

Inhibition of IFN α secretion in cells from patients with juvenile dermatomyositis under TBK1 inhibitor treatment revealed by single-molecular assay technology

Cyril Gitiaux, Vincent Bondet, Nassima Bekaddour, Patrick Nusbaum, Arnaud Hubas, Isabelle Melki, Christine Bodemer, Pierre Quartier, Isabelle Desguerre, Yanick Crow, et al.

▶ To cite this version:

Cyril Gitiaux, Vincent Bondet, Nassima Bekaddour, Patrick Nusbaum, Arnaud Hubas, et al.. Inhibition of IFN α secretion in cells from patients with juvenile dermatomyositis under TBK1 inhibitor treatment revealed by single-molecular assay technology. Rheumatology, 2020, pp.kez508. 10.1093/rheumatology/kez508. pasteur-02376263

HAL Id: pasteur-02376263 https://pasteur.hal.science/pasteur-02376263

Submitted on 22 Jul 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. Inhibition of IFNα secretion in cells from patients with juvenile dermatomyositis under TBK1 inhibitor treatment revealed by Single Molecular Assay technology

Cyril Gitiaux^{1,2*}, Vincent Bondet^{3*}, Nassima Bekaddour⁴, Patrick Nusbaum⁵, Arnaud Hubas⁵, Isabelle Melki^{6,7,10}, Christine Bodemer⁷, Pierre Quartier⁸, Isabelle Desguerre⁹, Yanick J Crow^{10,11}, Jean-Philippe Herbeuval⁴, Darragh Duffy³, Brigitte Bader Meunier^{8*}, Mathieu P Rodero^{*4}

* contributed equally

1: Reference Center for Neuromuscular Diseases, Necker-Enfants Malades Hospital, AP-HP.5, Paris, France. Department of Pediatric Neurophysiology, Necker-Enfants Malades Hospital, AP-HP.5, Paris University, Paris, France.

2 :INSERM U955-Team 10, Department of Neurosciences, Mondor Biomedical Research Institute, Paris-Est University, Créteil, France.

3: Immunobiology of Dendritic Cells, INSERM U1223, Institut Pasteur, Paris, France.

4: Chimie & Biologie, Modélisation et Immunologie pour la Thérapie (CBMIT), Université Paris Descartes, CNRS, UMR8601, Paris, France.

5: Laboratoire de Génétique et Biologie Moléculaires. Hôpital Cochin. Paris, France.

6: General Pediatrics, Infectious Disease and Internal Medicine Department, Hôpital Robert Debre, AP-HP, Paris, France, Referee center for Rheumatic, AutoImmune and Systemic diseases in children (RAISE), Paris, France

7: National Reference Centre for Genodermatosis and Rare Diseases of the Skin (MAGEC), Necker-Enfants Malades Hospital, APHP5, Paris, France. Department of Dermatology, Necker-Enfants Malades Hospital, Paris, France. Imagine Institute, Inserm U 1163, Paris University France. 8: RAISE reference centre, IMAGINE Institute and Pediatric Immunology-Hematology and Rheumatology Unit, Necker hospital, Assistance Publique-Hopitaux de Paris, Paris, France. Paris Descartes University, Paris, France

9: Reference Center for Neuromuscular Diseases, Necker-Enfants Malades Hospital, AP-HP.5, Paris, France. Department of Pediatric Neurology, Necker-Enfants Malades Hospital, AP-HP.5, Paris University, Paris, France.

10: Centre for Genomic and Experimental Medicine, MRC Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, United Kingdom.

11: Laboratory of Neurogenetics and Neuroinflammation, Institut Imagine, Paris Descartes University, Paris, France.

Correspondance to: Mathieu P Rodero, Chimie & Biologie, Modélisation et Immunologie pour la Thérapie (CBMIT), Université Paris Descartes, CNRS, UMR8601,45 rue des saints Pères 75004 Paris, France. Email: <u>mathieu.rodero@parisdescartes.fr</u>

Key message: Detection of IFN α s proteins secreted by cells from JDM patients opens new perspective for drug discovery.

Dear Editor,

A type 1 interferon (IFN) gene signature has been previously demonstrated in the peripheral blood and muscle of patients with juvenile dermatomyositis (JDM), correlating with disease activity scores. However, direct measurement of interferon alpha (IFN α) protein in samples from patients has remained a challenge until recently. We addressed this limitation by optimising an ultrasensitive single molecule array (Simoa) digital enzyme-linked immunosorbent assay (ELISA) utilizing high affinity autoantibodies isolated from APECED patients that recognize all human interferon- α species (1). Using this technology, we were able to detect and quantify serum IFN α in the blood of JDM patients (2), and the blood, cerebrospinal fluid, cell supernatant and tissues from patients with other complex and monogenic interferonopathies (2,3). Interestingly, the median serum concentration of IFN22 was at 56 fg/ml in JDM patients, almost 100 times below the limit of detection of classical anti IFN α ELISAs. In the present study, we evaluated the ability of anti-inflammatory drugs, particularly the TBK1 inhibitor BX795, to control IFNa secretion from cells of JDM patients. Previous studies have shown that TBK1 inhibition controls disease activity in a mouse model of systemic lupus erythematosus (SLE), and interferon signalling in fibroblasts from lupus patients (4) and PBMCs isolated from patients with gain-of-function of STING (3). TBK1 inhibitors are currently in preclinical evaluation for their use in inflammatory diseases such as SLE.

We first demonstrated that our Simoa assay allowed the detection of IFNα secretion by unstimulated peripheral blood mononuclear cells (PBMCs) isolated from JDM patients: culture of frozen PBMCs from all 7 patients tested demonstrated elevated levels of IFNα (median 1319 fg/ml [range: 211.7-5940 fg/ml]) in comparison to controls (median 0.0056 fg/ml [0.0004-45.05 fg/ml]; p=0.0025 (Figure 1A). In order to evaluate the ability of TBK1 to control IFNα secretion by PBMCs isolated from JDM

patients, we cultured PBMCs overnight with or without 2μM of BX795. Inhibition of TBK1 by BX795 resulted in a strong decrease of spontaneous secretion of IFNα in all tested patients (Figure 1B). As in our previous study on cells from STING-mutated patients(3), BX795 did not affect cell viability at this concentration (Figure 1C). We have also previously shown that myogenic precursor cells (MPCs) from JDM patients can produce type I-IFN (5). This may be explained in part by the over expression of nucleic acid sensors, including MDA5 (6)(Figure 1D). Thus, in order to mimic disease-related inflammation, we stimulated MPCs-derived from JDM patients by transfection of the MDA5 ligand poly I:C. After 24 hours, IFNα was measured in the supernatant by Simoa, and cell viability was evaluated by ATP measurement. Upon stimulation, MPCs from 11 of 15 patients demonstrated increased levels of IFNα (median non-stimulated 0.20 fg/ml [0.005-1.9 fg/ml]; median stimulated 1.03 fg/ml [0.015-137.8 fg/ml] p=0.017 (Figure 1E, 1F). IFNα production by MPCs under poly I:C stimulation was decreased by treatment with BX795 (median 1.03 fg/ml [0.015-137.8 fg/ml]; median 0.36 fg/ml [0.003-4.69 fg/ml] p=0.007 (Figure 1F) with no or minor effect on MPCs viability (Figure.1G).

The increased sensitivity to detect IFN α protein by Simoa technology has allowed us to reveal that circulating leukocytes from JDM patients demonstrate spontaneous *ex vivo* production of IFN α , which can be controlled by TBK1 inhibition. Similarly, we were able to measure, for the first time, the production of IFN α by stimulated primary MPCs of JDM patients, and to evaluate the inhibition of this secretion by a TBK1 inhibitor. As such, our data demonstrate that it is now possible to use IFN α secretion as a read-out to evaluate the modulation of interferon by JDM patient-derived material, and to interrogate the effect of potential therapeutic agents. Thus, we envision that Simoa technology will inform drug discovery and personalised approaches to therapy. In another hand, how type I IFNs are involved in JDM pathogenesis remain an open question. Notably, the sources of the interferon in JDM remain unknown, as does the impact of chronic exposure to interferon, even at very low levels, on endothelial and muscular cells within the tissues. The increased sensitivity to detect IFN α proteins by

Simoa should help to identify the interferon producing cells and paves the way for a better understanding of JDM physiopathology.



Figure Legend:

Figure 1:TBK1 inhibition blocks IFN α **secretion by JDM primary cells. (A)** IFN α secreted by PBMCs of 7 JDM patients can be detected by Simoa. **(B)** Treatment with a TBK1 inhibitor strongly reduces IFN α secretion. **(C)** Evaluation of TBK1 inhibition on cell survival. Results from 2 independent experiments on PBMCs from P6 and P7 show no toxic effect. **(D)** MPCs from JDM patients express more IFIH1 transcript than controls. **(E)** Stimulation of patient MPCs with poly I:C induced secretion of IFN α that can be measured by Simoa. **(F)** TBK1 inhibition controls Poly I:C induced IFN α secretion by MPCs,**(G)** with no or minor effect on MPCs viability. TR : transfection reagent; Wilcoxon matched-pairs rank test: * <0,05; **<0,01

Bibliography

- Llibre A, Bondet V, Rodero MP, Hunt D, Crow YJ, Duffy D. Development and Validation of an Ultrasensitive Single Molecule Array Digital Enzyme-linked Immunosorbent Assay for Human Interferon-α. J Vis Exp. 2018 Jun 14;(136).
- Rodero MP, Decalf J, Bondet V, Hunt D, Rice GI, Werneke S, et al. Detection of interferon alpha protein reveals differential levels and cellular sources in disease. J Exp Med. 2017 May 1;214(5):1547–1555.
- Frémond M-L, Uggenti C, Van Eyck L, Melki I, Bondet V, Kitabayashi N, et al. Brief Report: Blockade of TANK-Binding Kinase 1/IKKɛ Inhibits Mutant Stimulator of Interferon Genes (STING)-Mediated Inflammatory Responses in Human Peripheral Blood Mononuclear Cells. Arthritis Rheumatol. 2017 Jun 5;69(7):1495–1501.
- Hasan M, Dobbs N, Khan S, White MA, Wakeland EK, Li Q-Z, et al. Cutting edge: inhibiting TBK1 by compound II ameliorates autoimmune disease in mice. J Immunol. 2015 Nov 15;195(10):4573– 4577.
- Gitiaux C, Latroche C, Weiss-Gayet M, Rodero MP, Duffy D, Bader-Meunier B, et al. Myogenic Progenitor Cells Exhibit Type I Interferon-Driven Proangiogenic Properties and Molecular Signature During Juvenile Dermatomyositis. Arthritis Rheumatol. 2018;70(1):134–145.
- Suárez-Calvet X, Gallardo E, Nogales-Gadea G, Querol L, Navas M, Díaz-Manera J, et al. Altered RIG-I/DDX58-mediated innate immunity in dermatomyositis. J Pathol. 2014 Jul;233(3):258–268.

Conflicts of interest:

The authors have no conflicts of interest to disclose.

Funding and acknowledgment:

JPH and NB were supported by the Agence National de la Recherche sur le SIDA et les Hépatites ANRS (AAP 2017-166). This work was also supported by SATT idfinnov (www.idfinnov.com) through the

Project under Grant n° 303. D.D. and Y.J.C acknowledge support from the Agence National de Recherche ANR (CE17001002) and Immunoqure for provision of mAbs for Simoa assays.