accumulation, and demonstrate that the sole OX40 engagement can trigger the selective proliferation of lipid-laden Tregs.

FA synthesis, rather than uptake, shapes the TUM-Treg lipid pool and promotes Treg proliferation.

In the adipose tissue, lipid-laden Tregs also display higher levels of the FA translocator CD36 (19); therefore, we tested whether a differential uptake of exogenous FAs might account for the higher lipid content in TUM-Tregs. We found that CD36 was modestly upregulated at the tumor site in both Tregs and Tconvs, but without any specific enrichment in TUM-Tregs (Fig. 3A-C). Then, we measured the absorption in vivo of a fluorescently labeled FA (Bodipy FL C16, a palmitic acid molecule conjugated to the Bodipy fluorophore), previously injected into tumor-bearing mice: both Tregs and Tconvs tended to acquire less

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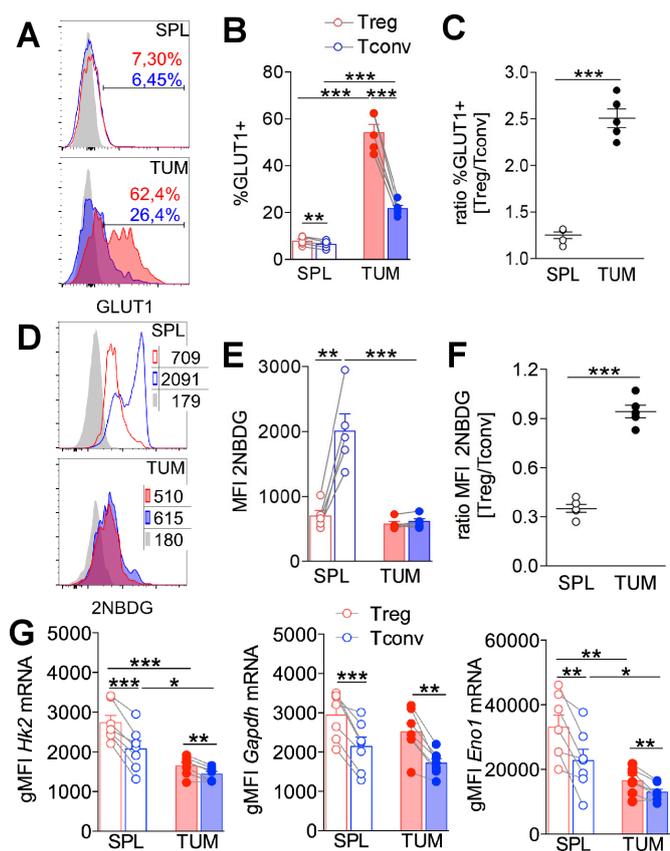


Fig. 5. Glucose uptake and usage occurs preferentially in Tregs in tumors. (A-F) Mice bearing tumors of 8x8 mm in size received an i.p. injection of the fluorescent glucose analogue 2-NBDG, and spleen (SPL) and tumor (TUM) were collected 15 minutes after injection. 2-NBDG uptake and GLUT1 expression were quantified by flow cytometry in CD25+Foxp3+ Tregs and CD25-Foxp3-Tconvs. **(A)** Representative histograms and **(B)** frequency of cells expressing GLUT1 in gated Tregs (red) and Tconvs (blue) in SPL (empty peaks and bars) and TUM (solid peaks and bars); grey peaks represent the fluorescence-minus-one (FMO) controls. **(C)** Ratio between frequency of GLUT1 expressing Tregs and Tconvs in SPL and TUM. **(D)** Representative histograms and **(E)** mean fluorescence intensity (MFI) of 2-NBDG acquisition in gated Tregs (red) and Tconvs (blue) in SPL (empty peaks and bars) and TUM (solid peaks and bars). **(F)** Ratio between 2-NBDG MFI of Tregs versus Tconvs at different sites. **(G)** Expression levels of the glycolysis-related genes *Hk2*, *Gapdh* and *Eno1*, evaluated through PrimeFlow RNA assay in gated CD4+CD25+ Tregs (red) and CD4+CD25- Tconvs (blue) from SPL (empty bars) and TUM (filled bars). Each dot corresponds to a single mouse; data shown are from a representative out of three independent experiments each including 3-7 mice. Error bars show mean \pm SEM, * p < 0.05, ** p < 0.01, *** p < 0.005, by Student t test, paired between Tregs and Tconvs in the same sample; unpaired, between populations of different samples.

FA in the tumor site compared to the spleen (despite the modest CD36 upregulation) but without any major difference between the two subsets (**Fig. 3D-F**). These results suggested a local defect in FA distribution and uptake, shared by Tregs and Tconvs and unrelated to their CD36 levels, and excluded preferential FA scavenging by TUM-Tregs: thus, we investigated whether intracellular FA metabolic routes were instead involved in lipid accumulation and proliferation of Tregs. First, we evaluated, in Tregs and Tconvs from tumor-bearing mice, the expression levels of 3 selected genes, *Pparg*, *Acacb* and *Cpt1a*, respectively required for FA storage, synthesis and oxidation. To this aim, we adopted PrimeFlow RNA Assay, allowing the simultaneous detection of transcripts and proteins by flow cytometry at single cell resolution. Thanks to this approach, we could detect that TUM-Tregs expressed all 3 genes at significantly higher levels

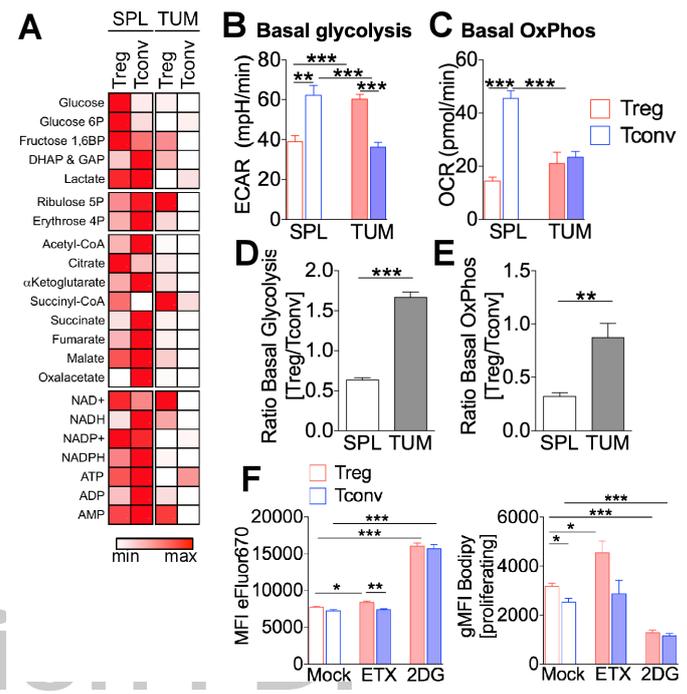


Fig. 6. TUM-Treg display higher levels of both glycolytic and oxidative metabolism. (A) CD4+CD25+ Tregs and CD4+CD25- Tconvs were magnetically purified from pooled spleens and tumors of tumor-bearing mice, and metabolomic profiling was performed directly ex vivo by LC-MS/MS. The heatmap displays the levels of each metabolite (fg/cell) in the indicated samples (min/max related to each metabolite). **(B-E)** CD4+CD25+ Tregs and CD4+CD25- Tconvs were magnetically purified from pooled spleens and tumors of tumor-bearing mice, and quantification of extracellular acidification rate (ECAR) and oxygen consumption rate (OCR) were performed by Seahorse analysis after a 12-hours stimulation with anti-CD3/anti-CD28. **(B)** Basal glycolysis, **(C)** basal OxPhos, and **(D-E)** respective Treg/Tconvs ratios for each index, in the indicated conditions. Data shown are from a representative out of two independent experiments. Data are expressed as mean \pm SEM of three measurements, each collected in 2-8 replicates. ** p < 0.01, *** p < 0.005, by Student t test, unpaired. **(F)** Magnetically purified Tregs and Tconvs were co-cultured at 1:1 ratio for 4 days with or without EtX (100 μ M) or 2-DG (1 mM), stained with Bodipy and Foxp3, and analyzed by flow cytometry. Analysis of eFluor670 MFI or Bodipy gMFI (gated on proliferating cells) Tregs (red bars) or Tconvs (blue bars) is shown in the indicated conditions. Data shown as mean \pm SEM; each condition was tested in triplicates; data shown are from a representative out of two independent experiments. * p < 0.05, ** p < 0.01, *** p < 0.005, by Student t test, unpaired.

compared to TUM-Tconvs (**Fig. 3G**). Of note, *Pparg* and *Acacb* expression tended to co-segregate in all cell types and at higher levels in TUM-Tregs (Fig. 3H), suggesting a link between PPAR γ expression and FAS program specifically in these cells.

Therefore, we tested whether TOFA, an inhibitor of acetyl-CoA carboxylase (ACC), key enzyme in FAS cascade, impacted on Treg proliferation. First, we administered TOFA to tumor-bearing mice, performing repeated intra-tumor injections starting when nodules were palpable: this treatment significantly suppressed tumor growth; however, this effect was unlikely due to immune-mediated events, as we failed to detect any significant increase in CD8 T cell frequency or IFN γ production in TOFA-treated compared to mock-treated nodules (SI Appendix, **Fig. S4A-B**). Rather, direct TOFA toxicity against tumor cells might account for the anti-tumor activity observed in vivo: indeed, TOFA reduced also in vitro the viability of the MCA38 cell line in a dose-dependent fashion, as evidenced in XTT cytotoxicity assay (SI Appendix, **Fig. S4C**).

To assess the role of FAS directly on Tregs, we took advantage of a model of Treg proliferation in vitro, which recapitulated

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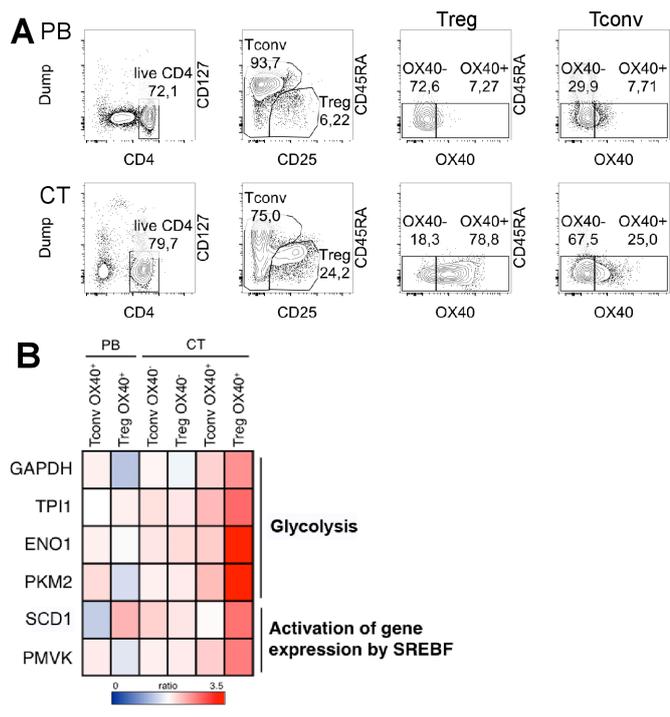


Fig. 7. Human Treg proliferation, in vitro and ex vivo from diseased liver, is associated with glycolytic gene expression. (A-B) OX40+/OX40- CD45RA^{low} Tregs and Tconvs were extracted and sorted from PB and cirrhosis/tumor (CT) of 5 patients with chronic HCV infection, and gene expression analysis was performed. (A) Sorting strategy. (B) The fold-change was calculated for each gene over the respective control (OX40- PB Tconvs for all Tconv samples, and OX40- PB Tregs for all Treg samples), and 216 genes upregulated >2fold only in Treg OX40+ CT were selected for pathway analysis (www.reactome.org). The heatmap displays the fold-change over control in the genes accounting for two pathways of interest, among those showing statistically significant enrichment (p<0.05).

the lipid accumulation occurring in TUM-Tregs in vivo. To this aim, Tregs were enriched from the spleens of naïve mice and polyclonally stimulated in vitro in the presence of IL-2: in this context, we could detect an increased incorporation in Tregs of the Bodipy lipophilic fluorophore, proportionally to IL-2 dosage and to the extent of proliferation (Fig. 4A); notably, Bodipy levels were higher in proliferating than in resting cells, indicating active lipid accumulation during Treg cell division (Fig. 4B). In this setting, TOFA treatment significantly inhibited, in a dose-dependent fashion, the proliferation of Tregs, and completely abolished FA accumulation selectively in proliferating cells, demonstrating a key role for FAS in both expansion and lipid pool generation of Tregs (Fig. 4C). Of note, contrary to tumor cells, TOFA exposure did not exert a direct cytotoxic effect over Tconvs or Tregs, but rather increased their survival (SI Appendix, Fig. S5).

To ascertain whether lipid accumulation and FAS reliance were preferential features of Tregs, we purified Tregs and Tconvs from spleens of naïve mice, labeled them with a proliferation tracer, and co-cultured them in the presence of IL-2: according to the literature (26), IL-2 supply allowed both Tregs and Tconvs to proliferate to a similar extent (Fig. 4D). Here, in basal conditions, we found that Tregs showed significantly higher Bodipy content, yet same proliferation levels (as suggested by eFluor670 mean fluorescence intensity levels), compared to Tconvs, denoting that also in vitro Tregs were more prone to accumulate lipids during cell division; notably, the addition of TOFA significantly reduced proliferation and Bodipy staining in both Tregs and Tconvs, however Tregs were significantly more sensitive to such inhibition (Fig. 4D-E).

Finally, we sought to test whether TOFA-treated Tregs were less able to suppress, when adoptively transferred in vivo, the immunity elicited by a primary growing tumors against a secondary challenge, i.e. “concomitant immunity” (27). To this aim, mice bearing palpable tumors received a second injection of tumor cells alone or admixed with TOFA- or mock-pretreated Tregs: the analysis of growing secondary tumors-infiltrating CD8 T cells revealed a trend for a decreased IFN γ and TNF α production in mice receiving tumor cells plus Tregs compared to tumor cells alone; notably, this suppressive effect was rescued when TOFA-treated Tregs were co-transferred with tumor cells (SI Appendix, Fig. S6). Together with in vitro data, these results point to a key role of FAS in Treg proliferation and possibly also in their suppressive function in vivo.

Glycolysis and fatty acids oxidation both concur to Treg advantage in cancer.

Memory CD8 T cells were shown to synthesize FAs from glucose (16), therefore we hypothesized that also Tregs might fuel FAS through a greater absorption of glucose, relative to effector T cells, which are instead damaged by glucose deprivation in the TME (17, 18). First, we analyzed, in Tregs and Tconvs from tumors and spleens, the expression of Glut1, a prominent glucose transporter in CD4 T cell activation (28) that is also relevant for Treg proliferation (12): we found that Glut1 was up-regulated in both Tconvs and Tregs at the TUM site, compared to SPL; however, such induction was significantly higher in Tregs, resulting in an increased relative ratio of Glut1 expression in Tregs versus Tconvs in tumors (Fig. 5A-C). To directly track glucose uptake in vivo, we injected the fluorescent glucose analogue 2NBDG into tumor-bearing mice: in line with previous data (17, 18), Tconvs showed a massive defect in glucose uptake in TUM compared to SPL; conversely, Tregs displayed a relatively low glucose absorption in SPL that was maintained in TUM, resulting in a significant rescue of the Treg/Tconv relative ratio of glucose uptake at the tumor site (Fig. 5D-F).

Then, we analyzed the expression of selected glycolysis-related genes (*Hk2*, *Gapdh*, and *Eno1*), through PrimeFlow RNA Assay, in Tregs and Tconvs: all 3 genes were significantly more represented in TUM-Tregs compared to TUM-Tconvs (Fig. 5G). Interestingly, *Hk2* and *Eno1* (being the latter a master regulator of human Treg metabolism and development (11)), were repressed in tumors, compared to spleens, in both Tregs and Tconvs, in line with a general contraction of glycolysis in the tumor context; however, their expression remained higher in Tregs in both compartments, pointing to a preferential preservation of glycolytic activity in Tregs in the TME (Fig. 5G, right).

The above results delineate a scenario in which Treg advantage in tumor may be sustained by the concomitant engagement of glucose and lipid metabolic routes. To address this aspect, we performed a metabolomic analysis of Tregs and Tconvs isolated from tumors or spleens: profoundly different profiles emerged from these T cell subsets (Fig. 6A). In more detail, we could detect a series of metabolic changes in TUM-Tregs, compared to SPL-Tregs: TUM-Tregs displayed a preserved pentose phosphate (PP) pathway but higher glycolytic flux, higher consumption of acetyl-CoA and of the first intermediates of TCA cycle: citrate and α -ketoglutarate (SI Appendix, Fig. S7A). Of note, this profile was paralleled by increased NADH/NAD⁺ and NADPH/NADP⁺ ratios, and lower ATP/ADP ratios (SI Appendix, Fig. S7B), suggesting that in TUM-Tregs an improved glucose usage may fuel TCA cycle, whose intermediates are likely diverted to FA-biosynthetic pathways rather than being fully oxidized to generate ATP.

To functionally validate the peculiar TUM-Treg metabolic signature, we analyzed the bioenergetic profile of Tregs and Tconvs, isolated from SPL or TUM and cultured in vitro for 12 hours, through Seahorse assays. Basal glycolysis, as measured by extra-

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cellular acidification rate (ECAR), in TUM compared to SPL, was repressed in Tconvs, as expected (18), but strikingly enhanced in Tregs (Fig. 6B). The rate of basal oxidative phosphorylation (OxPhos), as assessed by oxygen-consumption rate (OCR), was again inhibited in Tconvs at the tumor site, but maintained in Tregs (Fig. 6C). For all the metabolic indexes analyzed, Tregs showed a relative advantage over Tconvs specifically at the tumor site, as evidenced by higher Treg/Tconv ratios (Fig. 6D-E).

These data demonstrated that tumor-associated Tregs engage both glycolysis and OxPhos, for their expansion. To assess whether these metabolic routes are differentially required for Treg and Tconv proliferation, we performed the Treg/Tconv co-culture assay described above, in the presence of 2DG or etomoxir (Etx), selective inhibitors of glycolysis or FAO, respectively. In line with previous results with human Tregs (14), 2DG profoundly restrained proliferation and lipid accumulation in both Tconvs and Tregs to a similar extent, suggesting that both cell types required glycolysis to support cell division and FAS; conversely, Etx affected, slightly though significantly, only Treg proliferation, and only in Tregs it promoted a further increase in Bodipy staining, indicating that FAO consumed the intracellular lipid pool and contributed to cell division specifically in Tregs (Fig. 6F). In analogy with the above-described effect of TOFA, we could not test also in vivo whether Etx blocked Treg proliferation in tumors, due to its direct toxicity against tumor cells that overshadowed any immune-mediated effect (Fig. S3D-F). However, the greater sensitivity to FAO blockade in vitro in the co-culture assay, and the higher rate of OxPhos ex vivo from tumors, together highlight the reliance of Tregs on FA utilization, and suggest that Tregs may display a relative resistance to glucose restriction in the TME due to their special ability to engage multiple metabolic routes, such as glycolysis and FA synthesis/oxidation.

Glycolytic and lipid biosynthetic pathways are active in human tumor-associated Tregs

Compared to mouse tumors, Tregs infiltrating human cancers display superior complexity and heterogeneity, which is not completely understood especially in immunometabolic aspects. Human cancer-infiltrating Tregs reveal a peculiar gene signature, characterized by specific chemokine receptors, immune checkpoints, and costimulatory molecules such as OX40 (29, 30). Based on our recent work, demonstrating that OX40-expressing Tregs were expanded in hepatic cirrhosis and liver and colon cancers, and that OX40 was connected to high proliferation, stability and suppression of tumor-associated Tregs (21, 22), we explored the gene signature of OX40+ Tregs, freshly extracted from specimens of liver cirrhosis and tumor (CT) or from peripheral blood (PB) of patients with chronic HCV infection and hepatocellular carcinoma (HCC), in comparison to OX40- Tregs and same subsets of Tconvs (Fig. 7A). We selected 216 genes showing a >2-fold increase selectively in CT OX40+ Tregs, and performed a pathway enrichment analysis: notably, among the significantly enriched pathways, we could reveal two metabolic routes, namely glycolysis ($p=0.00148$) and activation of gene expression by SREBF ($p=0.0466$) (Fig. 7B and SI Appendix, Tables S1-S2). Glycolysis-related genes included *GAPDH*, *TPII*, *ENO1*, and *PKM2*, all involved in the glycolytic cascade, and *PKM2* in particular in substrate phosphorylation; the up-regulated genes belonging to the SREBF-related pathway were *SCD1* and *PMVK*, respectively involved in monounsaturated FA synthesis and in mevalonate pathway for cholesterol biosynthesis. In conclusion, these data demonstrate that also human Tregs shift between different metabolic routes in specific tissue contexts.

Discussion

The TME poses metabolic hurdles, such as hypoxia and glucose restriction, to the development of protective anti-tumor immunity. Nevertheless, this context favors the accumulation of

proliferating and activated Tregs: our study demonstrates that such advantage may rely on their capacity to compete for glucose and perform FAS at higher rates compared to Tconvs.

Whether Tregs utilize glycolysis for their functions has been a matter of controversy in recent years. While it has been shown that murine iTregs (mostly polarized in the presence of TGF β) show low glycolytic rates, and that glycolysis inhibition promotes their differentiation (7-10), human iTregs (differentiated with suboptimal stimulation) strictly rely on glycolysis for development and function (11). Human Tregs are highly glycolytic when freshly analyzed from human blood, and strongly require both FAO and glycolysis for their proliferation in vitro, while Tconvs rely only on glycolysis (13). Our present data confirm the same requirements in murine cells in vitro, and also demonstrate that Treg expansion in vivo in the tumor context is sustained by a strong increase in their glycolytic rate compared to Tconvs.

Others have shown that Glut1 is higher in proliferating Tregs from the spleens of naïve mice, and that Glut1 overexpression increased both glycolytic and oxidative metabolism in Tregs and provoked Treg expansion in the spleen; however, these Tregs showed signs of reduced stability and suppressive function (12, 28). These results suggest that an artificial boost in glycolysis may uncouple proliferation from suppressive function in Tregs. However, more and more data indicate that these two events are tightly interconnected, especially in tissue contexts where Tregs are activated: indeed, despite their refractoriness to conventional stimulation in vitro, Tregs are a highly proliferative population in vivo in both mice and humans, and many pieces of evidence demonstrate that activated Treg, displaying a higher proliferation rate, retain or even increase their suppressive ability in several contexts (31). Few examples are the spontaneous development of lethal autoimmunity due to impaired expansion of suppressive Tregs in mTORC1 deficiency (15), the impaired ability of OX40-null Tregs to suppress colitis because of defective competitive fitness (20), the ability of OX40/OX40L to induce the proliferation of highly suppressive human Tregs (21), and the lower proliferative potential of Tregs in multiple sclerosis patients with a more severe disease (32). In cancer mouse models and cancer patients, a huge amount of data demonstrates that highly proliferative and highly suppressive Tregs accumulate in tumors (2). Accordingly, our current results show, at the tumor site, higher frequency of Ki67+ and OX40+ Tregs, in association with increased glycolytic rate, higher Glut1 expression and improved competition for glucose. Combined with the well-established notion of the superior suppressive function of tumor-infiltrating Tregs, our data contribute to delineate a picture in which a glycolytic shift may boost proliferation and, consequently, suppressive function of Tregs in certain tissues and especially in cancer.

Foxp3 expression alone is sufficient to shift the metabolic program from glycolysis to oxidative phosphorylation, and enables Tregs to resist better to low-glucose, high-lactate exposure (12, 33): such default program may help Tregs to maintain their quiescent state in adverse conditions. However, our results indicate the presence of an active Treg proliferation in the tumor context, possibly sustained by glycolysis, and associated with the expression of OX40 as a marker of Treg activation; furthermore, OX40 engagement directly promotes the selective proliferation of lipid-laden Tregs in vivo. Overall, considering these results, it may be proposed that, on the one side, Foxp3 induces a default oxidative program to maintain Treg survival in a quiescent state; on the other side, the signal elicited through OX40, and possibly other costimulatory receptors and other members of the TNF receptor superfamily, may superimpose a glycolytic and anabolic boost over the oxidative program, allowing Tregs to exit from quiescence, undergo active proliferation and perform efficient suppressive function.

817 De novo synthesized intracellular lipids may be utilized not
818 only for metabolic but also functional purposes. Lipids are a
819 preferential source of acetyl groups for histone acetylation and
820 epigenetic reprogramming (34), and short-chain FA uptake has
821 been shown to support colonic Treg homeostasis through regu-
822 lating histone acetylation (35, 36). The stability of Foxp3 itself
823 is increased by acetylation (37) and may be thus sensitive to the
824 supply of lipid-derived acetyl groups. Lipids are also necessary for
825 the palmitoylation of crucial signaling proteins, for building cell
826 membranes and shaping their lipid composition and fluidity, for
827 the synthesis of pro- as well as anti-inflammatory lipid mediators.
828 Our results suggest that TUM-Tregs may perform FAS not only
829 to fuel FAO but also for other purposes. Indeed, the high neutral
830 lipid content, and also the metabolite signature, observed in
831 TUM-Tregs ex vivo indicates that FA synthesis may prevail over
832 FA consumption; the glycolytic rate was boosted in TUM-Tregs to
833 a greater extent when compared to oxidative indexes, and the co-
834 induction of *Pparg* and *Acacb* was higher than *Cpt1a* increase in
835 TUM-Tregs; finally, in vitro, Treg proliferation was more sensitive
836 to the blockade of FAS than of FAO. Human OX40+ tumor-
837 infiltrating Tregs displayed a gene signature oriented to glycoly-
838 sis and lipid biosynthesis. Overall these data indicate that lipid
839 biosynthetic pathways may dominate in active Treg expansion
840 in cancer and also in inflammatory conditions; accordingly, im-
841 paired Treg homeostasis in mTORC1-deficient mice associates to
842 defective lipid biosynthesis (15).

843 According to our gene expression analysis in OX40+ Tregs
844 extracted from human tumor, a SREBP-driven program may
845 support lipid biosynthesis in these cells. While no comprehensive
846 data exist on the role of this family of transcription factors in Treg
847 homeostasis and suppressive function, their function has been
848 clearly elucidated in CD8 T cell activation: SREBP was essential
849 for T cell transition from quiescence to activation through the
850 active promotion of membrane synthesis (38). We would specu-
851 late that, likewise, a program of lipid biosynthesis, addressed
852 to the construction of functional membranes, might support the
853 local proliferation of OX40+ Tregs in the TME. Indeed, the two
854 SREBP-related genes, *SCD1* and *PMVK*, upregulated in tumor
855 OX40+ Tregs, may concur to the biosynthesis of those lipids,
856 namely monounsaturated FAs and especially cholesterol, that
857 more profoundly impact on membrane fluidity, lipid-raft forma-
858 tion and TCR clustering and signaling, as recently demonstrated
859 in CD8 T cells (39).

860 We report here that OX40 triggering in naïve mice prompt the
861 proliferation of lipid-laden Tregs: therefore, the OX40/OX40L
862 signal, occurring in the HCC microenvironment (40), may directly
863 sustain Treg lipogenic program. Indeed, OX40 ligation induces
864 TRAF6 activation in CD4 T cells (41), and TRAF6 is a crucial
865 driver of lipid metabolism in memory T cell development (42);
866 therefore, we would suggest that the signal of OX40, possibly
867 together with other TNF receptors, may sustain the transition
868 of Tregs into the effector phase through the promotion of lipid
869 biosynthetic pathways.

870 Our data show that Tregs accumulate intracellular lipids not
871 only in cancer but also in a context of inflammation-driven expan-
872 sion (early cholangitis in *Mdr2*^{-/-} mice), but not in hepatic steatosis
873 in the absence of inflammation (HCVTg mice at advanced age).
874 Therefore, Treg lipid loading seems to occur as a result of Treg-
875 intrinsic pathways (such as OX40 signaling) and irrespective of
876 the lipid profile in the surrounding tissue. Intriguingly, we could
877 find high expression of PPAR γ also in TUM-Tregs. PPAR γ is a
878 nuclear factor that, in response to lipid ligands, controls a pro-
879 gram of FA uptake, glucose uptake and lipogenesis in the adipose
880 tissue, but also regulates a variety of other functions in other or-
881 gans and in immune cells (43). In VAT-Tregs, PPAR γ was shown to
882 drive CD36 expression and FA incorporation; however, treatment
883 of PPAR γ -transfectants with a synthetic agonist induced not only

884 *Cd36* gene, but also other genes involved in lipid metabolism, 885
886 such as *Scd1* and *Cpt1a* among others (19). Therefore, PPAR γ 887
888 may be active in both VAT- and TUM-Tregs, driving a similar 889
890 program of lipid rearrangement that is specifically instrumental 891
892 to Treg expansion. Whether certain endogenous FAs, generated 893
894 by lipogenesis, may bind (at relatively low affinity) and activate 895
896 PPAR γ , is still controversial (44). It is reasonable to think that 897
898 PPAR γ may be activated as a consequence of increased FAS in 899
900 TUM-Tregs: in line with this hypothesis, we found a preferential 901
902 co-segregation of *Pparg* and *Acacb* expression in TUM-Tregs. 903

904 At the systemic level, starvation leads to a transient Treg 905
906 expansion through the mediation of the leptin/mTOR axis (13): 907
908 indeed, conditions of poor nutrient availability may shift the bal- 909
910 ance from growth and defense to homeostasis maintenance, and 911
912 switch the immune status from inflammation to immune regula- 913
914 tion. We would propose that the tumor setting might configure as 915
916 a context of nutrient starvation at the microenvironmental level, 917
918 fostering immune regulatory events. Whether leptin deprivation 919
920 plays a role in the local tumor-associated immune dysfunction, 921
922 and especially in Treg expansion, remains to be investigated. 923
924 While the link between obesity, chronic inflammation and cancer 925
926 risk is well established, it is still unclear whether systemic/regional 927
928 nutrient availability, adiposity or metabolic inflammation affects 929
930 immune regulation at the tumor site; however, we have recently 931
932 elucidated that OX40+ Tregs accumulate in the VAT of obese 933
934 human subjects unless affected by colorectal cancer, suggesting 935
936 that adipose tissue of obese individuals may represent a regional 937
938 reservoir for Tregs to be recruited to the tumor site (45). Our 939
940 present results suggest that perturbations in nutrient accessibility 941
942 at the microenvironmental level may not only impact on tumor 943
944 cell but also on immune cell functions. For instance, therapies 945
946 targeting FAS, directly or indirectly, are under development for 947
948 cancer treatment (46): based on our data, this approach may “off- 949
950 target” Tregs, and especially activated tumor-associated Tregs, 951
952 and it may thus act as a double-edge sword against both tumor 953
954 cell proliferation and immune regulation. 955

956 Material and Methods

957 Tumor cell lines

958 MCA38 colon adenocarcinoma and B16F10 metastatic 959
960 melanoma cell lines, kindly provided by M. P. Colombo 961
962 (Fondazione IRCCS Istituto Nazionale Tumori, Milan, Italy), 963
964 were cultured in complete Dulbecco's modified eagle medium 965
966 (DMEM) with high glucose (Gibco) supplemented with 10% 967
968 foetal bovine serum (FBS, Gibco), 2mM L-glutamine (Sigma- 969
970 Aldrich), penicillin/streptomycin, non-essential aminoacids, 971
972 sodium pyruvate (Euroclone), 50 μ M 2-mercaptoethanol (Sigma- 973
974 Aldrich) and 10mM Hepes (Aurogene) at 37 °C in humidified 975
976 5% CO₂ atmosphere. To obtain transplanted tumors, tumor cells 977
978 were subcutaneous injected into the right flank of C57BL/6Ncrl 979
980 wild-type male mice (Charles River Laboratories, Calco, Italy). 981

982 Mouse models

983 HCVTg mice (of the FL-N/35 transgenic lineage, express- 984
985 ing the HCV polyprotein under the albumin promoter (24)) 986
987 were kindly provided by H. Lerat (Institut National de la Santé 988
989 et de la Recherche Médicale, Unité U955, Université Paris- 990
991 Est, Créteil) and backcrossed for more than 10 generations 992
993 to the C57BL/6Ncrl background before analysis. HCVTg and 994
995 C57BL/6Ncrl mice were bred and maintained under conven- 996
997 tional conditions at animal facility of the Dipartimento di Scienze 998
999 Anatomiche, Istologiche, Medico legali e dell'Apparato locomo- 1000
1001 tore (SAIMLAL), in Sapienza Università di Roma, under proto- 1002
1003 cols approved by the Italian Ministry of Health (Authorization 1004
1005 #481/2015-PR). *Mdr2*^{-/-} (FVB.129P2-*Abcb4*^{tm1BorJ}) mice were 1006
1007 obtained from the Jackson Laboratory and maintained under 1008
1009 pathogen-free conditions at the Pasteur Institute animal facility 1009
1010 (Paris). For all the experiments, mice were sacrificed by cervical 1011
1012 dislocation. Tumor-bearing C57BL/6Ncrl mice were sacrificed 1013

953 after two weeks from subcutaneous tumor cell transplantation
954 and tumor volume (mm³) was calculated using the formula:
955 (smaller diameter)² x larger diameter. For OX40 stimulation in
956 vivo, C57BL/6 mice received intraperitoneal injection of 500 µg
957 of anti-OX40 mAb (OX86, BioXcell); after 6 days, flow cytometry
958 analysis was performed on splenocytes.

959 *In vivo uptake assays*

960 To test in vivo uptake of glucose and lipids, tumor-bearing
961 mice received a single i.p. injection of 50 µg Bodipy-conjugated
962 palmitate (Bodipy FL C16, Life Technologies), 1 hour before
963 analysis, or a single i.p. injection of 100 µg fluorescent glucose
964 analogue 2-NBDG (2-(N-(7-Nitrobenz-2-oxa-1,3-diazol-4-yl)A-
965 mino)-2-Deoxyglucose, Life Technologies), 15 minutes before
966 analysis.

967 *TOFA/Etx intra-tumor treatments*

968 To test TOFA or Etx anti-tumor activity in vivo, tumor-
969 bearing mice received repeated intra-tumor injections of 250 µg
970 TOFA (Sigma) or 200 µg Etx (Sigma) at days 5, 8, 11 and 13
971 after s.c. tumor transplantation. As control, DMSO or water were
972 administered, respectively; mice were sacrificed at day 14 for
973 analysis.

974 All in vivo experiments were authorized by the institutional
975 ethical committees and performed in accordance to institutional
976 guidelines and national laws.

977 *Lymphocyte extraction from murine tissues*

978 Murine tumor samples were mechanically dissociated using
979 gentleMACS Octo Dissociator according to manufacturer's in-
980 structions (Miltenyi Biotec), and mononuclear cells were en-
981 riched through 40/80 Percoll (GE Healthcare) density gradient,
982 collecting cells at the interface between 40% and 80% Percoll
983 solutions. The same protocol was applied for hepatic lymphocyte
984 extraction. Splenocytes were obtained by spleen mechanical dis-
985 sociation on 70-µm filter followed by ACK lysis buffer (Gibco) in-
986 cubation for 4 minutes at 4°C. Lymph nodes were mechanically dis-
987 sociated on 70-µm filter. Cells were collected in complete RPMI-
988 1640 Dutch modified medium containing 10% FBS (Gibco), 2
989 mM L-glutamine (Sigma-Aldrich), penicillin/streptomycin, non-
990 essential aminoacids, sodium pyruvate (Euroclone), and 50 µM
991 2-mercaptoethanol (Sigma-Aldrich).

992 *Flow cytometry and PrimeFlow RNA Assay*

993 Bodipy (BODIPY™ 505/515, 4,4-Difluoro-1,3,5,7-
994 Tetramethyl-4-Bora-3a,4a-Diaza-s-Indacene, ThermoFisher
995 Scientific, catalog number D3921) is a cell membrane-permeant
996 fluorophore specific for neutral lipid stores, known to stain
997 lipid droplets in CD8 T cells (16). For Treg analysis, cells
998 were incubated 15 minutes at 37°C with Fixable Viability
999 Dye eFluor780 (eBioscience) plus Bodipy or Nile Red
1000 (Enzo Life Sciences) dyes for neutral lipid staining; then,
1001 staining with the following antibodies for surface antigens was
1002 performed for 20 minutes at 4°C: CD4 BrilliantViolet605,
1003 CD25 BrilliantViolet510 and CD134 (OX-40) BrilliantViolet421
1004 (all from BioLegend), CD36 APC (BD Biosciences), Glut1
1005 PE (Novus Biologicals). Finally, intracellular staining with
1006 Foxp3 PE-eFluor610 and Ki-67 PE-Cyanine7 (both from
1007 eBioscience) was performed for 30 minutes at RT, after cell
1008 fixation and permeabilization for 30 minutes at 4°C, using
1009 Foxp3/Transcription Factor Staining Buffer Set according to
1010 manufacturer's instructions (eBioscience). For CD8 analysis,
1011 cells were restimulated 4 hours with Cell Stimulation Cocktail
1012 plus Protein Transport Inhibitor (eBioscience), stained with
1013 Viability Dye eFluor780, CD8a BrilliantViolet785 (BioLegend),
1014 CD44 BrilliantViolet510 (BioLegend), fixed/permeabilized
1015 with Cytfix/Cytoperm solution according to manufacturer's
1016 instructions (BD Biosciences), and finally stained with IFNγ PE
1017 (eBioscience), TNFα PECy7 (Biolegend).

1018 PrimeFlow RNA Assay (Affymetrix/eBioscience) was per-
1019 formed according to manufacturer's instructions. After surface
1020

staining and fixation/permeabilization, target probe hybridization
was performed using type 1 (AlexaFluor647-conjugated) probes
for *Pparg* and *Eno1* mRNAs, type 4 (AlexaFluor488-conjugated)
probes for *Cpt1a* and *Hk2* mRNAs, and type 6 (AlexaFluor750-
conjugated) probes for *Acacb* and *Gapdh* mRNAs. As a positive
control, mouse beta Actin (*Actb*), conjugated with each probe
type, was used. Cells were incubated for 2 hours with the target
probes at 40°C in a thermal incubator for microtubes, then were
incubated with the PreAmplification (PreAmp) reagent for 2
hours and the Amplification (Amp) reagent for an additional 2
hours at 40°C. After signal amplification, cells were incubated
with label probes at 40°C for 1 hour and finally cells were washed
and resuspended in staining buffer for acquisition.

Data were acquired on LSR Fortessa (Becton Dickinson) and
analyzed with FlowJo software (Tree Star Inc, version 10.1r5).

1036 *In vitro proliferation assays*

1037 CD4+CD25+ Tregs and CD4+CD25- Tconvs were mag-
1038 netically purified from splenocytes of wild-type C57BL/6 mice
1039 using the CD4+CD25+ Regulatory T Cell Isolation Kit (Miltenyi
1040 Biotec), labeled with 10 µM cell proliferation dye eFluor670
1041 (eBioscience) by incubation for 15 minutes at 37°C in complete
1042 RPMI Dutch modified medium, and cultured or co-cultured at
1043 1:1 ratios in presence of equal numbers of irradiated (3500 rad)
1044 autologous splenocytes plus soluble anti-CD3 (1 µg/ml, eBio-
1045 science) and IL-2 (100 IU/ml, Roche), and in presence or not of
1046 TOFA (Sigma), Etomoxir (Etx, Sigma) or 2-deoxy-D-glucose (2-
1047 DG, Sigma) at different concentrations. After 4 days of culture,
1048 analysis of eFluor670 dilution and Bodipy incorporation was
1049 performed in gated Foxp3+ (Tregs) or Foxp3- (Tconvs) by flow
1050 cytometry.

1051 *Adoptive Treg transfer and suppression of concomitant immu- 1052 nity in vivo*

1053 CD4+CD25+ Tregs were magnetically purified from spleno-
1054 cytes of wild-type C57BL/6 mice, and cultured for 40 hours in
1055 complete RPMI medium with coated anti-CD3 (1 µg/ml) plus IL-
1056 2 (100 IU/ml), in the presence of TOFA (5 µg/ml) or DMSO as
1057 control. Recipient mice first received a primary tumor inoculum
1058 of 10⁵ MCA38 cells subcutaneously in the right flank. After
1059 10 days, when primary tumors were palpable, mice received a
1060 challenge inoculum in the opposite flank, containing 10⁵ MCA38
1061 cells, either alone or admixed at 1:1 ratio with Tregs, prepared
1062 as described above. After further 13 days, flow cytometry was
1063 performed in lymphocytes extracted from draining lymph nodes
1064 and tumors.

1065 *Seahorse analysis*

1066 OCR and ECAR real-time measurements were performed
1067 using an XFe-96 Extracellular Flux Analyzer (Seahorse Bio-
1068 science) in basal condition in XF media and also in response to 5
1069 µM oligomycin, 1.5 µM of carbonylcyanide-4- (trifluoromethoxy)-
1070 phenylhydrazine (FCCP), or 1 µM of Antimycin and Rotenone
1071 (all from Sigma-Aldrich) for OCR profile, whereas in response
1072 to 10 mM glucose, 5 µM oligomycin and 100 mM 2-DG (all
1073 from Sigma-Aldrich) for ECAR analysis. Basal OxPhos (before
1074 oligomycin addition) and ATP-linked OxPhos (as difference be-
1075 tween basal OCR and oligomycin-induced OCR) were calculated
1076 from the OCR profile, as indices of mitochondrial respiratory
1077 function; basal glycolysis (after the addition of glucose), maxi-
1078 mal glycolysis (after the addition of oligomycin), and glycolytic
1079 capacity (as difference between oligomycin-induced ECAR and
1080 2-DG-induced ECAR) were calculated from the ECAR profile
1081 as indices of glycolytic pathway activation.

1082 *Metabolomic analysis*

1083 CD4+CD25+ Tregs and CD4+CD25- Tconvs were magnet-
1084 ically purified from pooled splenocytes and tumors of C57BL/6
1085 mice (n=15) using the CD4+CD25+ Regulatory T Cell Isolation
1086 Kit (Miltenyi Biotec). Pellets were then resuspended in 250µl
1087 methanol/acetonitrile 1:1 containing [U-¹³C₆]-Glucose- 1ng/µl
1088

1089 (internal standard, Sigma Aldrich, 389374) and lysed by Tissue
1090 lyser 3 min at the highest frequency. Lysates were spun at 20,000g
1091 for 5 min at 4°C. Supernatants were then passed through a
1092 regenerated cellulose filter, dried and resuspended in 100µl of
1093 MeOH for subsequent analysis. Metabolomic data were obtained
1094 on an API-4000 triple quadrupole mass spectrometer (AB Sciex)
1095 coupled with a HPLC system (Agilent) and CTC PAL HTS au-
1096 to-sampler (PAL System). The identity of all metabolites was con-
1097 firmed using pure standards. Quantification of different metabo-
1098 lites was performed with a liquid chromatography/tandem mass
1099 spectrometry (LC-MS/MS) method using cyano-phase LUNA
1100 column (50mm x 4.6mm, 5µm; Phenomenex). Methanolic sam-
1101 ples were analyzed by a 5 min run in negative ion mode with 30
1102 MRM transitions. The mobile phase A was: 5 mM ammonium
1103 acetate pH 7.00 in MeOH. The gradient was 100%A for all the
1104 analysis with a flow rate of 500µl/min. MultiQuant™ software
1105 (version 3.0.2) was used for data analysis and peak review of
1106 chromatograms. Quantitative evaluation of all metabolites was
1107 performed based on calibration curves with pure standards, then
1108 data were normalized on micrograms of proteins analyzed by
1109 Bradford method.

1110 *XTT cytotoxicity assay*

1111 To assess the cytotoxic effects of TOFA and Etx on tumor
1112 cells in vitro, MCA38 cells were seeded at the concentration of
1113 5×10^3 cells/well into 200 µl of complete DMEM medium in flat-
1114 bottomed 96-well plate, in presence of different concentrations
1115 of TOFA and Etx, or DMSO and water as respective controls.
1116 Conditions without cells or cell treatment with 0.05% saponin
1117 (Sigma-Aldrich) were used as negative and positive control, re-
1118 spectively. After 24 hours of culture, the analysis of cell viability
1119 was performed with XTT Cell Proliferation Assay Kit (Cayman
1120 Chemical), and absorbance was measured with Multiskan FC
1121 (Thermo Scientific).

1122 *Human samples*

1123 Peripheral blood from healthy volunteers was obtained from
1124 buffy coats of healthy blood donors, anonymously provided by the
1125 Immunohematology and Transfusion Center of Policlinico Um-
1126 berto I. Peripheral blood and liver specimens were obtained from
1127 5 patients with chronic hepatitis C, cirrhosis and hepatocellular
1128 carcinoma (4 males and 1 female, aged 53-80 years), undergoing
1129 surgery or liver transplantation at Istituto Nazionale dei Tumori
1130 "Regina Elena" or "Sapienza" Università di Roma - Policlinico
1131 Umberto I. Human studies have been performed in accordance
1132 to the ethical guidelines of the 1975 Declaration of Helsinki and
1133 were approved by the Institutional Ethical Committee (Autho-
1134 rization: RIF.CE: 4259). Informed consent was obtained from all
1135 patients.

1136 *Human Treg/Tconv sorting and gene expression analysis*

1137 Liver fragments from patients were perfused and grossly
1138 mashed with HBSS (Euroclone) containing 0.5 mg/ml collage-
1139 nase IV (Sigma-Aldrich) and 50 ng/ml DNase I (Worthing-
1140 ton). Samples were incubated 10 minutes at 37°C and then me-
1141 chanically disaggregated into single cell suspensions. Mononu-
1142 clear cells were enriched by Lympholyte (Cederlane) density
1143 gradient, also from patient PB. T cells were enriched from
1144 mononuclear cells using Pan T cell Isolation kit II (Miltenyi
1145 Biotec) and, after treatment with Fixable Viability Dye eFluor780
1146 (eBioscience) and with FcBlock (eBioscience) for 15 minutes
1147 at 4°C, were stained for surface markers: dump channel (CD14
1148 APCeFluor780, CD56 APCeFluor780, CD8 APCeFluor780, all
1149 from eBioscience), CD4 AlexaFluor488 (eBioscience), OX40
1150 PE (ACT35, BD), CD45RA PerCPCy5.5 (BioLegend), CD127
1151 PECy7 (BioLegend) and CD25 APC (BioLegend). Therefore,
1152 OX40+ and OX40- subsets, in gated CD45RA low Tregs and
1153 Tconvs, were sorted using a FACSAria II (Becton Dickinson).

Total RNA was isolated using mirVana Isolation Kit (Am-
bion) following the standard protocol. Briefly, the lysates were
extracted once with Acid-Phenol Chloroform and further purified
to yield total RNA. Extracted RNA was quantified with Ribo-
Green Quantitation Kit (Molecular Probes) on an Infinite F200
plate reader (Tecan Trading AG). All extracted RNA samples
were quality controlled for integrity with 2100 Bioanalyzer (Agi-
lent Technologies).

Gene expression of whole transcriptomes was performed
according to the standard protocol. Total RNA was isolated,
quality controlled and quantified as described above; for each
sample 100 ng of total RNA were reverse transcribed accord-
ing to the Illumina TotalPrep RNA Amplification kit (Ambion)
and biotinylated cRNA was generated by in vitro transcription.
Washing, staining and hybridization were performed according to
the standard Illumina protocol: briefly, 750 ng of cRNA of each
sample in a final volume of 15 µl were hybridized onto Illumina
HumanHT-12 v4 Expression BeadChip arrays. Hybridization and
scanning were performed according to the manufacturer's indi-
cations on an Illumina iScan System and data were processed
with BeadStudio v.3; arrays were quantile normalized, with no
background subtraction, and average signals were calculated on
gene-level data for genes whose detection p-value was lower than
0.001 in at least one sample.

1181 *Statistical analysis*

1182 Statistical analysis was performed using Prism software (ver-
1183 sion 6.0, GraphPad). Two tailed-unpaired Student's t test was used
1184 to analyze in vitro data and to compare ex vivo data from Tregs
1185 and Tconvs in different samples, while two tailed-paired Student's
1186 t test were applied to compare ex vivo data from Tregs and
1187 Tconvs in the same sample. Every in vitro and ex vivo assay was
1188 performed in duplicate, triplicate, or quadruplicate when possible
1189 and for all experiment the number of repetitions is indicated in
1190 the figure legend. In all graphs, bars show means \pm SEM. In all
1191 tests, $P < 0.05$ was considered statistically significant.

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