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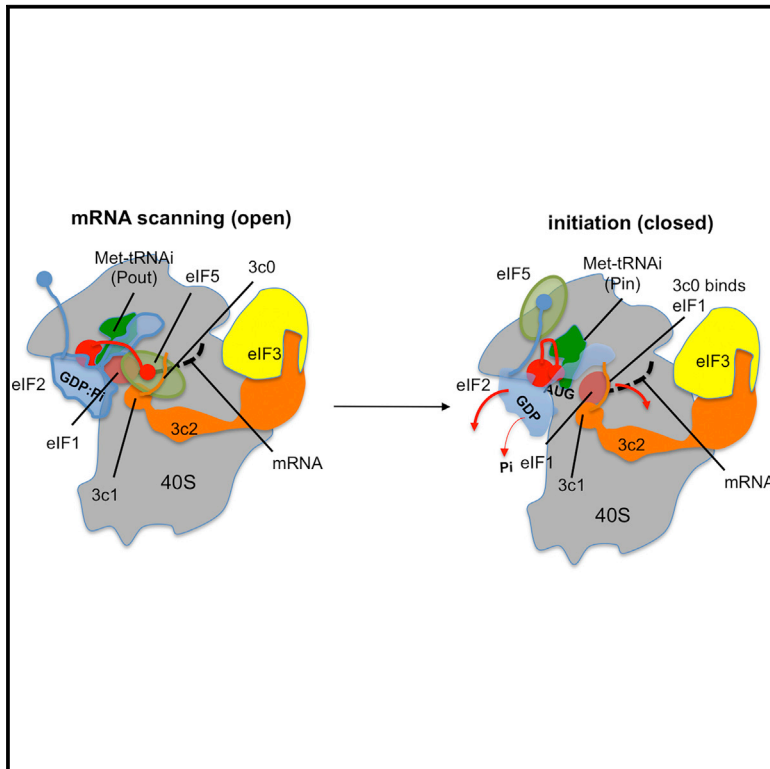


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Cell Reports

Molecular Landscape of the Ribosome Pre-initiation Complex during mRNA Scanning: Structural Role for eIF3c and Its Control by eIF5

Graphical Abstract



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In Brief

During translation initiation, eIF3 binds the solvent-accessible side of the 40S ribosome. Obayashi et al. propose that the N-terminal domain of eIF3c reaches into the decoding center to not only anchor the gate-keeper eIF1 but also facilitate eIF1 release on AUG selection. eIF5 appears to play a role in this regulation.

Highlights

- eIF3c N-terminal domain is divided into three regions, 3c0, 3c1, and 3c2
- 3c1 and eIF5 anchor eIF1 to the 40S ribosome during mRNA scanning
- On AUG, 3c0 binds eIF1 ribosome-binding site, facilitating eIF1 release
- eIF5 prevents 3c0 from binding eIF1 before AUG selection

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Molecular Landscape of the Ribosome Pre-initiation Complex during mRNA Scanning: Structural Role for eIF3c and Its Control by eIF5

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SUMMARY

During eukaryotic translation initiation, eIF3 binds the solvent-accessible side of the 40S ribosome and recruits the gate-keeper protein eIF1 and eIF5 to the decoding center. This is largely mediated by the N-terminal domain (NTD) of eIF3c, which can be divided into three parts: 3c0, 3c1, and 3c2. The N-terminal part, 3c0, binds eIF5 strongly but only weakly to the ribosome-binding surface of eIF1, whereas 3c1 and 3c2 form a stoichiometric complex with eIF1. 3c1 contacts eIF1 through Arg-53 and Leu-96, while 3c2 faces 40S protein uS15/S13, to anchor eIF1 to the scanning pre-initiation complex (PIC). We propose that the 3c0:eIF1 interaction diminishes eIF1 binding to the 40S, whereas 3c0:eIF5 interaction stabilizes the scanning PIC by precluding this inhibitory interaction. Upon start codon recognition, interactions involving eIF5, and ultimately 3c0:eIF1 association, facilitate eIF1 release. Our results reveal intricate molecular interactions within the PIC, programmed for rapid scanning-arrest at the start codon.

INTRODUCTION

Ribosomes initiate translation with levels of stringency varying between bacteria (low) and eukaryotes (high) (Asano, 2014).

The high accuracy of initiation in eukaryotes results from suppressing initiation from non-AUG codons like GUG and UUG. This stringency is imposed partly by eukaryotic initiation factors (eIFs) that bind the small (40S) ribosomal subunit in the 43S preinitiation complex (PIC), i.e., eIF1A, eIF1, eIF2, eIF3, and eIF5 (Asano, 2014; Hinnebusch, 2014). Like its bacterial counterpart IF1, eIF1A binds the 40S A-site. The other four factors engage in numerous mutual interactions to form the multifactor complex (MFC) with Met-tRNA^{Met} bound to eIF2-GTP in the ternary complex, whereby MFC can be isolated free of ribosomes from various eukaryotes (Asano et al., 2000; Dennis et al., 2009; Meleppattu et al., 2015; Sokabe et al., 2012). eIF4F, comprising m⁷G-cap binding subunit eIF4E, RNA helicase eIF4A, and scaffold eIF4G, mediates attachment of the mRNA 5' end to the PIC in its open, scanning-competent conformation (Kumar et al., 2016). A key event in start codon selection is dissociation from the 40S of eIF1, a gatekeeper molecule that maintains the open conformation of the PIC (Pestova and Kolupaeva, 2002; Saini et al., 2010). During scanning, the eIF1 physically opposes full accommodation of tRNA_i in the P-site, keeping it in the P_{OUT} conformation (Lomakin and Steitz, 2013; Rabi et al., 2011; Weissner et al., 2013). Once tRNA_i base pairs to the AUG start codon, eIF1 is released, Met-tRNA_i is fully accommodated in the P-site (P_{IN} state), and the PIC adopts the closed conformation incompatible with scanning. The resulting 40S initiation complex is ready for subsequent 60S subunit joining.

In this work, we examine the structural role of the N-terminal domain (NTD) of the eIF3c-subunit of eIF3, a crucial binding partner of eIF1 and eIF5 in the MFC, and key regulator of start codon



selection (Asano et al., 2000, 2001a; Karásková et al., 2012; Phan et al., 1998; Valásek et al., 2004). eIF3 is a multisubunit complex (Asano et al., 1997) that binds the solvent-accessible side of the 40S (Srivastava et al., 1992). Cross-linking and integrated modeling studies suggest that eIF3c-NTD extends into the 40S decoding center proximal to eIF1 (Erzberger et al., 2014). eIF5 is the GTPase activating protein for eIF2 (Asano et al., 2001b; Huang et al., 1997). Independently of the catalytic NTD, the eIF5 C-terminal domain (CTD) interacts with eIF1A, eIF2 β , eIF3c, and eIF4G at various stages of initiation (Luna et al., 2012, 2013; Reibarkh et al., 2008; Singh et al., 2012; Yamamoto et al., 2005). While an initial cryoelectron microscopy (cryo-EM) study revealed density potentially corresponding to eIF5-CTD facing eIF1 and eIF2 in the PIC (Hussain et al., 2014), this was not observed in more recent PIC structures (Lácer et al., 2015). Thus, the location and structural role of eIF5-CTD in the PIC also remains unclear.

Genetic studies have revealed that eIF3c-NTD contains two distinct elements with opposing roles in initiation accuracy. Box12 is required for accurate initiation, and substitution mutations in this element increase non-AUG initiation (for the *Sui*[−] or suppressor of initiation codon mutation phenotype). The Box6 element is required for initiation at non-AUG codons, and substitutions in Box6 suppress effects conferred by a *Sui*[−] mutation (for the *Ssu*[−] or suppressor of *Sui* phenotype) (Karásková et al., 2012; Valásek et al., 2004). Henceforth, Box6 and Box12 are designated as an *Ssu*⁺ (Box6_{Ssu+}) and a *Sui*⁺ element (Box12_{Sui+}), respectively. Certain Box6 or Box12 mutations decrease eIF1 binding to the eIF3c-NTD, suggesting that the eIF3c-NTD helps to stabilize eIF1 in the PIC not only during mRNA scanning, but also during the switch to the closed state upon start codon selection. Herein, we employed a battery of biophysical methods including nuclear magnetic resonance (NMR) spectroscopy to dissect eIF3c-NTD into three units, 3c0, 3c1, and 3c2 and locate the latter two within the recently solved cryo-EM PIC structure (Erzberger et al., 2014). Based on physical interaction studies involving eIF1, eIF3c-NTD, and eIF5, we propose that, by interacting with the N-terminal unit 3c0, eIF5 modulates the ability of eIF3c-NTD to either anchor or release eIF1. Our model explains distinct contributions of eIF3c Box6_{Ssu+} and Box12_{Sui+} to the accuracy of start codon selection in vivo.

RESULTS

Functional Dissection of eIF1-Binding Elements in eIF3c-NTD

To map eIF1 binding sites in the eIF3c-NTD, we divided the latter into three regions: 3c0 encompassing amino acids (aa) 1–58, including the conserved N terminus required for eIF5 binding (Karásková et al., 2012) and most of Box6_{Ssu+}; 3c1 encompassing aa 59–87, which contains a conserved hydrophobic segment; and 3c2 comprising aa 88–163, including predicted α helices (<http://bioinf.cs.ucl.ac.uk/psipred/>) and Box12_{Sui+} (Figures 1A and S1A). GST fusions to eIF3c-NTD fragments with different combinations of these regions (eIF3c-A to -G, Figure 1A) were tested for eIF1 binding using GST pull-down assays. Fragment eIF3c-D_{58–163} essentially covers the previously determined minimal eIF1-binding site (aa 60–137) (Karásková et al., 2012).

The strongest eIF1 binding was observed with eIF3c-A_{1–163}, -B_{36–163}, -C_{36–87}, and -F_{1–87}, which all include the C-terminal half of 3c0 and the entire 3c1 (Figures S1B, lanes 2, 3, 6, and 8, and S1C). Isothermal titration calorimetry (ITC) assays demonstrated apparent K_d values of $\sim 1 \mu\text{M}$ for these constructs (Figures 1A and S2A) with SDs of $<15\%$ ($n = 3$, Figure S2B). eIF3c-D_{58–163}, containing regions 3c1 and 3c2, exhibited weaker association with eIF1 (Figure S1B, lane 4) with an apparent K_d of $\sim 8 \mu\text{M}$ (Figures 1A, S2A, and S2B). eIF3c-D_{58–163} regions therefore bind eIF1 with a significantly lower affinity than the constructs with regions 3c0–3c1 ($p < 0.006$, $n = 3$). In contrast, the two NTD segments lacking 3c1, eIF3c-E_{87–163} (3c2), and eIF3c-G_{1–58} (3c0) did not appear to interact with eIF1 in GST pull-down assays (Figure S1B, lanes 5 and 7), but displayed appreciable binding when the eIF1 concentration was increased ~ 50 -fold to $\sim 30 \mu\text{M}$ (Figure 1B, lanes 5 and 9, eIF1 detected by anti-eIF1; Figure S1C, lanes 3 and 7, eIF1 indicated by arrowheads in Coomassie staining). Note that in Figure 1B, amounts of eIF1 bound to GST-eIF3c-E_{87–163} (lane 9) and eIF3c-G_{1–58} (lane 5) are $<10\%$ of that bound to GST-eIF3c-F_{1–87} with 3c0 and 3c1 (lanes 7 and 10; where 10% and 90% of the pull-down fraction were loaded. Anti-eIF1 signal in lane 10 is saturated due to overloading). Consistent with the pull-down results, the K_d for eIF3c-E_{87–163} binding to eIF1 is $>100 \mu\text{M}$ (Figure 1A).

The ITC assay revealed that eIF3c-D_{58–163} forms a stoichiometric complex with eIF1 ($N = 1.0$, Figures 1A and S2B), while other segments containing 3c1 and 3c0 (A_{1–163}, B_{36–163}, C_{36–87}, and F_{1–87}) display N values (number of eIF3c molecules bound per eIF1 molecule) significantly less than 1.0 ($p < 0.03$, $n = 3$). These results suggest that eIF1 has more than one binding site for eIF3c regions 3c0 and 3c1.

Based on these results, we identify 3c1 as the core eIF1-binding site in eIF3c-NTD. Low-affinity eIF1 binding by flanking region 3c0 containing Box6_{Ssu+} contributes to the high-affinity eIF1 binding ($\sim 1 \mu\text{M}$) by fragments containing 3c1 and 3c0, likely through interaction with more than one site on eIF1. Because we failed to generate an eIF3c segment containing only 3c1, the contribution of C-terminal flanking 3c2 remained unclear. However, based on the low-affinity eIF1 binding to eIF3c-E_{87–163}, 3c2 containing Box12_{Sui+} likely contributes to the relatively high-affinity binding ($\sim 8 \mu\text{M}$) observed for eIF3c-D_{58–163}.

CSP Mapping with ¹⁵N-eIF3c-NTD Identifies aa Involved in eIF1 Binding

Next, we used NMR chemical shift perturbation (CSP) mapping to delineate eIF3c residues directly involved in eIF1 binding. We first determined the structure of eIF3c-NTD by NMR spectroscopy using [¹³C, ¹⁵N] eIF3c-B_{36–163} segment (see Supplemental Information and Table S3 for details), which demonstrated that the region covering most of 3c2 (residues 105–159) folds into α -helical globule (Figure 1C). The eIF3c backbone resonance assignments were then used for CSP studies. As shown in Figure S3, CSPs induced by eIF1 binding are nearly identical between ¹⁵N-eIF3c-A_{1–163} and -B_{36–163}, which is consistent with our GST-pull down and ITC studies (Figures 1A, S1B, S1C, and S2) (Karásková et al., 2012). We further observed large CSPs for A67 (circled in blue in Figure 1D; indicated by arrow in Figure S3, lower panels), and E51 residues (circled in blue in Figure 1D)

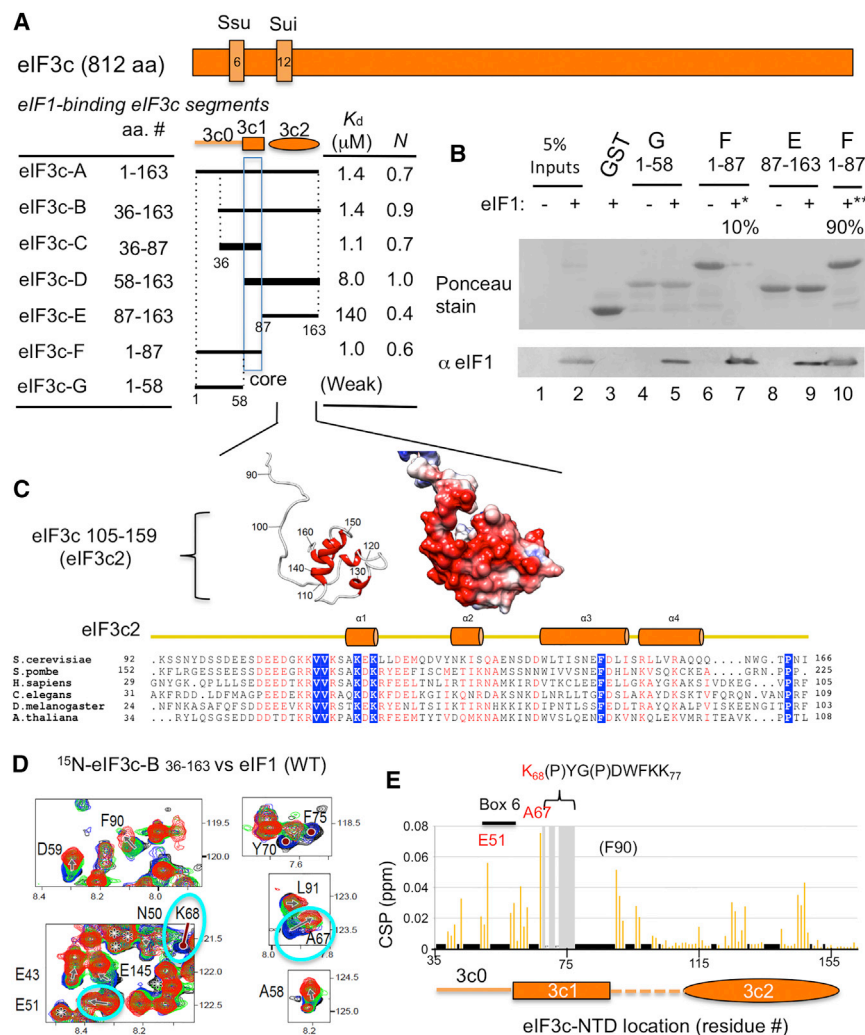


Figure 1. Functional Dissection of the eIF1-Binding Site within eIF3c-NTD

(A) Location of eIF1-binding site in eIF3c primary structure (orange rectangle), highlighting regions of Ssu and Sui mutation sites, Box6 and Box12 (boxes with numbers). Orange schematics below indicate functional elements identified in eIF3c-NTD, 3c0, 3c1, and 3c2. The lines further beneath depict eIF3c deletion constructs used in this study. Dotted lines define the boundaries of eIF3c-NTD regions i-iv (Figure S1A). Table summarizes the results of ITC analysis for eIF1 binding, K_d and N (stoichiometry; number of eIF3c molecules bound to a eIF1 molecule) (see Figure S2). Weak, the weakest binding observed with eIF3c-G in GST pull-down. (B) GST pull-down assay. Approximately 5 μ g of indicated GST-eIF3c fusion proteins (0.15–0.2 nmol) was allowed to bind 70 μ g of recombinant eIF1 (\sim 5 nmol; \sim 25 μ M) in *E. coli* lysates (lanes labeled “+”) and the protein complexes pulled down and analyzed with 5% input amounts of lysates by SDS-PAGE, followed by immunoblotting with anti-yeast eIF1 (bottom) and Ponceau staining (top). –, uninduced *E. coli* lysates were used as a negative control. In lanes 7 (*) and 10 (**), 10% and 90% of the GST-eIF3c-F complex were analyzed, respectively.

(C) Solution structure of eIF3c 105–159 found within yeast eIF3c-NTD (36–163), determined by NMR spectroscopy (see Supplemental Information and Table S3). Ribbon diagram is shown to the left. Right, electrostatic potential distribution (negative in red, neutral in white, and positive in blue) calculated according to Coulomb’s law. Bottom, α helices are aligned with aa sequences of eIF3c_N (Pfam).

(D and E) NMR CSP studies on interaction between 15 N-eIF3c-B_{36–163} and eIF1. (D) Close-up views of ^1H - ^{15}N heteronuclear single-quantum correlation (HSQC) spectra of ^{15}N -eIF3c eIF3c-B in the absence (black) or presence of WT eIF1 (panel 1) (1:0.3 molar ratio in blue, 1:0.6 in green, 1:1 in red). See Figure S3B for the entire spectrum

of ^{15}N -eIF3c eIF3c-B with or without WT eIF1. (E) Chemical shift perturbation, $\Delta\delta$, was computed as described in Supplemental Experimental Procedures and presented for each assigned aa. “P,” proline. Short black bar, unassigned. Shaded, residue with line broadening by eIF1. Three aa showing largest $\Delta\delta$ are labeled. Labeled in red are aa whose CSP were resolved by eIF1 mutations defective in eIF3c binding. See also Figures S1–S4 and Tables S1 and S3.

accompanied by the strong resonance line broadening in the stretch: K₆₈(P)YG(P)DWFKK₇₇ (K68, Y70, and F75 highlighted in Figure 1D; others highlighted in Figures S3A and S3B; prolines are in parentheses). In contrast, all CSPs in 3c2 were minor (<0.04 ppm) (Figure 1E) except for F90, which was considered spurious inasmuch as it was not eliminated by an eIF1 mutation that abolishes interaction with eIF3c (shown below in Figure S4, panel 1). Collectively, these results indicate that the eIF1-binding site on eIF3c NTD resides in the area covering Box6_{Ssu+} of 3c0 (containing E51) and core region 3c1 (aa 58–87, contains A₆₇K(P)YG(P)DWFKK₇₇) (Figures 1A and S1A).

Structure of eIF3c-NTD_{105–159} and Integrated Modeling of eIF3:eIF1:40S Complex Structure Define Two Globular Units within eIF3c-NTD

In a recent cryo-EM study of the eIF1/eIF3/40S complex, which integrated extensive crosslinking information, it was proposed

that the eIF3c-NTD projects from the solvent side along the 40S subunit into the decoding center, where eIF1 is bound (Erzberger et al., 2014). However, structural information for the eIF3c-NTD was lacking. We therefore incorporated NMR structure of eIF3c segment 105–159 (Figure 1C) into the integrated modeling platform and calculated a new localization for the whole eIF3 complex (Figure S5). The resulting localization densities for eIF3c-NTD had a resolution of 18 Å (Figure 2A, left), guided by four high-confidence crosslinks (Figure 2A, right), which is a clear improvement from the 38 Å precision in our previous model (Erzberger et al., 2014). The eIF3c-NTD is resolved into two globular units that span the \sim 60 Å distance between eIF1 and rpS13/uS15 (Figures 2A and S5). The one is located near rpS13/uS15 and was assigned as α -helical globular structure in 3c2 (aa 105–159) shown in Figure 1C. The other is adjacent to eIF1 and, thus, was assigned as the core eIF1-binding region 3c1 (aa 59–87) (Figure 1A). Indeed, recent medium-resolution

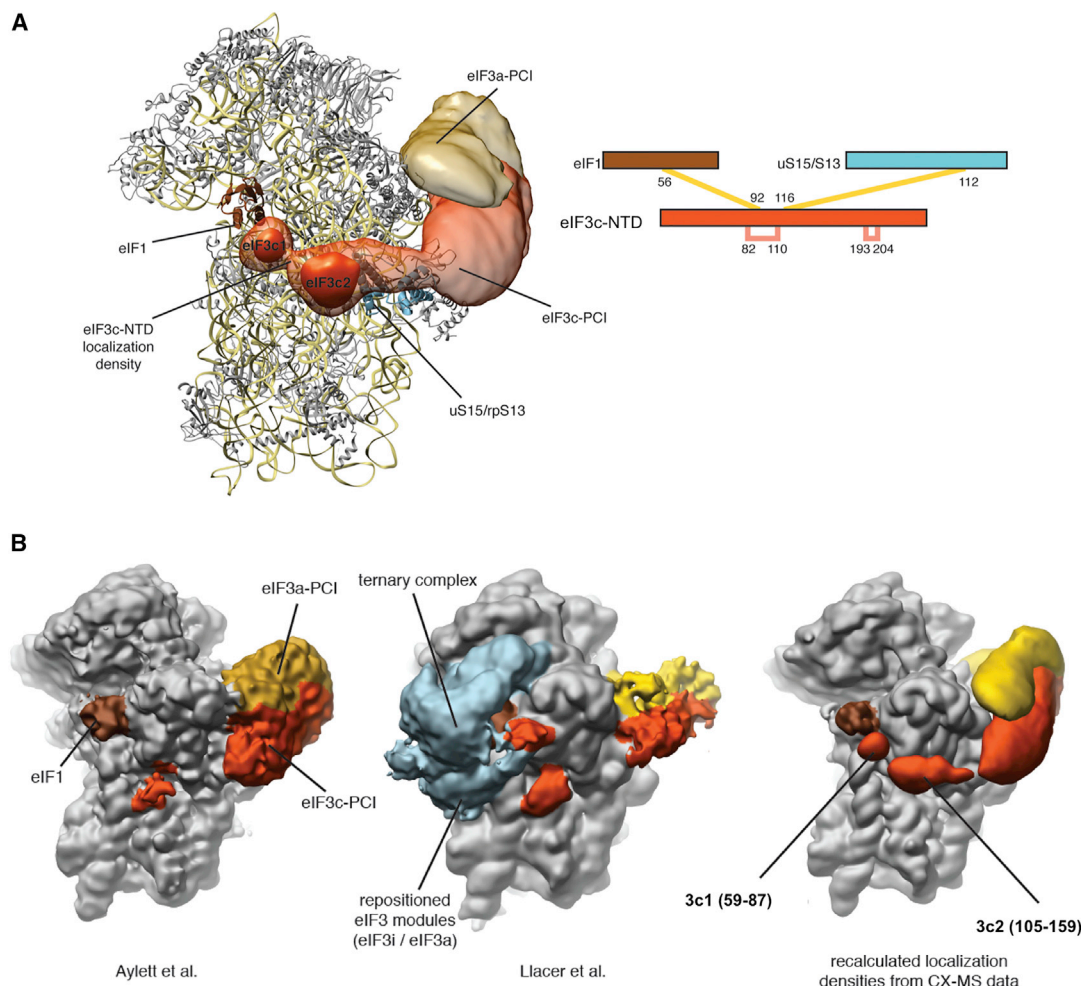


Figure 2. Location of eIF3c2 105–159 Globule within the eIF1:eIF3:40S Structure

(A) Recalculated integrated modeling localization densities (Erzberger et al., 2014) incorporating the eIF3c-NTD NMR structure. Left, 40S subunit shown as a ribbon diagram with overlaid localization densities for eIF3a (gold) and eIF3c (orange-red). A higher contour level of the eIF3c-NTD is used to emphasize the predicted two-domain architecture. Right, eIF3c-NTD-specific cross-links that anchor the globular domain of eIF3c-NTD. Interstrand crosslinks in yellow, intrastrand crosslinks in pink.

(B) Comparison of recent cryo-EM reconstructions of eIF3 complexes (left, Aylett et al., 2015; center, Llácer et al., 2015) with the current localization densities derived from integrative modeling superposed on a 40S-eIF1 structure (right). Densities for eIF3a and eIF3c are colored as in (A), eIF1 is shown in brown and additional densities present in the Llácer et al. structure shown in light blue. In the Erzberger structure, 3c1 is defined as eIF3c aa 58–87, based on the NMR studies in Figures 3 and 4. See also Figure S5.

cryo-EM reconstructions (Aylett et al., 2015; Llácer et al., 2015) reveal densities consistent with the positions of 3c1 and 3c2 (Figure 2B). The N-terminal region 3c0 (Figure 1A) was not localized in the eIF3:eIF1:40S structure, presumably because it cannot bind eIF1 when eIF1 is bound to the 40S subunit (as discussed below). Therefore, integrative modeling that incorporates the NMR structure of eIF3c_{105–159} pinpointed the locations of the 3c1 and 3c2 elements within the PIC, with 3c1 directly contacting eIF1.

NMR Evidence that eIF3c-NTD Segments 3c1–3c2 Interact with a Limited Surface of eIF1 Compatible with 40S Binding

eIF1 comprises an unstructured N-terminal tail (NTT) and a globular domain with a β 1- β 2- α 1- β 3- β 4- α 2- β 5 fold (Fletcher et al.,

1999; Reibarkh et al., 2008) (Figure 3A; Table S4). To determine the eIF1 residues contacted by the 3c1–3c2 units in the complex formed with eIF3c D_{58–163}, we performed CSP experiments using ¹⁵N-eIF1. As shown in Figure 3B and summarized in Figure 3A, strong CSPs were observed for R53, K56, I93, and L96 residues on eIF1 thereby indicating that these residues on eIF1 direct its interaction with eIF3c D_{58–163}. In contrast, resonances corresponding to residues within or nearby the two eIF1 ribosome-binding sites (Martin-Marcos et al., 2013; Rabl et al., 2011), including K60 at the α 1 C terminus and T40/T41 near the β 1- β 2 loop (loop 1), were only marginally affected (Figures 3A and 3B). As summarized in Figure 4A, the eIF3c-D_{58–163}-binding site on eIF1 comprises the N-terminal and central portions of α 1 and the adjacent hydrophobic area containing I93 (residues

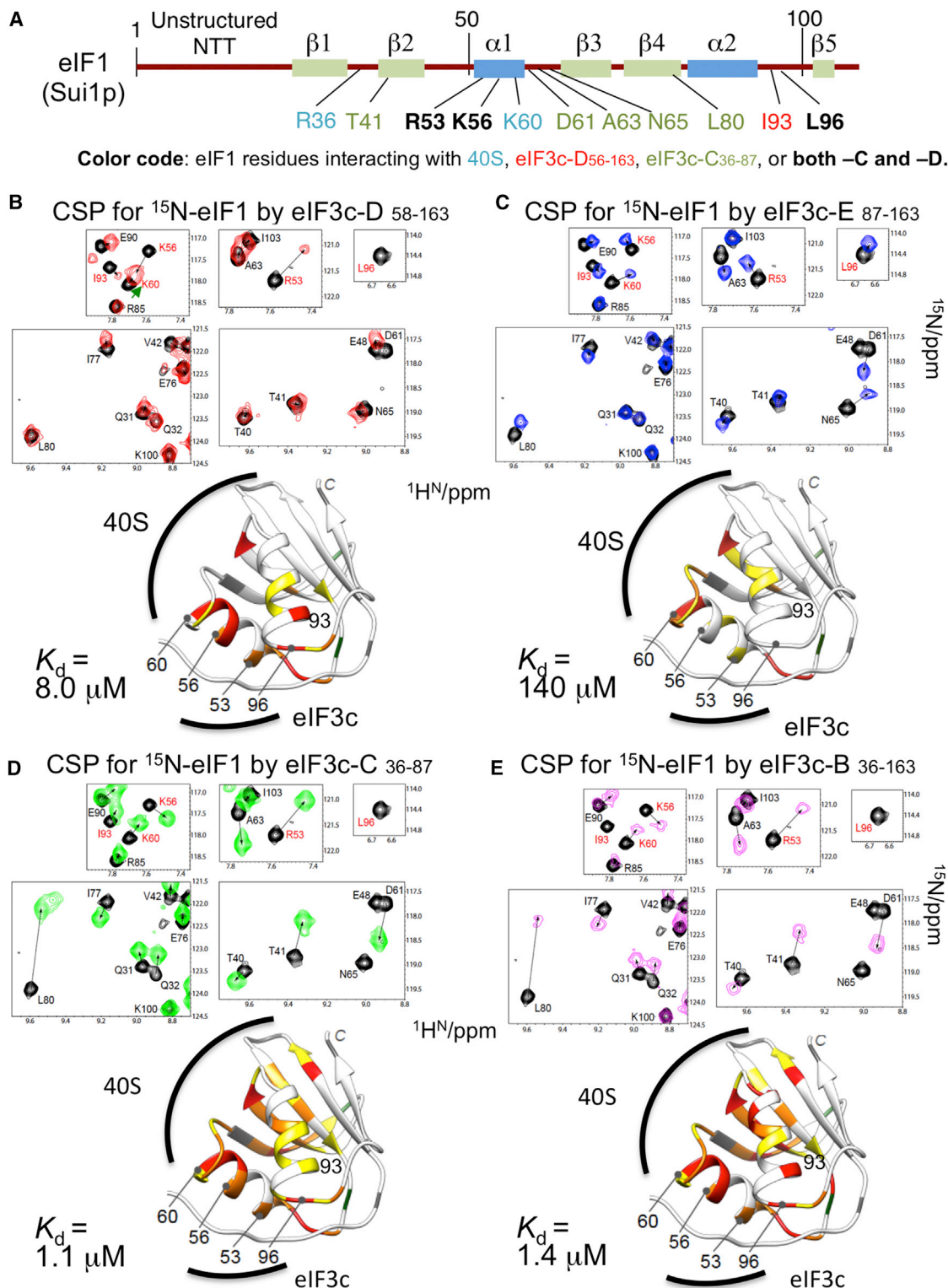


Figure 3. NMR CSP Mapping of eIF3c Binding Site on eIF1

(A) Primary structure of yeast eIF1 (brown horizontal line) with boxes indicating secondary structure elements. aa whose resonance was shifted due to addition of distinct eIF3c fragments are shown in colors based on the code on the bottom.

(B–E) Top, CSP of ^{15}N -eIF1 resonances caused by eIF3c-D_{58–163} (B), eIF3c-E_{78–163} (C), eIF3c-C_{36–87} (D), and eIF3c-B_{36–163} (E) were highlighted with arrows in the specified areas of ^1H - ^{15}N HSQC spectra. The spectra taken in the presence and absence of eIF3c fragments (1:1.2) are shown in color and gray, respectively. The

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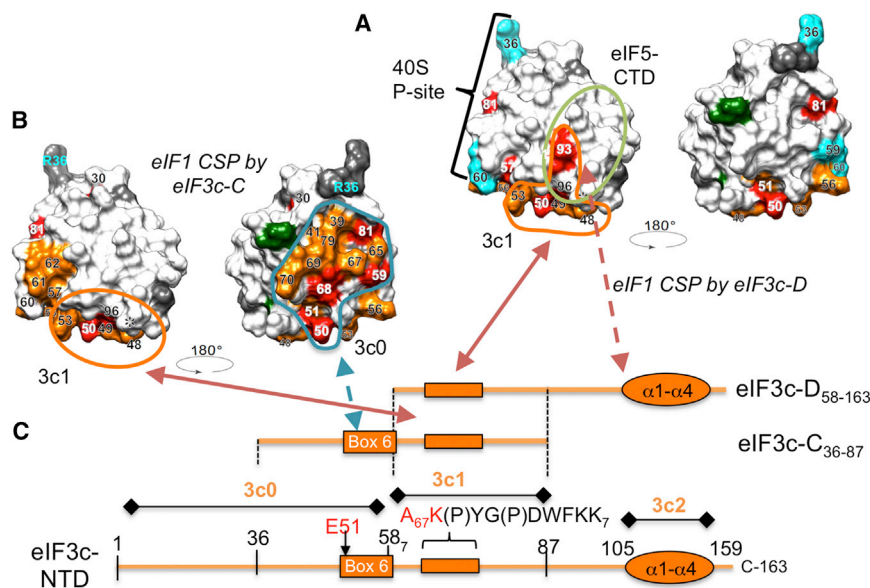


Figure 4. Summary of eIF3c:eIF1 Interaction Models

(A and B) Left, the eIF1 residues showing strong CSP or line broadening by eIF3c-D₅₈₋₁₆₃ (A) or eIF3c-C₃₆₋₈₇ (B) are presented (with unassigned and proline residues) by the same colors on the space-filled model viewed from the same angle as in Figures 3B and 3D, respectively. For simplicity, however, residues showing moderate CSP (yellow in Figure 3) are not presented. Residues 36, 59, and 60 known to contact the ribosome are shown in cyan (Rabl et al., 2011). 3c1- and eIF5-binding sites implicated in the scanning PIC are circled with orange and dark green lines, respectively. Right, the space-filled model of eIF1 rotated 180° relative to the model to the left. In (B), 3c0-binding site is indicated by blue line.

(C) Schematic on the bottom (horizontal orange line) describes the primary structure of eIF3c-NTD (aa 1–163) with orange boxes indicating the locations of Box 6 and the area of line-broadening by eIF1. Orange oval, α -helical globule of 3c2. Lines above denote the locations of eIF3c segments, D₅₈₋₁₆₃ and C₃₆₋₈₇, used for the CSP studies on ¹⁵N-eIF1. Arrows indicate the proposed interactions between defined areas of eIF3c and eIF1.

painted red or orange). In agreement with this, the previous EM study showed that K56 on eIF1 crosslinks with K92 on eIF3c, which is located in the vicinity of 3c1 region (Figure 2A). Because eIF1 interacts with the ribosome via residues K59 and K60 at the C terminus of α 1, and R36 in loop 1 (residues painted cyan in Figure 4A), stoichiometric eIF1 binding to the 3c1-3c2 segment of eIF3c appears to be compatible with eIF1:ribosome association.

By comparing CSPs between eIF3c-D₅₈₋₁₆₃ (Figure 3B) versus eIF3c-E₈₇₋₁₆₃, containing only 3c2 (Figure 3C), it is clear that eliminating the 3c1 core eIF1-binding region dramatically reduces affinity of eIF3c NTD for eIF1 (Figures 1A, S1, and S2). However, weak/moderate CSPs (0.05 < ppm < 0.1) were observed in the N-terminal half of α 1 and β 4 of eIF1 (Figure 3C), while stronger CSPs were located in the C terminus of α 1 (K60) and the α 1- β 3 loop (N65). Considering the small *N* value (0.4) observed for E₈₇₋₁₆₃ in ITC assays indicating multiple binding sites on eIF1 (Figures 1A and S2B), we suggest that eliminating 3c1 disrupts the stoichiometric binding to eIF1 seen for eIF3c-D₅₈₋₁₆₃, which allows isolated 3c2 (E₈₇₋₁₆₃) to engage in weak and likely non-physiological interactions with multiple surfaces on eIF1.

In conclusion, eIF3c-D₅₈₋₁₆₃ containing 3c1 and 3c2, but not 3c2 alone binds eIF1 in a manner compatible with eIF1 binding to the ribosome. Thus, the role of 3c2 in stimulating eIF1 binding to eIF3c-NTD, if any, appears to be indirect.

NMR Evidence that Segment 3c0-Box6_{SSU+} Interacts with the Ribosome-Binding Surface of eIF1

Relative to 3c1-3c2 segment D₅₈₋₁₆₃, fragment C₃₆₋₈₇, containing 3c1 and part of 3c0, displayed CSPs of greater intensity for a larger number of ¹⁵N-eIF1 resonance peaks (Figure 3D). Herein, in addition to R53 and L96 eIF1 residues, extensive CSPs were also observed for D61, A63, and N65, which are localized in the α 1- β 3 loop, T41 in β 2 near loop 1, and L80 (Figure 3D, summarized in Figure 3A). This suggests that C₃₆₋₈₇ fragment binds an entire side of β sheets 1–4 of eIF1 that is adjacent to K60 at the α 1 C terminus, the 40S contact site, and is likely to overlap with the second 40S contact site in loop 1, R36 (cyan lettering in Figure 4B). Interestingly, the resonance corresponding to I93 was slightly shifted in the presence of C₃₆₋₈₇ without attenuation of its signal (Figure 3D, yellow for weak/moderate interaction) but did not disappear (line broadening) as observed for D₅₈₋₁₆₃ (red in Figure 3B). As summarized in Figure 4B, this pattern suggests that C₃₆₋₈₇ still retains interaction with R53 and L96 of eIF1 through the core element, 3c1, while its interaction with eIF1-I93 is diminished due to lack of 3c2. This supports an indirect stimulatory role for 3c2 in eIF1 binding to eIF3c-NTD (dotted line in Figure 4A).

Importantly, these data also suggest that the presence of the C-terminal half of 3c0 in C₃₆₋₈₇ confers more extensive interactions with the ribosome-binding surfaces of eIF1 (Figure 4B).

eIF1 residues assigned to the resonances are shown with their aa numbers. aa of high relevance (R53, K56, K60, I93, L96) are highlighted in red. Bottom, the eIF1 residues affected by each eIF3c segment are painted orange or yellow for strong or moderate CSP of >0.1 ppm or 0.05–0.1 ppm, respectively, in the ribbon diagram of yeast eIF1 structure. The eIF1 residues whose resonances caused line broadening were painted red. Locations of aa of high relevance are indicated. Prolines (11, 46, and 72) and unassigned residues (23, 34–36, 66, and 107) are painted green and gray, respectively. In (B), note that, upon eIF3c-D₅₈₋₁₆₃ addition, the cross peak for K60 was shifted only slightly (green arrowhead in the spectrum), overlapping with that for K56, which shifted a greater distance (long black arrow).

See also Tables S1 and S4.

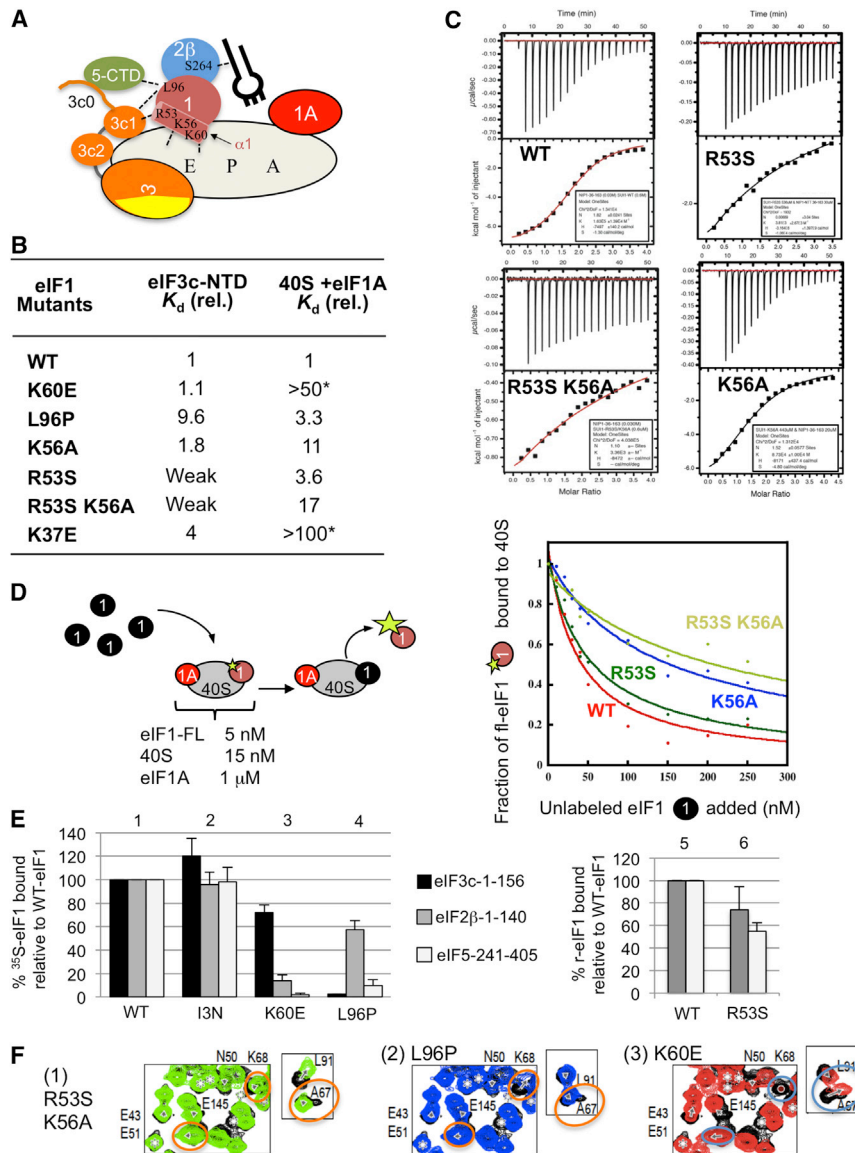


Figure 5. Effect of eIF1 Mutations on Interaction with eIF3c-NTD, eIF5-CTD, and the 40S Subunit

(A) Interactions described here to stabilize the open PIC are presented by dotted lines. Circles indicate eIFs, its subunits or domain. eIF1 or eIF2β aa involved in the interactions are presented within the circles. The cylinder attached to eIF1 is its α1. Plug, Met-tRNA_i. The largest oval, 40S subunit with tRNA-binding sites (A, P, and E).

(B) Affinity of eIF1 or its mutants for 40S or eIF3c-NTD. Shown are relative K_d values compared to the value obtained with WT eIF1. Original values are shown in Figure S6. Asterisk, values based on a previous study (Martin-Marcos et al., 2013).

(C) ITC thermograms for eIF3c-B binding to eIF1 and its mutants indicated.

(D) 40S binding assay by eIF1 mutants. (Left) Schematics illustrating experimental schemes. Binding of fluorescently labeled eIF1 (brown circle) to 40S (oval):eIF1A (red circle) complex was monitored by FA in the presence of given concentrations of indicated unlabeled eIF1 forms (filled circle). The size of the star indicates the degree of FA. (Right) Fraction of 40S bound to the labeled eIF1 was plotted against the concentration of unlabeled eIF1 species.

(E) GST-pull-down assay. The values for the binding of eIF1 mutants to GST-fusion proteins indicated on top are presented relative to those obtained with WT eIF1 with bars indicating SEM ($n = 2$ or more). eIF1-I3N was used as a negative control.

(F) Close-up views of ^1H - ^{15}N HSQC spectra of ^{15}N -eIF3c-B (aa 36–163) in the absence (black) or presence of indicated mutant eIF1 protein species (panels 1–3) (1:1.2 in color). See also Figures S6 and S7 and Table S1.

This is in agreement with its N value in ITC experiments of 0.7, indicating more than one binding site on eIF1 (Figures 1A and S2B). Hence, we propose that 3c0 does not engage eIF1 in the scanning PIC because its binding site on eIF1 overlaps with the 40S-binding surface.

The conclusion that the C-terminal half of 3c0 containing Box6_{SSU+} engages ribosome-binding surface of eIF1 is further supported by ITC analysis indicating that the K37E substitution in eIF1 loop 1 reduces eIF1 binding to eIF3c-NTD by 4-fold (Figures 5B and S6A). Importantly, 3c0:eIF1 interaction may explain the Ssu⁻ phenotype of the Box6R mutation (Valásek et al., 2004). Notwithstanding that due to the relatively low affinity of eIF1 for eIF3c-NTD ($K_d = 1 \mu\text{M}$) 3c0 is unlikely to displace eIF1 from PIC (eIF1:40S subunit $K_d = 1$ –10 nM) (Martin-Marcos et al., 2013), by competing with the eIF1:40S subunit interaction 3c0 may increase the chance that eIF1 is inappropriately released from the 40S sub-

unit at a non-AUG codon. By disrupting this competition, the Box6R mutation of 3c0 is expected to stabilize the scanning PIC and diminish non-AUG initiation (Ssu⁻ phenotype). Thus, combined with the genetic findings (Valásek et al., 2004), the CSP study in Figure 3D suggests that 3c0:eIF1 interaction impedes eIF1 binding to the ribosome.

The largest fragment examined, eIF3c-B_{36–163}, containing the C-terminal half of 3c0, and full 3c1 and 3c2, induced a combination of CSPs observed for both eIF3c-D_{58–163} (3c1+3c2) and eIF3c-C_{36–87} (3c0+3c1) (Figure 3E). These included major perturbations in the following eIF1 residues: R53 and L96 (due to 3c1), I93 (due to 3c2) and residues in the proximity of both eIF1 ribosome-binding surfaces (attributed to 3c0). Altogether, these results suggest that 3c0 and 3c2, flanking the core eIF1 binding element 3c1, may differentially modulate interaction of eIF3c-NTD with eIF1.

Arg-53 and Leu-96 of eIF1 Make Critical Connections to the eIF3c-NTD within the Scanning PIC

CSP analysis implicated eIF1 residues R53 and L96, in the N-terminal end of α1 and nearby hydrophobic patch, in interaction

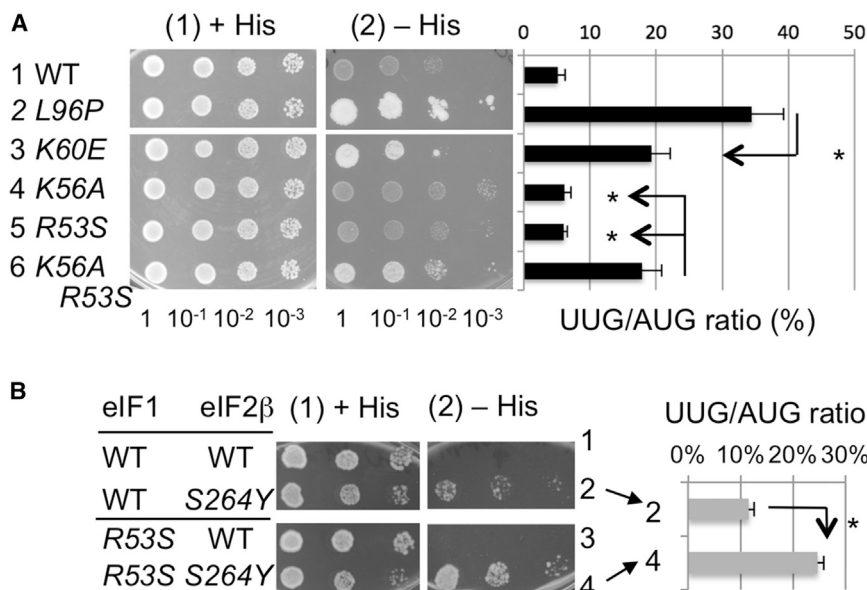


Figure 6. Effect of eIF1 Mutations on Stringent Translation Initiation In Vivo

(A and B) Sui^- phenotype tests. Indicated yeast eIF1 mutants are assayed for *his4-UUG* expression (His⁺ test, panel 2 with – His plate; panel 1 with + His plate shown as a loading control) or UUG/AUG initiation ratio (graph to the left); bars indicate SEM. * $p < 0.05$ (A, $n = 5$; B, $n = 4$). See Supplemental Information for details. See also Tables S1 and S2.

with all three eIF3c-NTD constructs that bind eIF1 with strong affinity (Figures 3B, 3D, and 3E). Accordingly, we tested the effect of substituting these residues on eIF1 binding to eIF3c-B_{36–163} in vitro. As controls, we examined eIF1 substitutions K56A and K60E, which are involved in 40S binding (see Figures 3A and 5A for eIF1 residues altered). In the ITC assay, R53S substitution reduced eIF3c binding below the detection limit, whereas L96P substitution reduced the affinity by 10-fold (Figures 5B and 5C). In contrast K56A and K60E exerted little effect on eIF3c-NTD:eIF1 binding (Figures 5B and S6A), which is consistent with NMR data. These results were verified by CSP experiments (Figures 5F and S4). These results also agree with our previous GST pull-down assays indicating that simultaneous substitution of eIF1 residues K52, R53, K56, K59, and K60 distributed along $\alpha 1$ (*sui1-M5*) (Reibarkh et al., 2008) and I93, L96, and G97 in the hydrophobic patch (*sui1-93-97*) (Cheung et al., 2007) reduces eIF1 binding to eIF3c (italicized are aa whose single substitution was found here to reduce the interaction).

The eIF1 substitutions L96P and K60E are known to allow misinitiation from UUG codons in vivo (Sui^- phenotype) (Martin-Marcos et al., 2013), which we verified using a *UUG-his4* allele and *UUG-lacZ* reporter (Figure 6A, rows 1–3). The K60E substitution strongly impairs 40S binding in vitro (Martin-Marcos et al., 2013). Since the L96P substitution reduces eIF1 interaction with eIF3c-NTD (Figure 5B), its strong Sui^- phenotype (Figure 6A) could be attributed to defective interaction with eIF3c. However, neither eIF1 R53S nor K56A elevate UUG initiation (Figure 6A, rows 4 and 5), even though R53S had a greater effect than L96P on eIF3c-NTD binding (Figure 5B). Thus, the dramatic reduction in initiation stringency conferred by L96P likely results from disrupting eIF1 interactions with other components of the scanning PIC besides eIF3c-NTD.

To test this tenet, we examined the effects of L96P on eIF1 interactions with its other known binding partners: the 40S subunit, the eIF2 β -NTT, and the eIF5-CTD (Figure 5A). We determined the K_d for the 40S-eIF1 complex by measuring changes in fluores-

cence anisotropy (FA) of fluorescently labeled eIF1 in the presence of increasing 40S concentration (Maag et al., 2005). By this approach, we showed that K60E (Martin-Marcos et al., 2013) essentially eliminated, while L96P (this study) modestly reduced eIF1 affinity for 40S, respectively (Figure 5B, column 3). In contrast, L96P strongly reduced eIF1 interaction with the eIF5-CTD in GST

pull-down assays (Figure 5E, column 4). This is consistent with our previous CSP and spin-labeling studies identifying the hydrophobic patch harboring L96 as the eIF5-CTD binding site (Luna et al., 2012; Reibarkh et al., 2008) (Figure 4A). However, L96P only slightly reduced eIF1 binding to the eIF2 β -NTT (Figure 5E; see Figure 5A for summary of interaction involving eIF1-L96). Thus, the strong Sui^- phenotype of L96P (Figure 6A) likely arises from combined defects of reduced eIF1 binding to the eIF3c-NTD, eIF5-CTD (Figure 5E), and perhaps the 40S subunit (Figure 5B).

Despite the fact that eIF1 substitution R53S essentially abolishes binding to the eIF3c-NTD (Figure 5B), it has no effect on initiation accuracy (Figure 6A), implying that eIF1-R53S retains other interactions with the PIC that compensate for impaired interaction with eIF3c. Employing a variation of the FA assay in which excess unlabeled eIF1 competes with wild-type (WT)-labeled eIF1 for ribosome binding (Figure 5D, left), we found that R53S has only a slight effect on 40S binding (Figures 5B and 5D, green curve). Moreover, GST pull-down assays revealed only modest effects of R53S on binding to the eIF2 β -NTT and eIF5-CTD (Figure 5E, right). The CSP assay with ¹⁵N-eIF1-R53S also demonstrates robust eIF5-CTD interaction with this mutant, as observed with WT ¹⁵N-eIF1 (Figure S7) (Reibarkh et al., 2008). Thus, R53S specifically abolishes eIF1 interaction with the eIF3c-NTD (Figure 5B), which is not sufficient to impair accuracy of start site selection in vivo.

To demonstrate a role for eIF1-R53 in stabilizing the scanning PIC in vivo, we generated double mutants. Combining R53S and K56A in eIF1 did not alter the defect in eIF3c-NTD:eIF1 binding seen for R53S alone (Figure 5B) and conferred only a moderate decrease in 40S:eIF1 binding affinity beyond the 11-fold reduction in K_D induced by K56A alone (Figures 5B and 5D, blue and light green curves). Nevertheless, the R53S,K56A double mutant displayed a marked increase in UUG initiation that was not observed for single mutants (Figure 6A, row 6). Since K56A has no effect on eIF1 binding to eIF2 β -NTT and eIF5-CTD when

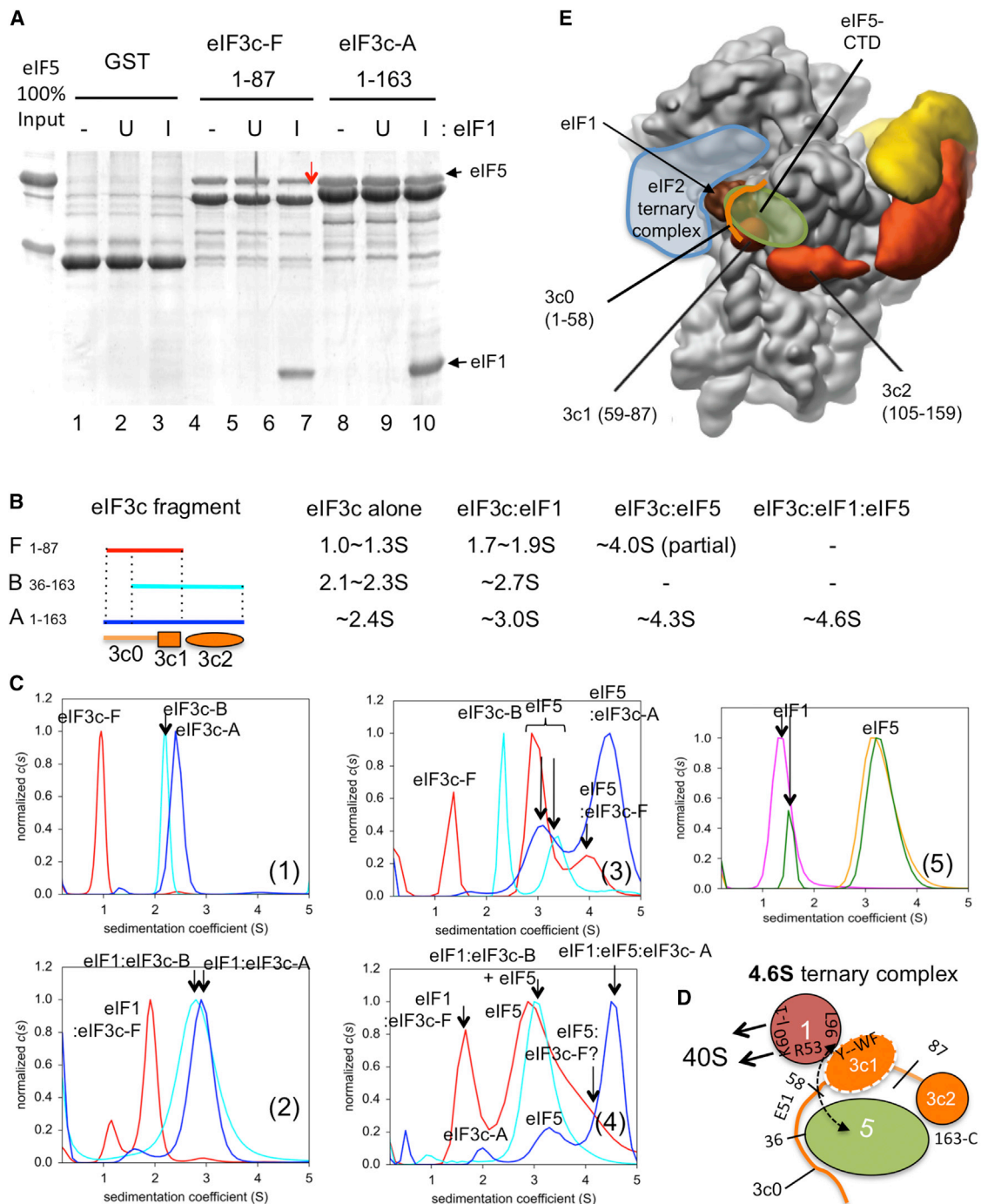


Figure 7. GST Pull-Down and AUC Experiments Characterizing Interaction between eIF3c-NTD, eIF1, and eIF5

(A) GST pull-down assay demonstrating inhibition of eIF5 binding to eIF3c by excess eIF1. ~5 μ g of indicated GST-eIF3c fusion proteins (~0.2 nmol) were allowed to bind ~5 μ g of eIF5 (~0.1 nmol) in the presence of 70 μ g of recombinant eIF1 (~5 nmol) present in induced (I) lysates and the complex analyzed by SDS-PAGE and Coomassie staining. U and -, uninduced lysate or buffer, respectively, was added in place of induced lysates.

(B) Summary of AUC interaction studies. Left, eIF3c-NTD fragments used are shown with bars indicating their relative locations in eIF3c primary structure. Second, third, fourth, and fifth columns list sizes of eIF3c species or complexes formed. -, no complex formation.

(C) AUC analysis. The sedimentation coefficient (c(s)) distributions of reactions containing eIF3c-F (red), B (cyan) and A (blue), either alone (panel 1) or in the presence of eIF1 (panel 2), eIF5 (panel 3) or both (panel 4). Panel 5, eIF1 (pink), eIF5 (orange) and the mixture thereof (green). Proposed peak assignments are presented for each experiment.

(legend continued on next page)

combined with four other eIF1 substitutions in $\alpha 1$ (M5 mutation) (Reibarkh et al., 2008), we conclude that the synthetic *Sui*⁻ phenotype of the R53S,K56A substitution (Figure 6A) results from the combined loss of eIF1 interaction with eIF3c-NTD conferred by R53S and weakened 40S binding conferred by K56A (Figure 5B). As shown in Figure 6B (panel 2, row 2 versus 4), the eIF1-R53S substitution also exacerbates the elevated UUG initiation caused by the eIF2 β -S254Y variant (encoded by *SUI3-2*), previously attributed to increased GTP hydrolysis (Huang et al., 1997) and stabilizing the P_{IN} conformation of Met-tRNA_i at UUG codons (Martin-Marcos et al., 2014). Our findings imply that the defective stabilization of the closed/P_{IN} conformation at UUG codons conferred by eIF2 β -S254Y is normally mitigated by the eIF1/eIF3c-NTD interaction (disrupted by eIF1-R53S) to diminish acceptance of codon-anticodon mismatches in the P-site.

In conclusion, these results show that eIF1-R53 and -L96 are key eIF3c-NTD interaction sites in vivo. Within the scanning PIC, eIF3c-NTD appears to be the sole binding partner of eIF1-R53, whereas eIF1-L96 appears to engage both eIF3c-NTD and eIF5-CTD. Thus, multiple interactions between eIF3c, eIF5, and the ribosome collaborate in retaining eIF1 within the scanning PIC (Figure 5A; also see Supplemental Results).

eIF5 Regulates eIF3c-NTD Interactions with eIF1

We speculated that the 3c0:eIF1 interaction competes with 40S:eIF1 interaction to favor eIF1 dissociation from the PIC at the start codon. We therefore addressed whether the known interaction of eIF5-CTD with 3c0 (Karásková et al., 2012) can preclude this destabilizing effect of 3c0 on the scanning PIC. As shown in Figure S8, GST pull-down assays demonstrated that GST-eIF3c-A₁₋₁₆₃, -F₁₋₈₇, and -G₁₋₅₈, but not GST-eIF3c-E₈₇₋₁₆₃ associates with eIF5, supporting the idea that the minimal eIF5-binding segment in eIF3c-NTD spans across 3c0 residues 1–46 (Karásková et al., 2012). To examine competition between eIF1 and eIF5 for 3c0 binding, we used GST-eIF3c-F₁₋₈₇ and eIF3c-A₁₋₁₆₃, which both exhibit high affinity for eIF1 (Figure 1A), and monitored their binding to the full-length eIF5 (in 1:1 stoichiometry) in the presence of >10-fold molar excess of eIF1. As shown in Figure 7A, eIF1 inhibited eIF5 binding by fragment F₁₋₈₇ to 51% \pm 0.8% ($p < 0.0001$, $n = 4$) (lanes 6 versus 7, red arrow), supporting the idea that eIF1 competes with eIF5 for 3c0 binding. Importantly, this inhibition was not observed with A₁₋₁₆₃ fragment (lanes 9 versus 10), indicating that the presence of 3c2 in fragment A₁₋₁₆₃ allows eIF1 and eIF5 to avoid competition for binding to eIF3c-NTD.

To corroborate these findings, we used analytical ultracentrifugation (AUC) to examine the size and hence, stoichiometry,

of complexes formed by eIF1, eIF5, and different eIF3c-NTD fragments. When tested alone, eIF3c fragments A₁₋₁₆₃, F₁₋₈₇, and B₃₆₋₁₆₃ display single peaks (Figure 7C, panel 1), ranging in size from 1.7S to 3.0S (Figure 7B). Addition of eIF1 allowed formation of a dimeric complex with each eIF3c fragment (Figures 7C, panel 2, and 7B), as expected from their high affinity for eIF1 (Figure 1A). In assays where eIF5 fragments were included (Figure 7C, panel 3), F₁₋₈₇ bound eIF5 partially, whereas B₃₆₋₁₆₃, lacking the eIF5 binding site in 3c0, did not bind eIF5 at all (red and cyan lines). Interestingly, eIF5 assembled into a 4.3S complex with A₁₋₁₆₃, leaving no unbound fragment A₁₋₁₆₃ (blue line). This strong interaction with eIF5 requires 3c2 devoid of F₁₋₈₇ (red line).

As shown in Figure 7C, panel 4, the AUC assay confirmed formation of a stable 4.6S trimeric complex comprising eIF1, eIF5, and eIF3c-A₁₋₁₆₃ (blue line) (Asano et al., 2000; Singh et al., 2004). In contrast, eIF1, eIF5, and eIF3c-F₁₋₈₇ did not assemble into a trimeric complex (Figure 7C, panel 4, red line), even though eIF1 and F₁₋₈₇ formed a 1.7S complex. These results support competition by eIF1 and eIF5 for 3c0 binding, which can be relieved by 3c2 present in eIF3c-A₁₋₁₆₃ but not eIF3c-F₁₋₈₇.

When bound to eIF3c-B₃₆₋₁₆₃ defective in eIF5-binding, eIF1 was unable to bind eIF5, and free eIF5 and the B₃₆₋₁₆₃:eIF1 complex were found co-sedimenting at ~3S (Figure 7C, panel 4, cyan line). Thus, forming the trimeric complex requires eIF5 interaction with the N-terminal region of 3c0 (aa 1–35). Because eIF1 and eIF5 did not form a complex in the absence of eIF3c fragments (Figure 7C, panel 5), we conclude that the entire eIF3c-NTD (aa 1–163) bridges these two proteins, with eIF1 bound to its C-terminal portion, as found in eIF3c-D₅₈₋₁₆₃ (Figure 1A; Karásková et al., 2012). The proposed interactions in the eIF5:eIF3c-NTD:eIF1 trimeric complex are depicted in Figure 7D. Here, it should be noted that the eIF5-CTD:3c0 interaction precludes the 3c0:eIF1 interaction that otherwise competes with eIF1:40S association, and we propose that this stabilizes the scanning PIC. Based on these findings, we suggest approximate locations of the eIF5-CTD and eIF3c0 in the PIC (Figure 7E) compatible with the proposed roles of these segments in regulating the transition from scanning to start codon recognition.

DISCUSSION

The results of NMR and complementary quantitative binding assays presented in this work revealed two distinct eIF1 complexes formed with overlapping eIF3c-NTD segments that appear to function at different stages of the initiation pathway. The C-terminal segment of the eIF3c-NTD (fragment D₅₉₋₁₆₃)

(D) Schematic illustration of the proposed 4.6S trimeric complex. eIF3c-NTD is drawn as blue orange line representing unstructured segments, 3c0 (aa 1–58), and orange circles representing 3c1 (aa 59–87) and 3c2 (aa 105–159), as found in cryo-EM models in Figure 2 and redefined based on ¹⁵N-eIF3c-B CSP studies (Figure 3). eIF5 (dark green circle) is depicted as contacting both ends of eIF3c-NTD. eIF1 (brown circle) is bound to 3c1 via R53 and L96 (labeled). K60 and loop 1 (I-1) of eIF1 are open for 40S binding (arrows). E51, showing CSP with eIF1; Y-WF; Y70, W74, F75, showing line broadening with eIF1 (Figures 1E and S3). Numbers along eIF3c schematics indicate boundaries of eIF3c units. Dotted arrow indicates the interaction between eIF1-L96 and eIF5, suggested here to stabilize the trimeric complex, as it does in the scanning PIC (Luna et al., 2012; Reibarkh et al., 2008).

(E) Locations of eIF2 ternary complex (blue drawing) (Llácer et al., 2015), eIF5-CTD (dark green circle, this study) and 3c0 (aa 1–58) (orange line, this study) are superimposed onto the re-calculated cryo-EM structure, as shown in Figure 2B, right.

See also Figure S8 and Table S1.

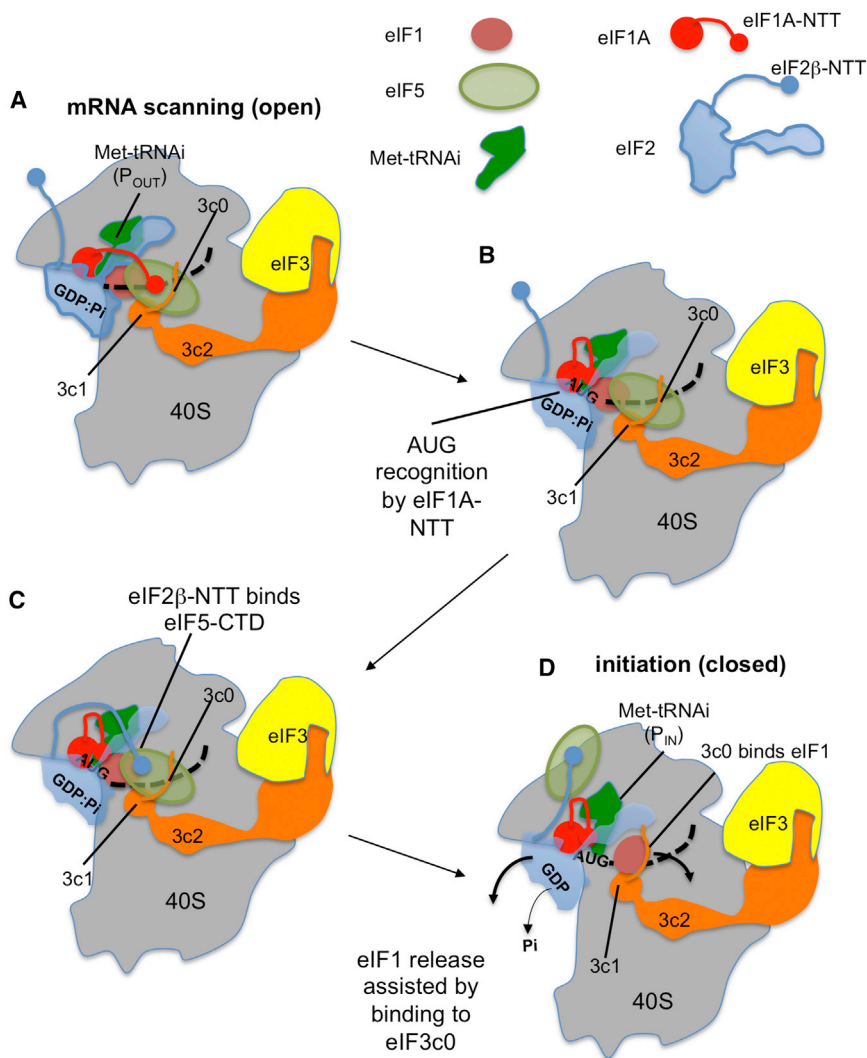


Figure 8. Model of MFC Rearrangement during Translation Initiation

(A) During mRNA (dotted line) scanning, eIF5 and eIF3c-NTD play crucial roles in eIF1 anchoring. eIF5 and 3c1 directly bind eIF1 to anchor it to the PIC (this study). eIF1 maintains the PIC in the open conformation and prevents Met-tRNA_i to accommodate in the P-site (P_{OUT}). eIF1A-NTT also binds eIF5 (Luna et al., 2013), preventing its binding by eIF2β-NTT. The binding partner of eIF2β-NTT at this stage may be eIF1 (Nanda et al., 2013) or alternatively, rRNA or mRNA, as it binds RNA (Singh et al., 2012).

(B) Met-tRNA_i base-pairing to the AUG codon causes a scanning arrest. This is enhanced by eIF1A-NTT binding to the codon:anticodon duplex, resulting in eIF1 distortion (Hussain et al., 2014; Lácer et al., 2015). eIF5 is now available for eIF2β-NTT binding.

(C) eIF2β-NTT binds eIF5, resulting in disruption of eIF5 binding to eIF1 (Luna et al., 2012) and 3c0. (D) 3c0 assists eIF1 release by binding to its ribosome binding site (this study). tRNA^{Met} bound to the start codon positions in the P-site (P_{IN}). eIF1 release is followed by Pi release from eIF2 (Algire et al., 2005), promoting ejection of eIF2:GDP in complex with eIF5 (Singh et al., 2006). The model that eIF1 remains associated with eIF3 after its release from the 40S decoding site was previously proposed (Karásková et al., 2012; Singh et al., 2012).

segment 3c0 (Figure 7D). Dissolving the eIF5-CTD:3c0 interaction thus emerges as a key step in the transition from the open to closed conformation of the PIC, and we propose a plausible mechanism for this rearrangement below.

In agreement with our proposal that the 3c1/3c2 segments of eIF3c-NTD cooperate to anchor eIF1 on the scanning PIC, eIF1 substitution L96P, which perturbs the interface with 3c1, reduces eIF1 binding to the eIF3c-NTD. By also impairing eIF1 binding to the eIF5-CTD, L96P dramatically elevates UUG initiation in the manner expected for destabilization of the scanning complex (Martin-Marcos et al., 2013). eIF1 substitution R53S, which affects the neighboring surface in helix α 1, dramatically reduces eIF3c-NTD binding but does not substantially impair the eIF1:eIF5-CTD interaction. Because R53S elevates UUG initiation only when combined with the α 1 substitution K56A, which weakens eIF1:40S interaction, we conclude that a network of eIF1 interactions with the eIF3c-NTD, eIF5-CTD, and 40S subunit cooperate to anchor eIF1 to the scanning PIC and block initiation at non-AUG codons (Figure 5A). Based on the cryo-EM model in Figure 2A, the role of 3c2 in anchoring eIF1 to the PIC appears to be indirect. Consistently, mutations altering Box12_{Sui+} (aa 111–120) within 3c2 can elevate UUG initiation by either increasing or decreasing eIF1 retention in native PICs (Karásková et al., 2012). This complexity may reflect dual role of 3c2 in promoting eIF1 binding to segment 3c1 and eIF5-CTD binding to 3c0 in the scanning PIC, while preventing

contains the core eIF1-binding unit 3c1 (aa 59–87) and the adjacent globular domain 3c2 (aa 105–159), which bind to a limited surface on eIF1, including R53 and L96, in a manner compatible with eIF1 binding to the 40S subunit (Figures 4A and 4C). We have assigned two densities projecting from the main body of eIF3 in the eIF1:eIF3:40S cryo-EM structure (Erzberger et al., 2014) as 3c1, which contacts eIF1, and 3c2 interacting with uS15 (Figure 2). In contrast, eIF3c fragment C_{36–87}, containing 3c1 and the C-terminal half of 3c0, interacts with a broader surface of eIF1 that includes R53 and L96 but additionally contains residues surrounding the two 40S binding sites at the C terminus of α 1 and loop 1 (Figures 4C and 4B). Based on the Ssu[−] phenotype of a mutation in Box6 (aa 51–60) within 3c0, we propose that interaction of eIF1 with 3c0 occludes the 40S-binding surface in eIF1 and thus facilitates eIF1 dissociation at the start codon—the event diminished at UUG codons by the Box6R Ssu[−] mutation. This destabilizing effect is likely to be opposed in the scanning PIC through eIF5-CTD binding to 3c0, which shifts eIF1 interaction from eIF3c-NTD elements 3c0/3c1 to 3c1/3c2 and thereby eliminates occlusion of the 40S binding surface on eIF1 by

the more stable eIF1 complex formed with 3c0/3c1 on AUG recognition. In addition, by directly contacting 40S protein uS15/S13, 3c2 is likely to stabilize eIF1 binding to the scanning PIC (Figure 2A).

Recent studies reveal structural rearrangements within 43S/48S PICs between different steps of initiation (Hussain et al., 2014; Llácer et al., 2015; Simonetti et al., 2016). However, it is unclear exactly how start codon selection induces transition from the open to closed conformations of the PIC. Based on our findings and other work done using yeast *S. cerevisiae* as a model system, we propose that the eIF1A-NTT plays such a signaling role (Saini et al., 2010) (Figure 8). During mRNA scanning, eIF1A-NTT interaction with the eIF5-CTD helps to retain eIF1 in the PIC (Luna et al., 2013) (Figure 8A). Thus, in addition to binding the 3c0 element and eIF1, the eIF5-CTD binds the basic eIF1A-NTT through a distinct acidic surface. This interaction is also important as it antagonizes eIF5-CTD interaction with the positively charged eIF2 β -NTT, which would otherwise promote eIF1 release (Luna et al., 2012; Nanda et al., 2013). On Met-tRNA^{Met} anticodon pairing to AUG, eIF1A-NTT binds to the codon:anticodon duplex in the P-site (Hussain et al., 2014) (Figure 8B). This releases the eIF5-CTD for interaction with eIF2 β -NTT, which, in turn, disrupts eIF5-CTD interaction with both eIF1 (Luna et al., 2012) and 3c0 (Figures 8B and 8C). The 3c0 segment is now free to engage eIF1 and occlude its ribosome-binding surface, interfering with eIF1 re-association with the 40S subunit and thus allowing Met-tRNA^{Met} to remain stably anchored in the P_{IN} state (Figure 8D). These effects are expected to amplify the subtle distortion of eIF1 structure and perturbation of its 40S binding site that accompanies Met-tRNA^{Met} isomerization to the P_{IN} state (Hussain et al., 2014). In this way, 3c0 ensures irreversible eIF1 release from the decoding center in response to AUG recognition and subsequent closure of the ribosome structure and formation of the 40S initiation complex.

It is noteworthy that human eIF1 also binds eIF3c-NTD (Fletcher et al., 1999) and eIF5-CTD (Luna et al., 2012). While the eIF3c-NTD segments corresponding to 3c0 are shorter in animals and plants, they contain an acidic element similar to Box6, lying next to the conserved core region 3c1 (Boxed in Figure S1B). Moreover, eIF3c-NTD in animals and plants is predicted to form an α -helical structure, as found in yeast 3c2 (Figure 1C). Further work on the human and yeast systems is expected to reveal Eukarya-wide conservation of the MFC's role in promoting scanning and AUG selection through the coordinated interactions of the eIF3c-NTD with eIF1, eIF5 and potentially other parts of eIF3 (Hussain et al., 2014; Simonetti et al., 2016; Valásek et al., 2003).

EXPERIMENTAL PROCEDURES

Protein Purification and Yeast Methods

Isotopically labeled or unlabeled proteins were expressed in *E. coli* transformants carrying appropriate plasmids (Table S1) and purified as described in Supplemental Information. Yeast *Saccharomyces cerevisiae* strains used in this study are constructed as described in Supplemental Information and listed in Table S2. Standard yeast molecular biology methods including growth and β -galactosidase assays were used throughout (Lee et al., 2007) (see Supplemental Experimental Procedures for details).

Biophysical Methods

ITC, NMR spectroscopy, FA, and AUC are all performed as described in Supplemental Experimental Procedures. Detailed NMR data and structural statistics for eIF3c-B_{36–163} and eIF1 are summarized in Tables S3 and S4, respectively. We re-ran integrative modeling prediction including the new information from the eIF3c-B NMR structure, with parameters and methods identical to those previously described (Erzberger et al., 2014).

ACCESSION NUMBERS

The accession numbers for the yeast eIF1 and eIF3c (aa. 36–163) data reported in this paper are PDB: 2rvh and 5H7U, respectively.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, Supplemental Results, eight figures, and four tables and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2017.02.052>.

AUTHOR CONTRIBUTIONS

E.O., R.E.L., T.N., P.M.-M., H.H., C.R.S., J.P.E., and K.A. designed and performed experiments and analyzed data. F.Z., H.A., and J.M. performed experiments and analyzed data. R.P., S.U., F.S., and T.U. analyzed data. C.M., I.H., E.P., H.Y., M.L.N., B.T., E.A., S.H., E.D., A.N., and P.G. performed experiments. R.E.L., A.G.H., G.W., and K.A. wrote the paper.

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Supplemental Information

Molecular Landscape of the Ribosome

Pre-initiation Complex during mRNA Scanning:

Structural Role for eIF3c and Its Control by eIF5

Eiji Obayashi, Rafael E. Luna, Takashi Nagata, Pilar Martin-Marcos, Hiroyuki Hiraishi, Chingakham Ranjit Singh, Jan Peter Erzberger, Fan Zhang, Haribabu Arthanari, Jacob Morris, Riccardo Pellarin, Chelsea Moore, Ian Harmon, Evangelos Papadopoulos, Hisashi Yoshida, Mahmoud L. Nasr, Satoru Unzai, Brytteny Thompson, Eric Aube, Samantha Hustak, Florian Stengel, Eddie Dagraca, Asokan Ananbandam, Philip Gao, Takeshi Urano, Alan G. Hinnebusch, Gerhard Wagner, and Katsura Asano

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Supplemental Results

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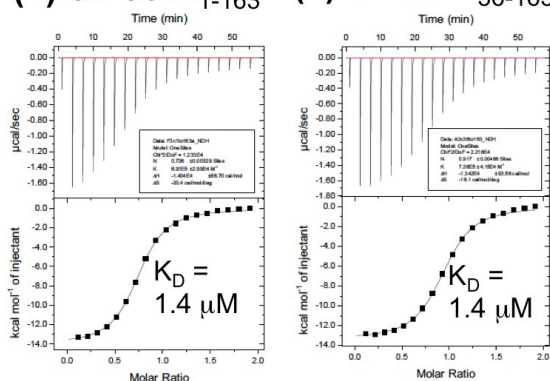
Supplemental References

1

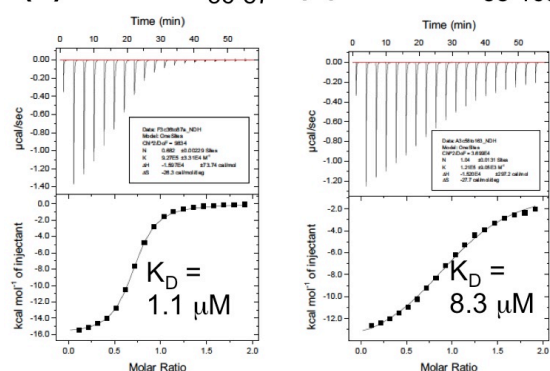
the eIF3c segment fused to GST, based on all the GST pulldown experiments presented in this study. Thus, eIF3c-F and -G, which did not bind eIF1 in panel B, is shown to bind minimal amounts of eIF1 based on other GST pulldown experiments (Fig. S1C and 1B). In (B), 0.2 nmol GST fusion proteins (0.2 μ M) were mixed in 1 ml binding buffer with 0.4 nmol eIF1 (0.4 μ M). A fraction of the complex was isolated by glutathione sepharose chromatography and analyzed by SDS-PAGE followed by Coomassie staining. In (C), 0.12~0.17 nmol GST fusion proteins (0.6~0.9 μ M) attached with the glutathione resin were mixed in 0.2 ml binding buffer with ~6 nmol eIF1 (~30 μ M). After washing the resin four times, the entire pellet fractions were analyzed by SDS-PAGE and Coomassie staining.

A eIF1 titration against eIF3c

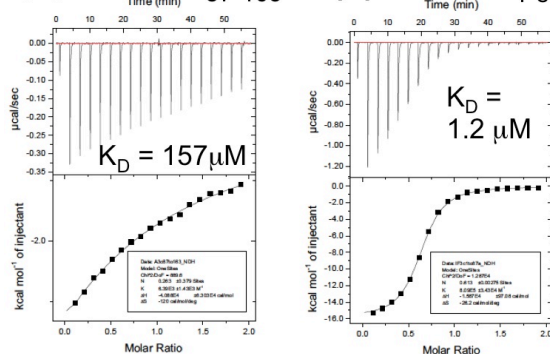
(1) eIF3c-A₁₋₁₆₃ (2) eIF3c-B₃₆₋₁₆₃



(3) eIF3c-C₃₆₋₈₇ (4) eIF3c-D₅₈₋₁₆₃



(5) eIF3c-E₈₇₋₁₆₃ (6) eIF3c-F₁₋₈₇



B Thermogram analysis results

Cell	Inject	K _d (μM)	ΔH (kcal/ mol)	ΔS (cal/ mol/ deg)	N
eIF3c-A 1-163	eIF1	1.4	-14.0	-20.4	0.72
		1.3	-13.5	-18.3	0.75
		1.5	-14.0	-20.2	0.73
eIF3c-B 36-163	eIF1	1.4	-13.4	-18.1	0.92
		1.4	-13.4	-18.1	0.95
		1.4	-13.4	-18.0	0.95
eIF3c-F 1-87	eIF1	1.2	-15.8	-25.2	0.61
		1.0	-15.9	-26.0	0.59
		0.86	-15.8	-25.5	0.55
eIF3c-C 36-87	eIF1	1.1	-16.0	-26.3	0.68
		1.2	-16.1	-26.9	0.66
		0.94	-15.8	-25.5	0.66
eIF3c-D 58-163	eIF1	8.3	-15.2	-27.7	1.04
		7.8	-14.8	-26.3	1.08
		7.8	-15.0	-27.1	1.02
eIF3c-E 87-163	eIF1	156.5	-40.9	-120	0.26
		125.9	-19.4	-47.2	0.48
		139.1	-18.0	-42.7	0.56

Fig. S2. Isothermal Titration Calorimetry (ITC) measurements. – Related to Figure 1

(A) The enthalpy changes caused by injections of eIF1 into the different fragments of eIF3c, **1)** A₁₋₁₆₃, **2)** B₃₆₋₁₆₃, **3)** C₃₆₋₈₇, **4)** D₅₈₋₁₆₃, **5)** E₈₇₋₁₆₃, and **6)** GST-F₁₋₈₇. The lower panels show the fitted binding parameters; the solid line in each lower panel represents a calculated curve using the best fit parameters obtained by a nonlinear least-squares fit. (B) Summary of thermogram analysis for each of the ITC experiments (n=3) using eIF1 and the six eIF3c constructs, as shown in Fig. 1A.

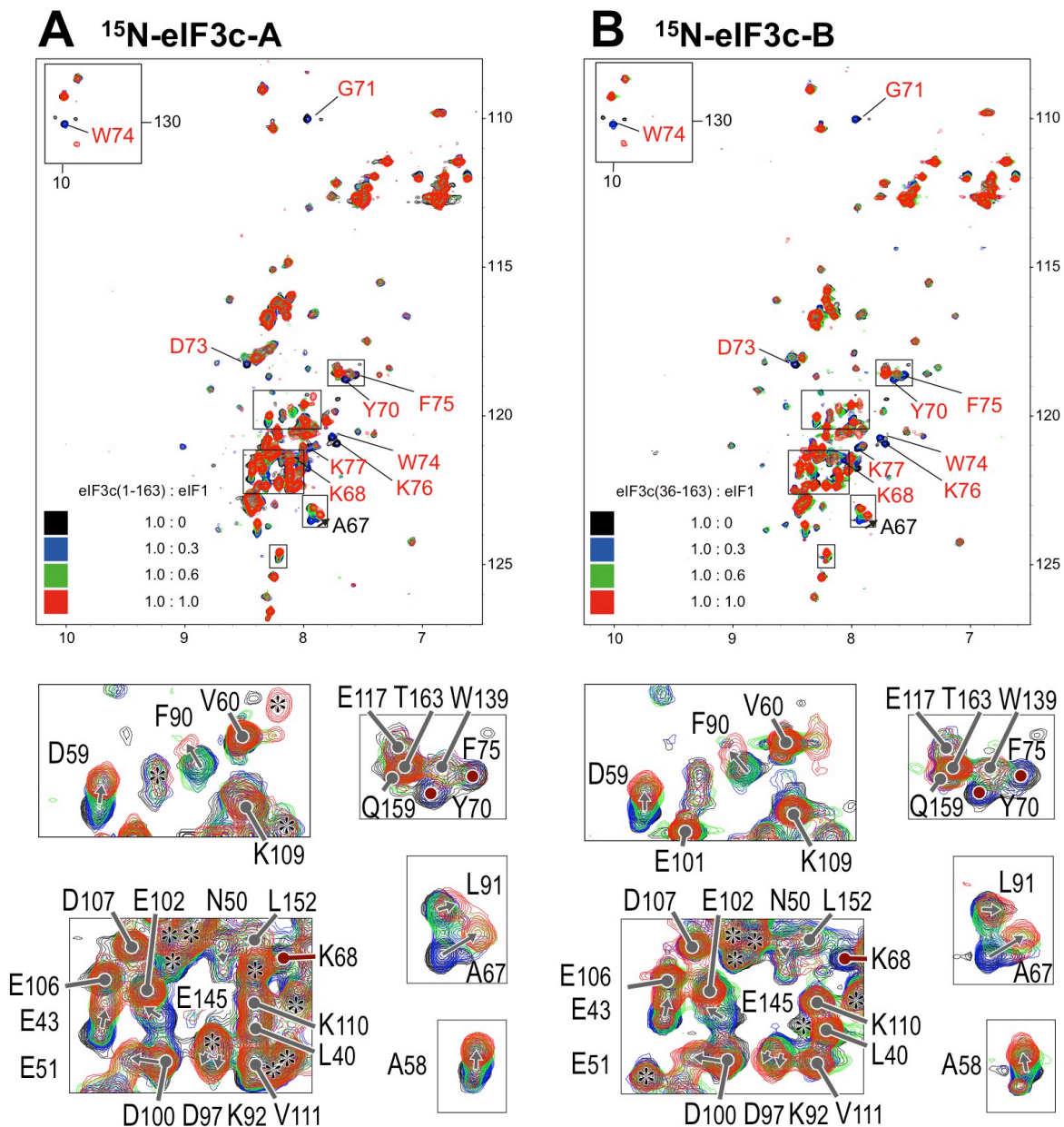


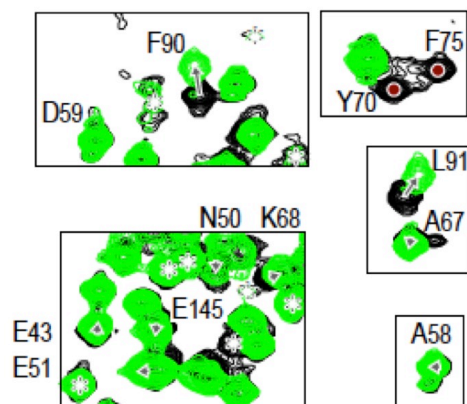
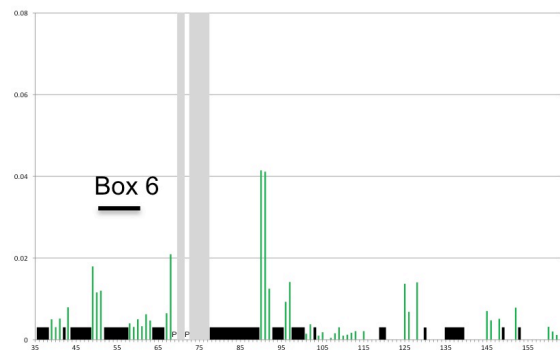
Fig. S3. CSP experiments with ^{15}N -eIF3c-A₁₋₁₆₃ or -B₃₆₋₁₆₃ to determine their amino acids affected by eIF1 binding. – Related to Figure 1

^1H - ^{15}N HSQC spectra of [^{15}N] eIF3c-A (panel A) and [^{15}N] eIF3c-B (panel B) in the presence of eIF1 at indicated molar ratio. Amino acids assigned by determining the structure of eIF3c-B₃₆₋₁₆₃ are indicated in both the panels. For each panel, close-up views display CSP at resonances highlighted by five thin squares in the spectra with CSP shown by arrows. Black dots indicate the locations of assigned amino acids whose resonances are not affected by eIF1 addition. Red brown dots indicate the locations of amino acids whose resonances displayed line-broadening. Asterisk, unassigned resonance.

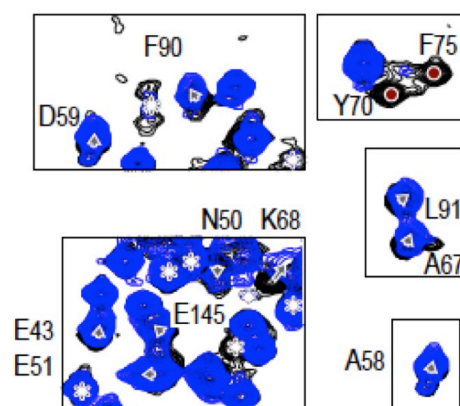
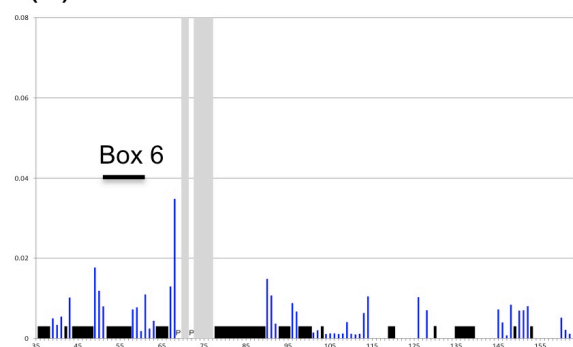
■ Line broadening due to eIF1 protein addition

■ Unassigned amino acids

(1) R53S K56A



(2) L96P



(3) K60E

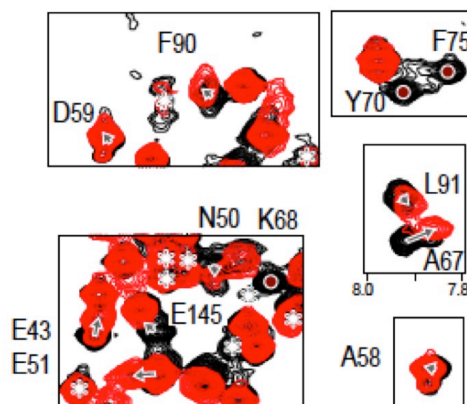
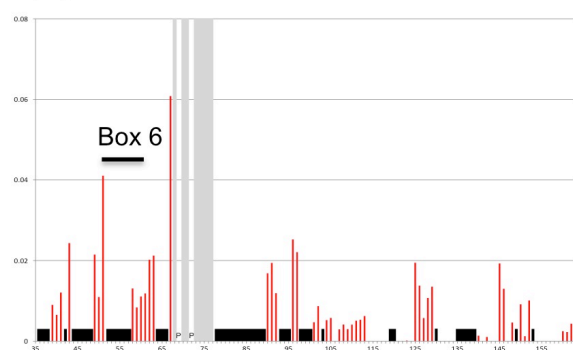


Fig. S4. Chemical shift perturbation observed for eIF3c-B₃₆₋₁₆₃ upon addition of different eIF1 mutants, R53S K56A (panel 1), L96P (panel 2), and K60E (panel 3). – Related to Figure 1

Left, Chemical shift perturbation (CSP), $\Delta\delta$, was computed as described in Supplementary Materials and methods and presented for each assigned amino acid. “P”s indicate proline residues. Black boxes indicate the residues that were not assigned. Shaded regions indicate the residues whose signal in the ^1H - ^{15}N HSQC spectra disappeared upon complex formation (line broadening). Right, ^1H - ^{15}N HSQC spectra of ^{15}N -eIF3c eIF3c-B in the absence (black) or presence of indicated mutant eIF1 protein species (panels 2-4) (1:1.2 in color). For CSP experiment with wild-type eIF1 control, see Fig. 1D and E.

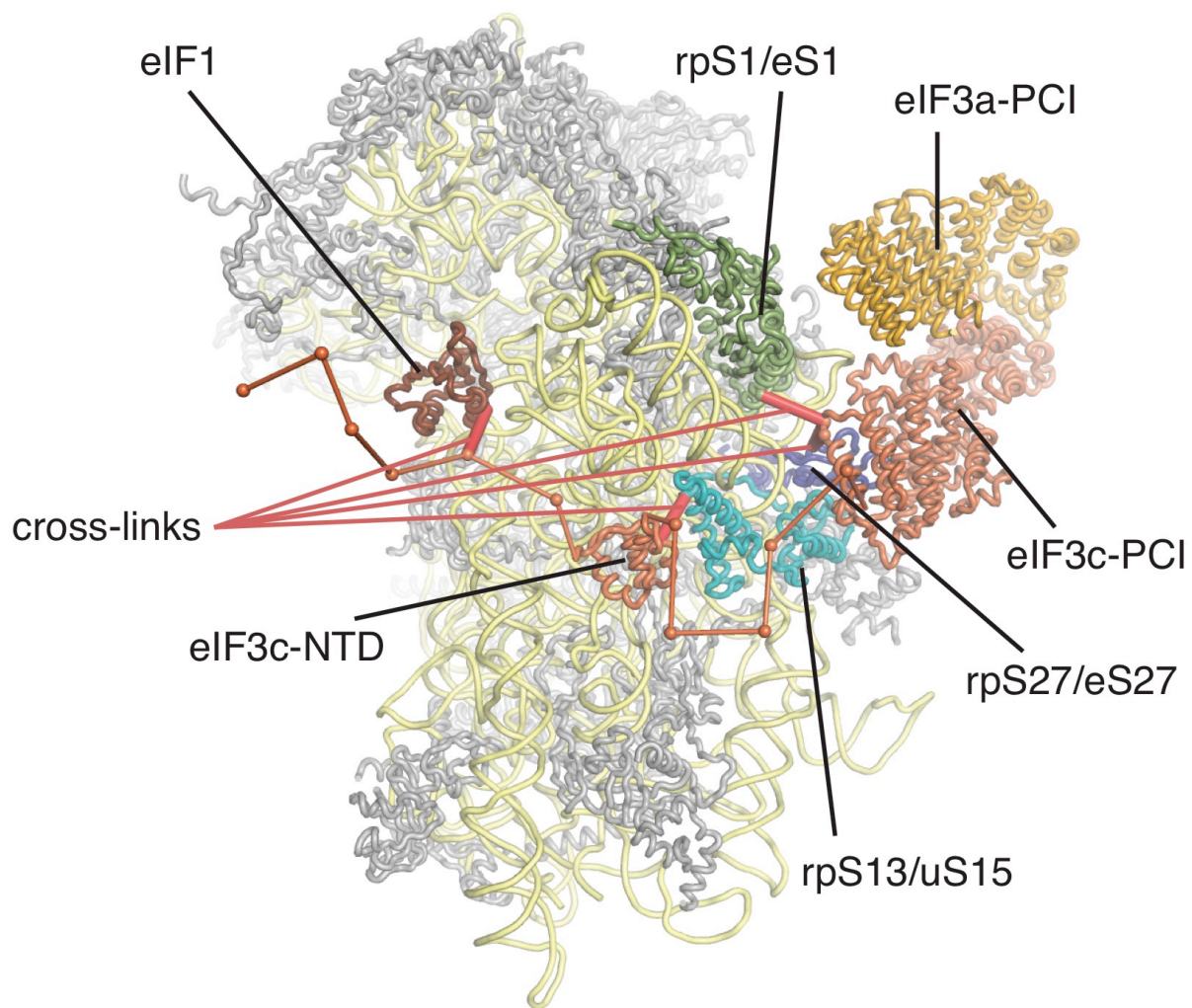


Fig. S5. Representative example of an integrative modeling solution. – Related to Figure 2

Cartoon representation of one of the 500 solutions generated by our modeling runs. eIF3a is colored gold, eIF3c in orange, eIF1 in brown, rpS13/uS15 in cyan, rpS27/eS27 in blue and rpS1/eS1 in green. Interstrand crosslinks are shown in red. Additional structural elements present in the modeling have been omitted for clarity.

A

Sample in Well	Titant in Syringe	Binding Affinities
eIF3c-WT (res 1-156)	eIF1-WT (res 1-108; full)	K_d ~ 7 μ M
eIF3c-WT (res 1-156)	eIF1-I3N (mutant)	K_d ~ 5 μ M
eIF3c-WT (res 1-156)	eIF1-R33A (mutant)	K_d ~ 11 μ M
eIF3c-WT (res 1-156)	eIF1-R36E (mutant)	K_d ~ 7.5 μ M
eIF3c-WT (res 1-156)	eIF1-K37E (mutant)	K_d ~ 28 μ M
eIF3c-WT (res 1-156)	eIF1-K60E (mutant)	K_d ~ 8 μ M
eIF3c-WT (res 1-156)	eIF1-L96P (mutant)	K_d ~ 67 μ M
eIF3c-WT (res 36-163)	eIF1-WT (res 1-108; full)	K_d ~6.1 μ M
eIF3c-WT (res 36-163)	eIF1-R53S (mutant)	Weak
eIF3c-WT (res 36-163)	eIF1-K56A (mutant)	K_d ~ 11 μ M
eIF3c-WT (res 36-163)	eIF1-R53S/K56A (mutant)	Weak

B

eIF1 Mutants	40S +eIF1A K_d (nM)	
	Exp I	Exp II
WT	1.6 \pm 0.4	10.5 \pm 2.0
K60E		>500*
L96P		35 \pm 2.0
K56A	17.5 \pm 1.9	
R53S	5.6 \pm 2.5	
R53S K56A	26.0 \pm 4.4	
K37E		> 1000*

Fig. S6. Additional quantitative interaction assays involving eIF1. – Related to Figure 5

(A) Summary of affinity of various eIF1 mutant proteins for eIF3c-N₁₋₁₅₆ (2) or eIF3c-B₃₆₋₁₆₃ (this study), as measured by ITC. Their affinity for WT eIF1 is slightly lower probably due to polyhistidine-tag attached to eIF1. Weak, interaction detected but not quantified due to the curve not fitting to a sigmoid curve. (B) Summary of affinity of eIF1 binding to 40S:eIF1A complex. In Experiment I, wild-type fluorescein-tagged eIF1 was allowed to bind the 40S subunit in the presence of eIF1A and various amounts of indicated eIF1 species, as depicted in Fig. 5B. K_D was computed, based on 3-6 assays. In Experiment II, K_D values were directly measured using the eIF1 variants (WT and mutants) labeled with fluorescein on their C termini. The average of 2 or 3 assays was presented. *, values from a previous study (3).

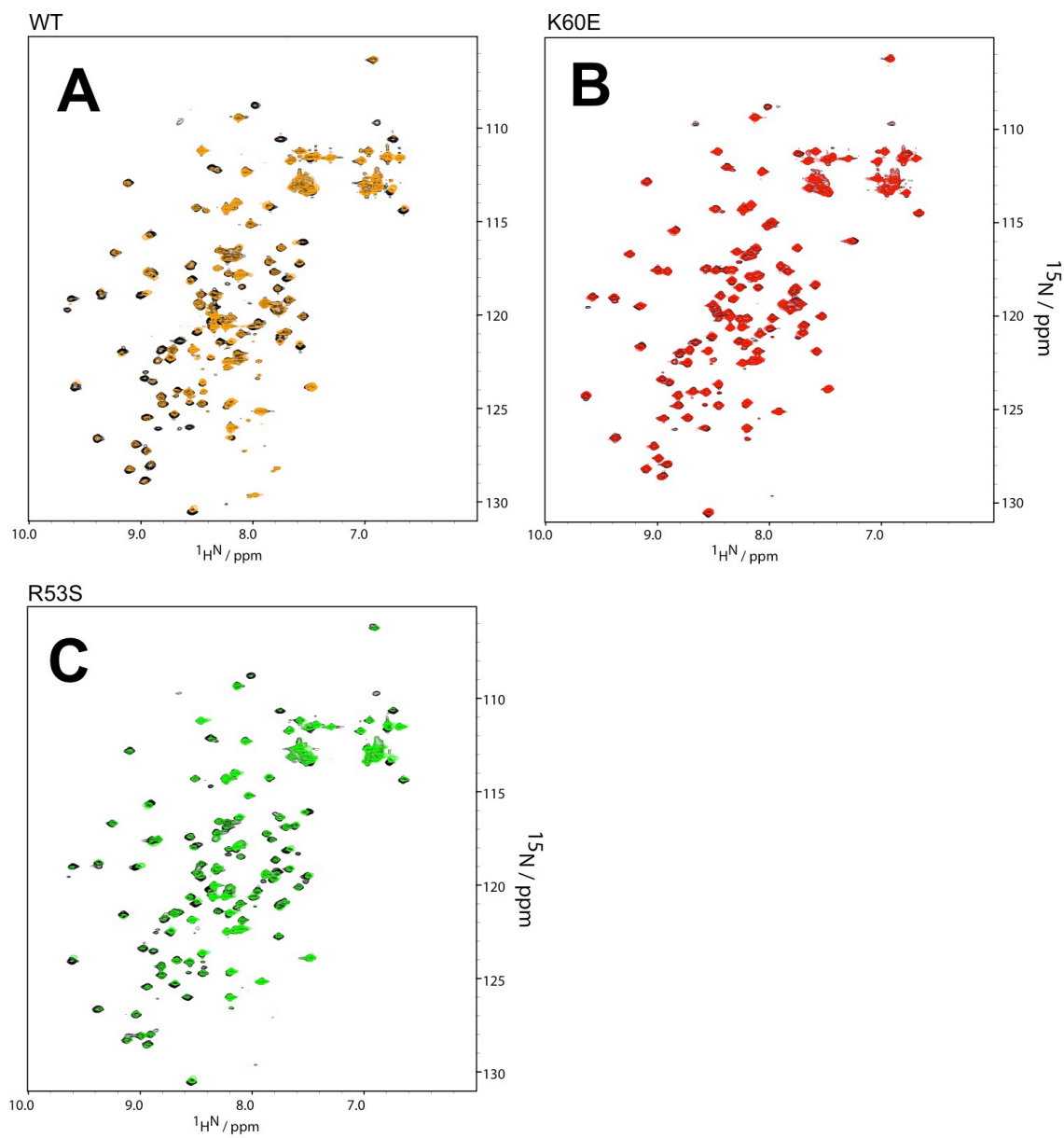


Fig. S7. CSP experiments with WT and mutant ^{15}N -eIF1 and unlabeled eIF5-CTD (aa. 242-405). – Related to Figure 5

(A) HSQC spectra of ^{15}N WT eIF1 free (gray) or ^{15}N eIF1: eIF5-CTD = 1 : 4 (orange). (B) HSQC spectra of ^{15}N eIF1-K60E free (gray) or ^{15}N eIF1-K60E: eIF5-CTD = 1 : 4 (red). (C) HSQC spectra of ^{15}N eIF1-R53S free or ^{15}N eIF1-R53S: eIF5-CTD = 1 : 4 (green).

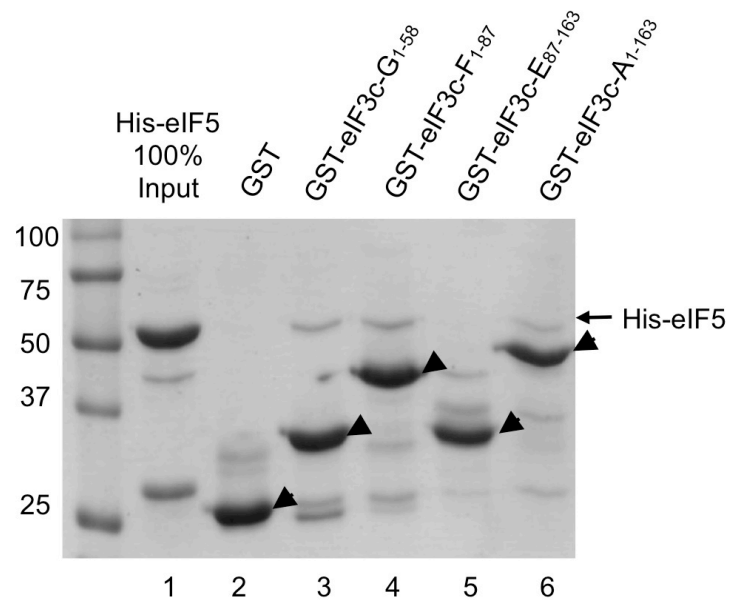


Fig. S8. GST-eIF3c pulldown assays with eIF5. – Related to Figure 8

5 μ g of GST or GST-eIF3c fusion proteins listed across top (0.12~0.17 nmol) are allowed to bind 5 μ g of His-tagged eIF5 (~0.1 nmol; shown as in-put in lane 1) in a 0.2 ml binding reaction and the complexes were isolated and analyzed by Coomassie staining as in Fig. 3B. Horizontal arrow, the location of His-eIF5. Arrowhead, the location of full-length GST fusion proteins.

Table S1. Plasmids employed in this study – Related to Figures 1, 3, 5, 6, and 7

p #	Systematic name	Description	Source	Used in:
188	pGEX-TIF5-B6	To express GST-eIF5 241-405	(4)	Fig. 5E
596	pGEX-SUI3ΔS	To express GST-eIF2b 1-140	(5)	
283	pGEX-NIP1-N	To express GST-eIF3c 1-156	(2)	
225	pT7-SUI1	To express eIF1 under T7 promoter	(6)	
PMB80	pT7- SUI1-I3N	pT7-SUI1 carrying I3N	This study	
PMB81	pT7- SUI1-K60E	pT7-SUI1 carrying K60E	This study	
PMB82	pT7-SUI1-L96P	pT7-SUI1 carrying L96P	This study	
1157	pET28-His-eIF1	To express His-TEV-eIF1 and purify (untagged) eIF1	This study	Fig. 1, 3, 5 and 7
1490	pET28-His-eIF5 242-405	To express His-eIF5 242-405	This study	Fig. S7
1489	pET28-His-eIF3c 1-163	To express His-eIF3c A 1-163	This study	Fig. 1, 3, 7BC, S2, S3 and S4
1491	pET28-His-eIF3c 1-87	To express His-eIF3c F 1-87	This study	
1492	pET28-His-eIF3c 36-163	To express His-eIF3c B 36-163	This study	
1493	pET28-His-eIF3c 58-163	To express His-eIF3c D 58-163	This study	
1494	pET28-His-eIF3c 87-163	To express His-eIF3c E 87-163	This study	
1507	pET28-His-eIF3c 36-87	To express His-eIF3c C 36-87	This study	
1496	pET28-GST-eIF3c 1-163	To express GST-eIF3c A 1-163	This study	Fig. 1B, 7A, S1BC and S8
1497	pET28-GST-eIF3c 1-87	To express GST-eIF3c F 1-87	This study	
1498	pET28-GST-eIF3c 1-58	To express GST-eIF3c G 1-58	This study	
1499	pET28-GST-eIF3c 36-163	To express GST-eIF3c B 36-163	This study	
1500	pET28-GST-eIF3c 58-163	To express GST-eIF3c D 58-163	This study	
1501	pET28-GST-eIF3c 87-163	To express GST-eIF3c E 87-163	This study	
1505	pET28-GST-eIF3c 36-87	To express GST-eIF3c C 36-87	This study	
1502	pET28-SUI1	To express wild-type eIF1	This study	Fig.1B,5E

				and S1C
1504	pET28-His-eIF5	To express His-eIF5	This study	Fig. 7, S8
1590	pET28-His-eIF1 R53S K56A	To express His-eIF1 R53S K56A	This study	Fig. 5, S6A and S7
1591	pET28-His-eIF1 K60E	To express His-eIF1 K60E	This study	
1594	pET28-His-eIF1 R53S	To express His-eIF1 R53S	This study	
1596	pET28-His-eIF1 L96P	To express His-eIF1 L96P	This study	
1598	pET28-His-eIF1 K56A	To express His-eIF1 K56A	This study	
347	pHis-NIP1-N	To express His-eIF3c 1-156	(2)	Fig. S6A
636	pET-SUI1-His	To express eIF1-His	(7)	
5707	pET-sui1-I3N-His	To express eIF1-I3N-His	(3)	
5708	pET-sui1-K60E-His	To express eIF1- K60E-His	(3)	
5709	pET-sui1-R33A-His	To express eIF1- R33A-His	(3)	
5710	pET-sui1-K37E-His	To express eIF1- K37E-His	(3)	
1595	pET28 eIF1 R53S	To express eIF1 R53S	This study	Fig. 5E
1124	YDpU-SUI3	lc <i>SUI3 URA3</i> plasmid	(8)	Fig. 6
1125	YDpU-SUI3-2	lc <i>SUI3-2 (eIF2β-S264Y) URA3</i>	(8)	
1647	YCpL-SUI1 K56A	sc <i>SUI1 LEU2</i> plasmid carrying <i>SUI1 K56A</i>	This study	
1648	YCpL-SUI1 R53S	sc <i>SUI1 LEU2</i> plasmid carrying <i>SUI1 R53S</i>	This study	
1649	YCpL-SUI1 R53S K56A	sc <i>SUI1 LEU2</i> plasmid carrying <i>SUI1 R53S K56A</i>	This study	
1789	YDpW-SUI3	lc <i>SUI3 TRP1</i> plasmid	This study	
1790	YDpW-SUI3-2	lc <i>SUI3-2 (eIF2β-S264Y) TRP1</i>	This study	

Table S2. Yeast *Saccharomyces cerevisiae* strains used in this study – Related to Figure 6

Strain	Genotype	Source
JCY03	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc URA3 SUI1)</i>	(9)
PMY32	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc LEU2 SUI1-K60E)</i>	(10)
PMY33	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc LEU2 SUI1-L96P)</i>	(10)
KAY1057	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc LEU2 SUI1)</i>	This study
KAY1070	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc LEU2 SUI1-K56A)</i>	This study
KAY1071	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc LEU2 SUI1-R53S)</i>	This study
KAY1072	<i>MATa ura3-52 leu2-3 leu2-112 trp1Δ63 his4-301(ACG) sui1Δ::hisG p(sc LEU2 SUI1-R53S K56A)</i>	This study

Table S3. Structural Statistics for eIF3c NTD (fragment B, aa. 36-163) – Related to Figure 1

NMR restraints	
Distance restraints	
Total NOE	790
Intra-residue	272
Inter-residue	
Sequential ($ i - j = 1$)	213
Medium-range ($1 < i - j < 5$)	168
Long-range ($ i - j \geq 5$)	137
Hydrogen bonds restraints ^a	0
Dihedral angle restraints ^a	
ϕ and ψ	14/14
χ^1 and χ^2	0/0
Structure statistics (20 conformers)	
CYANA target function (\AA^2)	0.46
Residual NOE violations	
Number $> 0.1 \text{ \AA}$	5
Maximum (\AA)	0.29
Residual dihedral angle violations	
Number $> 5^\circ$	0
Maximum ($^\circ$)	1.85
AMBER energies (kcal/mol)	
Mean AMBER energy	-4730
Mean restraints violation energy	5.34
Ramachandran plot statistics (%) ^b	
Residues in most favored regions	82.3
Residues in additionally allowed regions	15.6
Residues in generously allowed regions	1.6
Residues in disallowed regions	0.5
Average R.M.S.D. to mean structure (\AA) ^c	
Protein backbone	0.65 ± 0.20
Protein heavy atoms	1.34 ± 0.26

^a Used only in CYANA calculations.

^b Calculated with PROCHECK-NMR.

^c For residues Asp125-Arg155 of eIF3c 36-163.

Table S4. Structural Statistics for eIF1 – Related to Figure 3

NMR restraints	
Distance restraints	
Total NOE	1560
Intra-residue	463
Inter-residue	
Sequential ($ i - j = 1$)	394
Medium-range ($1 < i - j < 5$)	261
Long-range ($ i - j \geq 5$)	442
Hydrogen bonds restraints ^a	0
Dihedral angle restraints ^a	
ϕ and ψ	58/52
χ^1 and χ^2	2/11
Structure statistics (20 conformers)	
CYANA target function (\AA^2)	0.19
Residual NOE violations	
Number $> 0.1 \text{ \AA}$	1
Maximum (\AA)	0.23
Residual dihedral angle violations	
Number $> 5^\circ$	0
Maximum ($^\circ$)	2.60
AMBER energies (kcal/mol)	
Mean AMBER energy	-3633
Mean restraints violation energy	4.96
Ramachandran plot statistics (%) ^b	
Residues in most favored regions	89.3
Residues in additionally allowed regions	9.8
Residues in generously allowed regions	0.5
Residues in disallowed regions	0.4
Average R.M.S.D. to mean structure (\AA) ^c	
Protein backbone	0.48 ± 0.12
Protein heavy atoms	1.10 ± 0.12

^a Used only in CYANA calculations.^b Calculated with PROCHECK-NMR.^c For residues Asn24-Ile30, Leu39-Val69, Ile77-Phe108 of eIF1.

Supplemental Results

The effect of eIF1-K60E on eIF1 binding to its MFC partners

The K60E substitution strongly impairs 40S binding *in vitro*, thereby allowing mis-initiation from UUG codons *in vivo* (Sui⁻ phenotype) (3). Our GST pulldown assays indicated that K60E also disrupts eIF1 binding to eIF2 β -NTT and the eIF5-CTD (Fig. 5E, col. 3); a defect in eIF5-CTD binding was confirmed by CSP assay with ¹⁵N-eIF1-K60E (Fig. S7). It is noteworthy that the K60E substitution essentially eliminates eIF1 binding to the 40S subunit (Figs. 5B and S6B) (3) as well as to the eIF2 β -NTT and eIF5-CTD (Fig. 5B), and yet, confers a less dramatic increase in UUG initiation compared to L96P (Fig. 6A). One possibility is that eIF1-K60E's robust interaction with eIF3c-NTD (Fig. 5B and E) is sufficient to prevent a more dramatic reduction in accuracy for this substitution *in vivo*. Another possibility in light of the proposed role for 3c0-Box6_{Ssu+} in eIF1 release is that the inability of eIF1-K60 to bind eIF2 β -NTT dampened inaccurate UUG initiation by stabilizing the open state, assuming that eIF2 β -NTT contributes to eIF1 release by preventing eIF1 rebinding to the ribosome. These two possibilities must be distinguished by experiments in the future.

Supplemental Experimental Procedures

Plasmid construction – Plasmids used in this study are listed in Table S1 and new plasmids were constructed as follows: For protein purification, the genes encoding various yeast *S. cerevisiae* eIF proteins (wild-type or mutant) or their segments were amplified by PCR using budding yeast genomic DNA. The PCR products were digested with BamHI and NotI and then ligated into suitably cut, modified pET28b vector, in which a SD sequence, initial ATG, hexa-histidine tag or GST tag with a tobacco etch virus (TEV) protease cleavage site had been cloned between XbaI and BamHI restriction sites immediately upstream of the protein coding region (pET28-His or pET28-GST derivatives). pET28b derivatives expressing untagged eIF1 or its mutants were also constructed.

For yeast genetics, YCpL-SUI1 (eIF1) derivatives carrying eIF1 R53 and K56 mutations (used in Fig. 6) were constructed by subcloning the 0.2-kb NdeI-BamHI fragment containing the N-terminal half of eIF1-coding region of the corresponding derivatives of pET28-SUI1 plasmids into the same sites of YCpL-SUI1ΔNde (5). YDpW-SUI3 and YDpW-SUI3-2 were constructed by transferring 1.9-kb NotI-SalI fragment of YDpU-SUI3 and YDpU-SUI3-2 (8) into the same sites of pRS414 (lc *TRP1*).

Yeast strains and methods – Yeast strains used in this study are listed in Table S2. Derivatives of strain KAY1057 [*his4-301(ACG) sui1Δ*] were constructed by transforming JCY03 to Leu⁺ with YCpL-SUI1 (5) or its derivatives carrying the corresponding eIF1 mutations (Table S1), and the resident *SUI1*⁺ *URA3* plasmid was evicted by selecting against *URA3* using the drug 5-FOA. Standard yeast molecular biology methods including growth and β-galactosidase assays were used throughout (11).

For UUG initiation assays, we used histidine auxotrophic assay and β-galactosidase assay, using *his4-301(ACG)* allele and *HIS4*^{AUG} or *his4*^{UUG}-*lacZ* fusion plasmids as reporters, respectively. *Sui*⁻ mutations allow the *his4-301* allele to express using its third codon, UUG, as the start codon. Thus, yeast *his4-301* strains carrying eIF1 mutant alleles (used in Fig. 6A and listed in Table S2) were assayed for histidine auxotrophy. 5 μl of 1.5 A₆₀₀ units of these strains and their 10-fold serial dilutions are spotted onto synthetic complete (SC) medium lacking leucine (panel 1, +His) or the same medium except with trace histidine (3 μM; panel 2, - His) and incubated for 2 and 10 days, respectively. The growth in the - His plate depends on UUG-initiated translation of a histidine enzyme, and hence a phenotypic measurement of UUG initiation. To quantify UUG initiation frequency compared to AUG initiation, we transformed the eIF1 mutant strains with *URA3 HIS4*^{AUG} and *his4*^{UUG}-*lacZ* fusion plasmids and assayed for β-galactosidase, as described (11).

Likewise, to evaluate the combined effect of eIF1-R53S and eIF2β-S264Y (in Fig. 6B), overnight cultures of transformants of KAY1057 (WT eIF1) or KAY1071 (eIF1-R53S) carrying YDpSUI3 (WT eIF2β) or YDpSUI3-2 (eIF2β-S264Y) were spotted similarly onto SC medium lacking uracil (panel 1, +His) or the same medium except with trace histidine (1 μM; panel 2, - His) and incubated for 2 and 9 days, respectively. For *lacZ* reporter assays, we used the KAY1057 and KAY1071 transformants carrying YDpW-SUI3-2 (*SUI3-2 TRP1*) and *URA3 HIS4*^{AUG} or *his4*^{UUG}-*lacZ* fusion plasmids and assayed, as described above.

GST-pulldown assays – GST-pulldown assays were performed as described previously (12). In Fig. 3E,

GST-eIF3c-N₁₋₁₅₆, GST-eIF2β-NTT₁₋₁₄₀, and GST-eIF5-CTD₂₄₁₋₄₀₅ were allowed to bind ³⁵S-labeled eIF1 or its mutant species. The percentage of ³⁵S-eIF1 species pulled down with GST fusion proteins was quantified using a phosphoriager. Alternatively, we used WT and R53S versions of recombinant eIF1 expressed in bacteria for the pulldown assay. In this figure, the values for the binding of each eIF1 mutant are presented relative to those obtained with WT eIF1. Other experiments shown in Fig. 1B, 7A, S1B-C and S8 were done with GST-eIF3c-NTD derivatives and eIF1 and eIF5 that were expressed and purified in *E. coli*. In the case of Fig. 7A, we used an untagged, recombinant form of eIF1 present in *E. coli* lysate, in order to obtain a high concentration required for the competition assay. The amount of eIF1 in the lysate was determined by comparison with known amounts of eIF1 by immunoblotting with anti-eIF1. The same lysate (together with control uninduced lysate) was used for a regular binding assay with eIF1 in Fig. 1B.

Expression and purification of proteins – The pET28-His(TEV) plasmids encoding the desired proteins were employed for transformation of BL21(DE3)RIPL CodonPlus strain (Stratagene). The proteins were expressed in LB medium overnight at 15°C after induction with 0.5 mM IPTG. For His-tagged proteins, harvested cells were re-suspended in Ni-NTA binding buffer (20 mM Tris pH 8.0, 500 mM NaCl, 100 mM urea, 25 mM imidazole and 10 mM β-mercaptoethanol) and lysed using EmulsiFlex homogenizer (Avestin). After centrifugation, the supernatant was loaded onto Ni-NTA agarose (Qiagen) equilibrated with the same buffer. Proteins were eluted by a 25-500 mM linear gradient of imidazole. For GST-tagged proteins, cells were resuspended in GST binding buffer (20mM Tris pH 8.0, 100 mM NaCl and 1 mM DTT) and lysed using EmulsiFlex homogenizer (Avestin). After centrifugation, the supernatant was loaded onto glutathione sepharose (GE) equilibrated with the same buffer. Proteins were eluted by a 0-10 mM linear gradient of reduced glutathione. Peak fractions were incubated overnight with His-tagged TEV protease at room temperature while dialyzing against Ni-NTA low salt buffer (20 mM Tris pH 8.0, 100 mM NaCl, 25 mM imidazole and 10 mM β-mercaptoethanol). After complete cleavage the sample was loaded on Ni-NTA agarose again to remove His tag, His-tagged TEV protease and minor protein contaminants. The complex was then dialyzed against the buffer (20 mM Tris pH 8.0, 100 mM NaCl) for the measurements.

ITC experiments – All calorimetric titrations were carried out on VP-ITC and iTC200 calorimeters (MicroCal). Protein samples were dialyzed against the buffer containing 25 mM Tris pH 8.0 and 100 mM NaCl. The sample cell was filled with 50 μM solution of eIF1 and the injection syringe with 500 μM of the titrating eIF3c. For VP-ITC, each titration typically consisted of a preliminary 3 μl injection followed by 28 subsequent 10 μl injections every 210 seconds. For iTC200, each titration typically consisted of a preliminary 0.4 μl injection followed by 19 subsequent 2 μl injections every 150 seconds. All of the experiments were performed at 20°C. Data for the preliminary injection, which are affected by diffusion of the solution from and into the injection syringe during the initial equilibration period, were discarded. The data were fitted using ORIGIN software.

NMR spectroscopy – For structural determinations, [¹³C, ¹⁵N] eIF1 and [¹³C, ¹⁵N] eIF3c-B₃₆₋₁₆₃ were each concentrated to 0.4 mM in 20 mM sodium phosphate buffer (pH 7.0), containing 150 mM NaCl and 1 mM 1,4-DL-dithiothreitol (DTT), using Amicon Ultra15 filter (3000 MWCO, Millipore). In this study, we used an

untagged, native form of eIF1 by a TEV-protease cleavage method, determined its structure and used it for interaction mapping studies. All NMR data were collected at 298 K on Bruker AVANCE III HD 600, Bruker AVANCE 600 and 800 MHz NMR spectrometers, each equipped with a cryogenic probe. NMR spectra were processed with NMRPipe/NMRDraw (13). Spectral analysis was performed with KUIRA 0.984 (14), a program suite for interactive NMR analysis working with NMRView (15), according to the methods described previously (16). The backbone and side chain ^1H , ^{15}N and ^{13}C resonances of the proteins were assigned by standard double- and triple-resonance NMR experiments (17, 18), and were deposited in the BioMagResDB (BMRB accession numbers 11599 for eIF1 and 11600 for eIF3c-B). Distance restraints were derived from three-dimensional (3D) ^{15}N -edited and ^{13}C -edited nuclear Overhauser effect spectroscopy (NOESY)-HSQC spectra, each measured with a mixing time of 80 ms.

Structure calculations – Structure calculations of eIF1 and eIF3c-B₃₆₋₁₆₃ were performed using CYANA 2.0.17 (19-21). The structure calculations started from 200 randomized conformers, and used the standard CYANA simulated annealing schedule with 40,000 torsion angle dynamics steps per conformer. The 40 conformers with the lowest final CYANA target function values were further refined with AMBER9 (22), using the AMBER 2003 force field and a generalized Born model, as described previously (16). The 20 conformers that were most consistent with the experimental restraints were then used for further analyses. The final structures were validated and visualized by using the PROCHECK-NMR (23) and CHIMERA (24, 25). Detailed experimental data and structural statistics are summarized in Table S3 and S4. The final ensembles of 20 conformers were deposited in the Protein Data Bank (PDB IDs 2rvh for eIF1 and 5H7U for eIF3c-B).

Chemical shift perturbation experiments – All the proteins were dissolved in 20 mM sodium phosphate buffer (pH 7.0), containing 150 mM NaCl and 1 mM DTT. A series of 2D ^1H - ^{15}N HSQC spectra were recorded for the samples containing 70 μM [^{15}N] eIF3c-A₁₋₁₆₃ or [^{15}N] eIF3c-B₃₆₋₁₆₃ and unlabeled eIF1 at the molar ratios of 1.0:0.0, 1.0:0.3, 1.0:0.6, and 1.0:1.0. The samples containing 70 μM [^{15}N] eIF3c-B₃₆₋₁₆₃ and either eIF1-K60E, eIF1-L96P, or eIF1-R53S/K65A at the molar ratios of 1.0:0.0 and 1.0:1.2; and the samples containing 70 μM of either [^{15}N] eIF1, or [^{15}N] eIF1-K60E, [^{15}N] eIF1-R53S and eIF5₂₄₂₋₄₀₅ at the molar ratios of 1.0:0.0 and 1.0:4.0 were also subjected to 2D ^1H - ^{15}N HSQC spectra measurements. These spectra were processed with NMRPipe/NMRDraw¹ and analyzed with Sparky (26). Chemical shift perturbation (CSP), $\Delta\delta$, was defined as $\Delta\delta = [(\Delta\delta_{\text{H}})^2 + (\Delta\delta_{\text{N}}/6.5)^2]^{1/2}$, where $\Delta\delta_{\text{H}}$ and $\Delta\delta_{\text{N}}$ are the chemical shift differences for ^1H and ^{15}N , respectively, and 6.5 is the scaling factor determined from the ratio of the average variances of the ^1H and ^{15}N chemical shifts observed for the 20 common amino acid residues in proteins (27).

To verify the assignment of perturbed amino acids, we used [^{13}C , ^{15}N] eIF3c-C₃₆₋₈₇ and re-assigned amino acids in the presence of eIF1. This helped us verify the perturbation of ~ 10 overlapping chemical shifts. In the ^{15}N -eIF1 CSP experiments with eIF5-CTD, we also used eIF5₂₄₂₋₃₉₆ lacking the C-terminal tail. The CSP observed with this segment was indistinguishable with that observed with eIF5₂₄₂₋₄₀₅ (Fig. 1E, left), indicating that eIF5-CTD, not the C-terminal tail, interacts with eIF1.

Fluorescence Anisotropy Experiments – Initiation factors eIF1A and eIF1 WT and mutant variants of this protein were purified using the IMPACT system (New England Biolabs) as described before (28) using the appropriate pTYB2-derived constructs. eIF1 WT and mutant proteins were labeled at their C termini with cysteine-lysine-fluorescein dipeptide, using the expressed protein ligation system as described previously (29). 40S subunits were purified as described previously (28).

Fluorescence anisotropy measurements of equilibrium binding constants (K_d) were performed using a T-format Spex Fluorolog-3 (J. Y. Horiba) as described previously (29). The excitation and emission wavelengths were 497 and 520 nm, respectively.

The data were fit with a hyperbolic binding equation describing the binding of fluorescently labeled eIF1 mutants to 40S subunits to give K_d values (29). In competition experiments with unlabeled eIF1, the data were fit with a quadratic equation describing the competitive binding of two ligands to a receptor (29).

Analytical ultracentrifugation (AUC) – AUC sedimentation velocity experiments were carried out in the AUC buffer (20 mM sodium phosphate, pH 7.0, 150 mM NaCl), using an Optima XL-I analytical ultracentrifuge equipped with two optical systems, the Rayleigh interference and absorbance systems (Beckman Coulter). For centrifuge, we used an An-50 Ti rotor featuring cells with a standard 12-mm charcoal-epon double sector centerpiece and sapphire windows. Proteins were diluted with the AUC buffer to a final concentration of 20-25 μ M each. After dialysis with the same buffer, the sample was loaded into a cell in the An-50 Ti rotor. The experiments were conducted at 50,000 rpm at a temperature of 293 K. During the runs, changes in the protein concentration gradient were monitored with absorbance at 280 nm. All of raw data were analyzed by the program SEDFIT14.1, with the continuous $C(s)$ distribution model (30). The SEPHAT 10.58d program was used for analysis of the isotherm of weight-average s -values.

Supplemental References

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