



HAL
open science

Vertical transmission of arboviruses in mosquitoes: A historical perspective

Sebastian Lequime, Louis Lambrechts

► **To cite this version:**

Sebastian Lequime, Louis Lambrechts. Vertical transmission of arboviruses in mosquitoes: A historical perspective. *Infection, Genetics and Evolution*, 2014, 28, pp.681-690. 10.1016/j.meegid.2014.07.025 . pasteur-01976254

HAL Id: pasteur-01976254

<https://pasteur.hal.science/pasteur-01976254>

Submitted on 8 Feb 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Vertical transmission of arboviruses in mosquitoes: a historical perspective

Sebastian Lequime^{1,2*} and Louis Lambrechts^{1*}

¹ *Insect-Virus Interactions Group, Department of Genomes and Genetics, Institut Pasteur – Centre National de la Recherche Scientifique, Unité de Recherche Associée 3012, Paris, France*

² *University Pierre et Marie Curie, Cellule Pasteur UPMC, Paris, France*

*Corresponding author. Address: G5 Insect-Virus Interactions, Institut Pasteur, 28 rue du Docteur Roux, 75724 Paris Cedex 15, France.

E-mail addresses: sebastian.lequime@pasteur.fr (S. Lequime); louis.lambrechts@pasteur.fr (L. Lambrechts).

Abstract

1
2
3
4 Arthropod-borne viruses (arboviruses) are mainly transmitted horizontally among vertebrate
5 hosts by blood-feeding invertebrate vectors, but can also be transmitted vertically in the vector
6
7 from an infected female to its offspring. Vertical transmission (VT) is considered a possible
8
9 mechanism for the persistence of arboviruses during periods unfavorable for horizontal
10
11 transmission, but the extent and epidemiological significance of this phenomenon have
12
13 remained controversial. To help resolve this question, we reviewed over a century of
14
15 published literature on VT to analyze historical trends of scientific investigations on
16
17 experimental and natural occurrence of VT in mosquitoes. Our synthesis highlights the
18
19 influence of major events of public health significance in arbovirology on the number of VT
20
21 publications. Epidemiological landmarks such as emergence events have significantly
22
23 stimulated VT research. Our analysis also reveals the association between the evolution of
24
25 virological assays and the probability of VT detection. Increased sensitivity and higher-
26
27 throughput of modern laboratory assays resulted in enhanced VT detection. In general, VT
28
29 contribution to arbovirus persistence is likely modest because vertically infected mosquitoes
30
31 are rarely observed in nature. Taken together, however, our results call for caution when
32
33 interpreting VT studies because their conclusions are context- and method-dependent.
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Keywords: Arbovirus; mosquito; vertical transmission; systematic review; historical trend.

Highlights

- The extent and epidemiological importance of arbovirus VT are still debated.
- Arboviral emergence events have stimulated VT research.
- More recent laboratory assays are associated with enhanced VT detection.
- Typically <0.1% of mosquitoes are vertically infected in nature.
- The outcome of VT studies is context- and method-dependent.

1. Introduction

According to the United States Center for Disease Control and Prevention (CDC) Arbovirus Catalog (Centers for Disease Control and Prevention, 2010), there are currently at least 530 identified arthropod-borne viruses (arboviruses), of which about a hundred cause human disease. Among them, four major viral genera account for the majority of arboviral diseases: *Flavivirus* (e.g., dengue, West Nile, Japanese encephalitis, and yellow fever viruses), *Alphavirus* (e.g., chikungunya, Eastern equine encephalomyelitis, Western equine encephalomyelitis and Venezuelan equine encephalitis viruses), *Orthobunyavirus* (e.g., California encephalitis and LaCrosse viruses) and *Phlebovirus* (e.g., Rift Valley fever and sandfly fever viruses).

During the past few decades, several arboviruses have emerged globally and are now considered among the most important public health concerns for the 21st century (Gubler, 2002). Dengue, for example, has become the most prevalent arthropod-borne viral disease of humans over the last few decades (Messina et al., 2014); it has recently been estimated that there are 390 million human dengue infections each year (Bhatt et al., 2013). With only a few licensed vaccines and virtually no therapeutics available, antivectorial measures are often the only way to prevent arboviral diseases. Historically, however, the implementation of vector control measures has generally been difficult to sustain.

Arboviruses are naturally maintained in a transmission cycle between vertebrate and arthropod hosts (Gubler, 2001). The majority of arthropod hosts, generally referred to as vectors, are blood-feeding mosquitoes. Rather than a simple alternation within a single host-vector pair, arbovirus transmission often occurs through highly complex transmission networks that include various hosts and vectors (Diaz et al., 2012). Humans in particular, are not necessarily at the center of the transmission network and may only be incidental hosts

1 (e.g., West Nile virus). Whereas some host or vector species are central to epidemic arbovirus
2 transmission, others can be part of alternative transmission pathways, participating in the
3 maintenance of the virus in nature during inter-epidemic periods. As an example, for some
4 authors, fox squirrels may contribute to alternative transmission of West Nile virus in
5 suburban communities (Root et al., 2006; 2007). In many regions of the world, climatic
6 conditions do not allow mosquito reproductive activity all year long. During the dry season in
7 tropical areas or the cold season in temperate regions, the absence or low density of adult
8 mosquitoes is unlikely to support continuous host-to-vector (horizontal) transmission (Leake,
9 1984). Survival of mosquitoes during dry and cold seasons involves different physiological
10 and/or behavioral mechanisms that may impact virus transmission differently. Besides,
11 arboviral infections in vertebrate hosts typically produce a short-lived viremic period that
12 eventually results in immunization of the host. A high level of herd immunity in the host
13 community may thus prevent transmission above the minimum level required for sustained
14 horizontal transmission. The existence of reservoir host species, alternative transmission
15 mechanism or virus re-introduction, have been proposed to explain the maintenance of
16 arboviruses during unfavorable periods or when herd immunity is high (for review, see
17 Reeves, 2004).

18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41 One popular hypothesis to explain the persistence of arboviruses during unfavorable
42 periods is the occurrence of vertical transmission (VT) in the arthropod vector. In this article,
43 we define VT as the transmission of an arbovirus from an infected female mosquito to its
44 offspring, regardless of the underlying mechanism. VT may occur through two main
45 mechanisms. Transovarial transmission (TOT) occurs when the virus infects the germinal
46 tissues of the female mosquito, whereas trans-egg VT takes place during oviposition in the
47 fully formed egg (Rosen, 1988). TOT typically achieves a higher efficiency of VT than trans-

1 egg mechanisms especially when the germ cells are permanently infected so that most of the
2 offspring are infected in the following generation (Tesh, 1984).
3

4 Under the VT scenario, the arbovirus present in the mosquito eggs, larvae or adults,
5 including nulliparous females entering diapause, may survive throughout the unfavorable
6 period without the need for a vertebrate host. Such a mixed-mode transmission (i.e., both
7 horizontal and VT) is widespread among symbionts across taxa (Ebert, 2013). Here symbiosis
8 is defined as any type of persistent biological interaction, which includes mutualistic,
9 commensalistic and antagonistic relationships. Although the infection cost is often modest to
10 the vector, arboviruses are considered parasites of mosquitoes (Lambrechts and Scott, 2009).
11 Combining horizontal with VT enlarges considerably the range of ecological conditions in
12 which a symbiont can persist. In host species with diapause or discrete generations, VT may
13 allow the symbiont to endure periods when horizontal transmission is not possible. Trade-offs
14 between the two modes of transmission have been documented but are not universal (Ebert,
15 2013). In addition to ecological factors that may favor horizontal over VT (e.g., climate),
16 theory suggests that VT should be reduced in arboviruses with complex transmission
17 networks because horizontal transmission among genetically disparate hosts hinders co-
18 adaptation between vertically transmitted viruses and their hosts.
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40

41 Both the very existence and the epidemiological significance of arbovirus VT have
42 remained controversial since it was first suggested in the scientific literature over a century
43 ago, at the onset of arbovirology. Carlos Finlay, who first introduced the idea of vectorial
44 transmission of yellow fever virus by mosquitoes in 1881, extended his theory in 1899,
45 suggesting that the yellow fever agent could be transmitted by an infected mosquito to its
46 progeny (Finlay, 1899). During their investigations in Cuba, the Yellow Fever U.S. Army
47 Commission proved Carlos Finlay's original theory right in 1901 and experimentally tested,
48 albeit unsuccessfully, the possibility of yellow fever virus VT in mosquitoes (Reed et al.,
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1901). The same year, the Cuban physician Juan Guiteras also failed to succeed in demonstrating yellow fever virus VT (Guiteras, 1901).

Between 1901 and 1905, a group of French scientists from the Pasteur Institute carried out studies on yellow fever in Rio de Janeiro, Brazil. Among them, Paul-Louis Simond and Emile Marchoux finally demonstrated the VT hypothesis. They stated, however, that in their opinion the phenomenon was certainly infrequent (Marchoux and Simond, 1906; 1905). Following this discovery, many have tried to replicate the experiment, but none have succeeded (Davis and Shannon, 1930; Frobisher et al., 1931; Hindle, 1930; Rosenau and Goldberger, 1906).

The possibility of VT of other arboviruses was also investigated during the first half of the 20th century. Whereas early studies on dengue virus failed to provide evidence of VT (Siler et al., 1926; Simmons et al., 1931), a Japanese team demonstrated VT of Japanese encephalitis virus (Mitamura et al., 1939), but their work, published in German in a Japanese journal on the eve of World War II, went unnoticed by the scientific community. Studies on VT resumed in the 1950s and 1960s with a team from the Communicable Disease Center (ancestor of CDC) that was investigating viruses responsible for encephalitis (alphaviruses, such as Eastern equine encephalomyelitis, Western equine encephalomyelitis or Venezuelan equine encephalitis viruses; flaviviruses, such as St. Louis encephalitis virus). Their conclusions, however, were inconsistent with both positive (Chamberlain et al., 1956b; Kissling et al., 1956; 1957) and negative results (Chamberlain et al., 1956a; Chamberlain and Sudia, 1957; Chamberlain et al., 1959). The outcome of these studies did not seem to depend on the virus under consideration.

In 1972, Robert B. Tesh and colleagues provided a clear demonstration of vesicular stomatitis virus VT in *Lutzomyia* sandflies (Tesh et al., 1972). The next year, another team led by Douglas M. Watts, published evidence of VT for LaCrosse virus (*Orthobunyavirus*) in

1 experimentally infected *Aedes triseriatus* mosquitoes (Watts et al., 1973). Research on
2 arbovirus VT by mosquito vectors has been vigorous ever since (Fig. 1), although a closer
3
4 look reveals considerable heterogeneity associated with historical events and the evolution of
5
6 laboratory assays used to detect VT experimentally or in a natural setting.
7

8
9 In the present study, we conducted a systematic review of over a century of published
10
11 literature on arbovirus VT to analyze quantitatively historical trends of research on
12
13 experimental and natural occurrence of VT in mosquitoes.
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

2. Material and methods

2.1. Literature search

Between 22 June 2011 and 25 September 2013, a systematic literature search was conducted in NCBI PubMed, ISI Web of Science, Armed Forces Pest Management Board Literature Retrieval System and Pasteur Institute Media Library. Citations in the identified articles were also examined individually in order to recover additional references. When the article was not found using the databases mentioned above, the corresponding authors and/or journal were contacted to obtain a copy of the publication. Older publications were found through Internet Archive (<http://www.archive.org/>).

Publications were searched regardless of their language, including English, French, German, Japanese and Chinese. Japanese and Chinese publications without a full abstract in English were excluded for practical reasons.

The review focused on arboviruses transmitted by mosquitoes and therefore articles dealing with VT in ticks or other arthropods were excluded. Likewise, publications about insect-specific viruses were excluded. Within mosquito-borne arboviruses, the review was restricted to VT in three main arboviral families: *Bunyaviridae*, *Flaviviridae* and *Togaviridae*.

2.2. Databases

Three databases were created in MySQL using the Sequel Pro© software. Contents of the three databases are described below.

2.2.1. Historical trends. To analyze historical trends in VT research, a first database (database #1) was built that contains all publications related to VT according to the inclusion criteria indicated above. For each individual publication contained in this database, basic bibliographical data, virus taxonomy and study type were recorded. Study type consisted of

1 five categories: ‘Experimental’ studies looked for evidence of VT based on laboratory
2 experiments under controlled environments; ‘Natural’ studies attempted to demonstrate VT in
3 nature, by collecting immature, male or overwintering female mosquitoes; ‘Both’ are studies
4 that conducted both experimental and natural investigations; ‘Modeling’ studies used
5 mathematical models to evaluate the potential role of VT in arbovirus epidemiology;
6 ‘Reviews & Opinion’ articles synthesized earlier work and/or commented the possibility of
7 VT.
8
9

10
11
12
13
14
15
16
17 2.2.2. *Evolution of virological assays.* In order to examine how the evolution of
18 laboratory assays in virology influenced the outcome of VT studies, two additional databases
19 were assembled. Database #2 consists of ‘experimental’ studies and database #3 consists of
20 ‘natural’ studies. Both were derived from database #1, but were restricted to the
21 ‘experimental’, ‘natural’, and ‘both’ types of studies. Additionally, inclusion criteria were
22 more stringent for databases #2 and #3 than for database #1. Databases #2 and #3 only
23 included publications that specified mosquito and virus species tested, sample size and
24 detection technique (the full list of studies included in these databases is provided in
25 **Supplementary Table 1**). A total of 32 and 16 factors were included in databases #2 and #3,
26 respectively, and each unique combination of factors was considered a different entry (the full
27 list of factors recorded is provided in **Supplementary Table 2**). Laboratory assays used for
28 virus detection were divided into four broad categories. ‘Animal’ assays refer to the detection
29 of pathological effects (including death) in laboratory animals, usually suckling mice,
30 following inoculation with mosquito extracts; ‘Cellular’ assays refer to the detection of
31 cytopathological effects in cell culture *in vitro*, following inoculation with mosquito extracts;
32 ‘Immunological’ assays rely on the detection of viral antigens by antibodies (e.g.,
33 immunofluorescence assays) with or without previous amplification in cell culture or animal
34 tissues; ‘Molecular’ assays refer to detection of viral nucleic acids, generally by reverse
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1 transcription (RT)-PCR. Whereas animal/cellular assays detect infectious virus and molecular
2 assays detect viral nucleic acids (non-infectious), immunological assays include both
3
4 detection of viral antigens in primary samples (non-infectious) and detection of viral antigens
5
6 following amplification *in vivo* (e.g., intra-thoracic inoculation of *Toxorhynchites* mosquitoes)
7
8 or in cell culture (infectious). Whether the use of infectious or non-infectious assays
9
10 influenced the outcome of VT studies was also evaluated.
11
12
13
14
15
16

17 **2.3. Statistical analyses**

18
19 All statistical analyses were performed in the statistical environment R, version 3.0.2
20
21 (<http://www.r-project.org/>).
22
23

24 *2.3.1. Historical trends.* To normalize the overall scientific production about VT of
25 dengue, West Nile and chikungunya viruses, the yearly number of publications between 1950
26
27 and 2013 was obtained through queries to the ISI Web of Science version 15.13.1 database,
28
29 using the keywords ‘dengue’, ‘West Nile virus’ and ‘chikungunya’, respectively. For each
30
31 virus, the normalized yearly number of publication was calculated as the ratio of the yearly
32
33 number of publications about VT of this virus over the total yearly number of publications
34
35 about the virus. The normalized yearly number of publications was analyzed using non-
36
37 parametric Wilcoxon tests.
38
39
40
41
42

43 *2.3.2. Evolution of virological assays.* The evolution of laboratory assays used for VT
44
45 detection in databases #2 and #3 was visualized by plotting the yearly proportion of each
46
47 assay category (i.e., animal, cellular, immunological and molecular) over time. To weight
48
49 each category according to the sample size, the yearly proportion of each assay category was
50
51 calculated relative to the total number of individual mosquitoes tested for VT (i.e., the relative
52
53 likelihood of each category over time). Smoothed curves were obtained by kernel density
54
55
56
57
58
59
60
61
62
63
64
65

1 estimation using a Gaussian kernel and the bandwidth selection method proposed by Sheather
2 and Jones (1991).
3

4 The association between assay category and VT detection probability was further
5 explored with a subset of databases #2 and #3. The subset consisted of the *Aedes-Flavivirus*
6 pair, which is the most frequent vector-virus association in the databases (49.7% and 31.8% of
7 entries in databases #2 and #3, respectively). Analysis of the relationship between assay
8 category and VT detection probability was restricted to the *Aedes-Flavivirus* pair to rule out
9 the potential confounding effect of a differential VT probability among arbovirus families.
10 Indeed, some arbovirus families may be vertically transmitted more efficiently than others
11 (Turell, 1988). If VT of arboviruses with higher VT efficiency were predominantly examined
12 using one particular virological assay, then a spurious association would be detected between
13 this assay and higher VT detection probability. In experimental studies, only three assay
14 categories were compared for the *Aedes-Flavivirus* pair because the molecular assay category
15 was not represented in database #2. VT occurrence was defined as the proportion of database
16 entries that found evidence of VT. Evolution of VT detection over time for the *Aedes-*
17 *Flavivirus* pair was represented as the relative likelihood of yearly VT occurrence. The
18 relative likelihood function was estimated by kernel density estimation as described above.
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40

41 To disentangle the respective effects of assay and sample size, VT occurrence was
42 analyzed with a generalized linear model that included the \log_{10} -transformed sample size, the
43 assay category and their interaction. As VT occurrence is a binary variable (0=VT undetected,
44 1=VT detected), the model was fitted with a binomial error distribution and a logit link
45 function. Statistical significance of the effects was assessed by analysis of deviance (Hastie
46 and Pregibon, 1991). The same model was used to evaluate the influence of infectious
47 compared to non-infectious assays.
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

To compare sample sizes between assay categories, distributions of \log_{10} -transformed sample sizes for each assay category were compared with non-parametric Kruskal-Wallis tests. When significant, pairwise Wilcoxon tests were subsequently performed with a Bonferroni correction of the p -values.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

3. Results and discussion

3.1. Summary description of databases

The primary database (database #1) includes 257 articles related to arbovirus VT published between 1899 and 2013 (the full list of publications is provided in Supplementary Table 1). Two additional databases were derived from database #1 that were restricted to ‘experimental’ and ‘natural’ types of studies (database #2 and #3, respectively) and had more stringent inclusion criteria (see material and methods).

Database #2 includes a total of 1,119 entries from 94 distinct publications that examined VT in an experimental setting. Overall, VT occurrence was 60.2% in database #2 (i.e., evidence of VT was found in 60.2% of entries). Virological assays consisted of 58.5% immunological, 23.6% cellular, 16.8% animal and 1.1% molecular assays. The most frequent vector-virus pairs in database #2 were *Aedes-Flavivirus*, *Culex-Flavivirus*, *Aedes-Orthobunyavirus* and *Aedes-Alphavirus* that are represented by 49.7%, 22.6% 18.9% and 5.1% of entries, respectively. The remaining 3.7% consist of 11 less studied vector-virus pairs. The median VT prevalence, estimated as the number of positive pools over the total number of mosquitoes tested, was 0.15%, 0.0%, 13.5% and 0.0% in *Aedes-Flavivirus*, *Culex-Flavivirus*, *Aedes-Orthobunyavirus* and *Aedes-Alphavirus* pairs, respectively.

Database #3 includes a total of 368 entries from 97 distinct publications that examined VT in a natural setting. Overall, VT occurrence was 37.5% in database #3 (i.e., evidence of VT was found in 37.5% of the entries). Virological assays consisted of 29.6% animal, 28.5% immunological, 25.3% cellular and 16.3% molecular assays. *Aedes-Flavivirus*, *Culex-Flavivirus*, *Aedes-Orthobunyavirus* and *Aedes-Alphavirus* pairs are represented by 31.8%, 21.2% 16.8% and 2.7% of database #3 entries, respectively. The remaining 27.5% consist of 19 less studied vector-virus pairs. The median VT prevalence was 0.03%, 0.0%, 0.0% and

0.06% in *Aedes-Flavivirus*, *Culex-Flavivirus*, *Aedes-Orthobunyavirus* and *Aedes-Alphavirus* pairs, respectively. The 10- to 1000-fold lower estimates of VT prevalence observed in natural studies likely reflect two different processes. First, the infection prevalence in the previous adult generation is generally close to 100% in experimental studies because mothers are experimentally exposed to the arbovirus before VT is assayed in the their progeny. The prevalence of arbovirus infection in wild mosquito populations, by contrast, is typically low and the prevalence of VT infection observed in a given generation actually measures the product of adult population prevalence and VT efficiency. For example, only 0.1% of field-caught *Ae. aegypti* were found to be infected by dengue virus in an endemic region of Thailand (Yoon et al., 2012). Second, experimental VT studies possibly overestimate VT efficiencies because they tend to focus on naturally efficient combinations of vector-virus strains. VT efficiency may be further enhanced in experimental studies due to the unnatural mode of infection commonly used in the parental generation (i.e., intrathoracic inoculation). Moreover, VT efficiency can be quickly selected in the laboratory. For example, dengue virus VT efficiency increased more than 4-fold within two *Ae. aegypti* generations (Joshi et al., 2002). ‘Stabilized’ infections have also been described for arboviruses of the California serogroup (*Bunyaviridae: Orthobunyavirus*), such as California encephalitis virus in *Ae. dorsalis* (Turell et al., 1982) and San Angelo virus in *Ae. albopictus* (Tesh and Shroyer, 1980). Whereas TOT occurred in 10-20% of the progeny of horizontally infected females, transovarially infected daughters in the following generations consistently transmitted virus to over 90% of their progeny. The proposed mechanism is that following a systemic infection by horizontal transmission, only a small fraction of the developing oocytes of a female become infected transovarially. However, if the germinal cells of a female become infected, then nearly 100% of her progeny are infected. It has been argued that most experimental studies of TOT have in fact underestimated VT efficiency because they considered ‘non-stabilized’

1 infections (Tesh, 1984). An additional complicating factor is that TOT may not occur during
2 the first gonotrophic cycles of orally infected females because the ovaries only become
3 infected during the subsequent gonotrophic cycles. Only considering the first batch of eggs,
4 usually free of virus, would also underestimate VT efficiency in this case (Tesh, 1984).
5
6
7
8
9

10 11 **3.2. Historical trends**

12 We first examined whether epidemiological landmarks, such as emergence events,
13 large outbreaks or a significant change in geographic distribution of an arbovirus, influenced
14 VT research. We considered three major epidemiological events in the recent history of
15 arbovirology: the 1981 dengue hemorrhagic fever (DHF) epidemic in the Americas, the 1999
16 West Nile virus emergence in North America, and the 2005 chikungunya epidemic in the
17 Indian Ocean region. Whereas dengue was already a growing problem in the tropics in the
18 early 1980s (Messina et al., 2014), the emergence of West Nile virus in 1999 and that of
19 chikungunya in 2005 were sudden and largely unexpected.
20
21
22
23
24
25
26
27
28
29
30
31
32
33

34 *3.1.1. DHF in the Americas.* Dengue viruses (*Flaviviridae: Flavivirus*) consist of four
35 serotypes (DENV-1 to -4) that are currently estimated to collectively cause about 96 million
36 symptomatic cases of dengue fever each year, including the severe form DHF (Bhatt et al.,
37 2013). Dengue viruses are primarily transmitted by the mosquito *Ae. aegypti*, which was
38 largely eliminated from Central and South America during the 1950s and 1960s by the
39 eradication program of the Rockefeller Foundation. Approximately ten years after the end of
40 the eradication program, however, dengue epidemics started to occur in the Americas, quickly
41 followed by hyperendemicity and the emergence of DHF (Bennett et al., 2010). The first
42 cases of DHF were reported in Cuba during a DENV-2 epidemic in 1981 (Guzmán et al.,
43 1999).
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
This epidemiological event did not appear to have an obvious impact on dengue research overall, as shown by a query with the keyword ‘dengue’ to the ISI Web of Science database (Fig 2.). By contrast, research on dengue virus VT markedly increased after 1981 (Fig. 2). The normalized yearly number of VT publications was significantly larger during the period 1982-2013 than during the period 1950-1981 (Wilcoxon test, $p = 2.0 \times 10^{-8}$). We cannot exclude the existence of a concomitant factor, however, because this effect was no longer significant when considering a period spanning 5 years before and 5 years after the event (Wilcoxon test, $p = 1.2 \times 10^{-1}$). The burst of VT research after the early 1980s may have been driven by studies investigating the possibility of dengue virus maintenance in areas where *Ae. aegypti* re-established, following ‘rediscovery’ of arbovirus VT in the previous decade (i.e., a delayed consequence of the Watts et al. 1973 publication). Despite a lack of evidence in the early studies (Siler et al., 1926; Simmons et al., 1931), the VT hypothesis has been particularly popular in the case of dengue. Although a sylvatic transmission cycle exists whereby sylvatic dengue strains circulate between arboreal mosquitoes and non-human primates, there is no identified animal reservoir of dengue virus strains responsible for human epidemics (Vasilakis and Weaver, 2008). This probably explains why VT has often been suggested to explain dengue virus maintenance during inter-epidemic periods.

41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
3.1.2. West Nile virus emergence in North America. West Nile virus (*Flaviviridae: Flavivirus*) is primarily transmitted between birds and ornithophilic mosquitoes but can occasionally infect humans and horses, which are dead-ends for the virus (World Health Organization, 2011). Following its introduction in North America, it is estimated that between 1999 and 2010, West Nile virus infected 1.8 million people, resulting in 1,308 deaths (Kilpatrick, 2011). Research on this virus clearly increased after the North-American emergence (Fig. 3) but the normalized yearly number of studies on West Nile virus VT also increased significantly (Fig. 3; Wilcoxon test, $p = 6.1 \times 10^{-9}$). The effect was also significant

1 when considering a period spanning 5 years before and 5 years after the event (Wilcoxon test,
2 $p = 3.7 \times 10^{-3}$). More frequent studies on West Nile virus VT in *Culex* mosquitoes after the
3
4 1999 epidemic have probably been stimulated by the search for a possible arthropod reservoir.
5
6 Adult female mosquitoes are not active all year long in temperate regions of North America,
7
8 yet overwintering persistence of West Nile virus transmission has been observed for over a
9
10 decade (Reisen, 2013). The role of diapausing female *Culex* mosquitoes as a West Nile virus
11
12 reservoir is one of the hypotheses to explain West Nile virus overwintering in North America
13
14 (Reisen, 2013). Indeed, infected diapausing females have been reported in nature (e.g.,
15
16 Fechter-Leggett et al., 2012). VT is a prerequisite for this mechanism to operate, because
17
18 *Culex* females do not blood feed prior to diapause (Mitchell and Briegel, 1989; Nelms et al.,
19
20 2013). In warmer regions of North America, however, female *Culex* mosquitoes do not
21
22 necessarily enter diapause and may instead become quiescent, sometimes after taking a blood
23
24 meal (Eldridge, 1966; Nelms et al., 2013).
25
26
27
28
29
30

31 *3.1.3. Chikungunya emergence in the Indian Ocean region.* Chikungunya virus
32
33 (*Togaviridae: Alphavirus*) is an arbovirus originating in Africa that primarily circulates
34
35 between non-human primates and arboreal mosquitoes. In 2004, it emerged in the human
36
37 population on the coast of Kenya and was subsequently responsible for a vigorous epidemic in
38
39 the Indian Ocean region between 2005 and 2007. On La Réunion Island, 34.4% of the
40
41 population was infected overall, while 1.4 million cases were reported in 2006 in India
42
43 (Pialoux et al., 2007). The global emergence of chikungunya that followed the 2005 outbreak
44
45 triggered a renewed scientific interest for chikungunya virus, as shown by a query with the
46
47 keyword ‘chikungunya’ to the ISI Web of Science database (Fig. 4). Although the normalized
48
49 yearly number of publications on chikungunya virus VT after 2005 remains relatively low
50
51 (due to the dramatic increase in the total number of chikungunya publications used for
52
53 normalization), the frequency of VT publications significantly increased (Fig. 4; Wilcoxon
54
55
56
57
58
59
60
61
62
63
64
65

1 test, $p = 2.9 \times 10^{-3}$). The effect was also significant when considering a period spanning 5
2 years before and 5 years after the event (Wilcoxon test, $p = 3.6 \times 10^{-2}$). Increase of VT
3 research after the 2005 epidemic was, however, less dramatic than in the cases of the 1981
4 DHF epidemic in the Americas and the 1999 emergence of West Nile virus in North America.
5
6 This may be a consequence of earlier studies, which concluded that *Alphavirus* VT rarely
7 occurs and is unlikely to be epidemiologically significant when it does (Turell, 1988). Thus,
8
9 the modest increase of research on chikungunya virus VT after 2005 may reflect a general
10 lack of interest and/or a significant proportion of negative results, a well known bias in
11 systematic reviews (Littell et al., 2008). The risk of chikungunya virus emergence in Europe,
12 as illustrated by the Italian outbreak in 2007 (Rezza et al., 2007) and the current emergence in
13 the Americas (Leparc-Goffart et al., 2014) may stimulate additional research on chikungunya
14 virus VT.
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30

31 **3.3. Evolution of virological assays**

32 For some authors (Bocquet and Molero, 1996), the expansion of arbovirus VT research
33 in the 1970s may be related to the development of new investigation methods, notably with
34 the introduction of immunological reactions in laboratory assays. Overall, virological assays
35 used for VT detection changed dramatically over the last century (Fig. 5). In the 1970s,
36 traditional animal models were replaced by immunological methods that became
37 progressively more common through the 1990s. Immunological assays were then replaced by
38 cellular and molecular assays in the 2000s. Although the use of animal, cellular and
39 immunological assays followed a relatively similar evolution in both natural and experimental
40 studies of arbovirus VT, molecular assays have seldom been used in experimental studies to
41 date (Fig. 5A).
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

The advent of novel virological methods such as PCR-based molecular assays may have facilitated VT detection due to improved sensitivity and higher-throughput capacity. It is worth noting that molecular assays only detect viral RNA and therefore do not provide evidence of infectious virus unless they are confirmed by virus isolation. This is also true for immunological assays that detect viral antigens without direct evidence for infectivity. VT being a relatively rare event (Bocquet and Molero, 1996), one might expect that the likelihood of VT detection would have increased with the evolution of assay performance. We tested this hypothesis in the subset of studies that examined *Flavivirus* VT in *Aedes* mosquitoes. The *Aedes-Flavivirus* association is the most widely represented in the published literature and has been investigated with a variety of laboratory assays. The probability of VT detection in *Aedes-Flavivirus* studies significantly increased over time, both in experimental (Fig. 6A) and natural studies (Fig. 6B). Although a causal relationship cannot be conclusively inferred, these increasing trends coincide with the advent of cellular and immunological assays in experimental studies (Fig. 5A) and of cellular and molecular assays in natural studies (Fig. 5B). Thus, our analyses support the hypothesis that the probability of VT detection has been influenced by the evolution of virological assays.

The probability of VT detection could have been influenced by the sensitivity of the assay and/or the ability to process a larger number of samples by modern assays. More recent detection methods such as cellular or immunological assays used in experimental studies are indeed associated with larger sample sizes (Fig. 7A; Kruskal-Wallis test, $p = 5.2 \times 10^{-5}$; pairwise Wilcoxon tests: $p = 6.4 \times 10^{-3}$ and $p = 3.7 \times 10^{-5}$ for animal vs. cellular and animal vs. immunological assays, respectively, $p = 6.3 \times 10^{-1}$ for cellular vs. immunological assays). Similarly, in natural studies, significantly larger sample sizes are found in studies based on molecular assays (Fig. 7B; Kruskal-Wallis test, $p = 2.9 \times 10^{-2}$; pairwise Wilcoxon tests: $p = 2.6 \times 10^{-2}$ for animal vs. molecular assays, $p = 9.5 \times 10^{-1}$ for animal vs. cellular assays, $p = 2.3$

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

$\times 10^{-1}$ for animal vs. immunological assays, $p = 1.0$ for immunological vs. cellular assays, $p = 2.5 \times 10^{-1}$ for molecular vs. cellular assays, and $p = 9.6 \times 10^{-1}$ for molecular vs. immunological assays). To determine the respective contributions of sample size and detection assay to changes in VT detection probability, VT occurrence in the databases was analyzed with a model that incorporated both factors and their interaction. In experimental studies, we found that VT occurrence was statistically significantly influenced by both the \log_{10} -transformed sample size ($p = 2.5 \times 10^{-7}$) and the detection assay ($p = 3.7 \times 10^{-2}$), but not by their interaction ($p = 4.7 \times 10^{-1}$). Therefore, the increase in VT detection probability did not simply result from a higher-throughput capacity of assays but also from improved assay sensitivity. In natural studies, VT occurrence was significantly influenced by the \log_{10} -transformed sample size ($p = 3.9 \times 10^{-4}$), but not by the detection assay ($p = 2.2 \times 10^{-1}$) or their interaction ($p = 7.2 \times 10^{-2}$). In this case, the observed increase in VT detection probability primarily resulted from a significant increase in the number of samples tested.

We also evaluated whether the use of infectious or non-infectious assays influenced the probability of VT detection. Both animal and cellular assays detect infectious virus, whereas molecular assays detect viral nucleic acids (non-infectious). Immunological assays include both detection of viral antigens in primary samples (non-infectious) and detection of viral antigens following amplification *in vivo* or in cell culture (infectious). We did not find a statistically significant effect of whether the assay was infectious or non-infectious, both in experimental studies ($p = 1.8 \times 10^{-1}$) and in natural studies ($p = 1.8 \times 10^{-1}$), neither did we find an interaction with the \log_{10} -transformed sample size ($p = 1.5 \times 10^{-1}$ and $p = 8.6 \times 10^{-2}$, respectively).

4. Conclusions

Arbovirus VT in mosquitoes has rarely taken center stage in arbovirology, but has remained a debated topic since it was initially discovered more than a century ago. In the present study, fortunately, controversy about the extent and epidemiological importance of VT for arbovirus epidemiology has likely alleviated publication bias, a well-known pitfall of meta-analytical studies due to the under reporting of negative results. Despite their intrinsic value, negative results often remain unpublished because they are abandoned by authors and/or rejected by editors. Controversial topics like arbovirus VT do not entirely escape publication bias, but are thought to minimize it because negative results are more frequently published (e.g., Vazeille et al., 2009; Watts et al., 1985). By nature, however, publication bias is difficult to quantify. Therefore, caution must be used to interpret the results, keeping in mind that under reporting of negative results leads to overestimating VT.

Although the existence of VT is unequivocal for several mosquito-arbovirus pairs of public health significance, our databases showed that VT is collectively rare. Studies that estimated VT prevalence in a natural situation typically found that <0.1% of mosquitoes are vertically infected, regardless of the virus family or mosquito species. VT efficiency per se, however, cannot be inferred in field studies because infection prevalence in the adult population is unknown. VT prevalence in experimental studies is 10 to 1000-fold higher, which reflects more closely VT efficiency because the parental generation is usually 100% infected. Even when it is relatively efficient, VT is unlikely to explain the persistence of arboviruses in the prolonged absence of horizontal transmission. A recent mathematical model on dengue concluded that VT efficiency would have to be 5- to 30-fold higher than the experimentally measured VT efficiency to support long-term persistence of the virus (Adams and Boots, 2010). It has been argued that TOT alone could maintain the virus indefinitely in a

1 mosquito subpopulation whose germinal tissues are permanently infected (Turell et al., 1982)
2 even if the prevalence in the overall population is low (Tesh, 1984). Unless VT efficiency is
3
4 100%, however, occasional horizontal amplification is predicted to be necessary for the long-
5
6 term maintenance of arboviruses (Fine, 1975).
7
8

9 Our historical perspective highlighted two important aspects of VT research in
10 arbovirology. First, scientific production on arbovirus VT in mosquitoes has been extremely
11 uneven over time. In particular, epidemiological landmarks such as emergence events have
12 significantly stimulated VT research. Second, the evolution of virological assays used in VT
13 studies has profoundly influenced the ability to detect it. Increased sensitivity and higher-
14 throughput of more recent laboratory assays are associated with higher estimates of VT rates.
15
16 Taken together, our results call for caution when interpreting VT studies because their
17 conclusions are context- and method-dependent.
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Acknowledgements

1
2
3
4
5 The authors thank members of the Lambrechts lab for insightful discussions, Richard
6
7 Paul for critical reading of the manuscript, and two anonymous reviewers for useful
8
9 comments. S.L. was supported by a doctoral fellowship from University Pierre and Marie
10
11 Curie. L.L. received funding from the French Government's Investissement d'Avenir program,
12
13 Laboratoire d'Excellence Integrative Biology of Emerging Infectious Diseases (grant ANR-
14
15 10-LABX-62-IBEID).
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

References

- 1
2
3
4 Adams, B., Boots, M., 2010. How important is vertical transmission in mosquitoes for the
5 persistence of dengue? Insights from a mathematical model. *Epidemics* 2, 1–10.
6
7
8
9 Bennett, S.N., Drummond, A.J., Kapan, D.D., Suchard, M.A., Munoz-Jordan, J.L., Pybus,
10 O.G., Holmes, E.C., Gubler, D.J., 2010. Epidemic dynamics revealed in dengue
11 evolution. *Mol. Biol. Evol.* 27, 811–818.
12
13
14
15
16 Bhatt, S., Gething, P.W., Brady, O.J., Messina, J.P., Farlow, A.W., Moyes, C.L., Drake, J.M.,
17 Brownstein, J.S., Hoen, A.G., Sankoh, O., Myers, M.F., George, D.B., Jaenisch, T., Wint,
18 G.R.W., Simmons, C.P., Scott, T.W., Farrar, J.J., Hay, S.I., 2013. The global distribution
19 and burden of dengue. *Nature* 496: 504–507.
20
21
22
23
24
25
26 Bocquet, J.-P., Molero, M., 1996. Une hypothèse originale : la transmission transovarienne
27 des arbovirus de Finlay à nos jours, in: Waast, R., Moulin, A.M. (Eds.). Presented at the
28 Les sciences hors d'Occident au 20ème siècle - Médecines et Santé, Paris, pp. 83–88.
29
30
31
32
33
34 Centers for Disease Control and Prevention, 2010. Arbovirus Catalog [WWW Document].
35 [wwwn.cdc.gov](http://wwwn.cdc.gov/Arbocat/Default.aspx). URL <http://wwwn.cdc.gov/Arbocat/Default.aspx> (accessed 1.20.14).
36
37
38
39 Chamberlain, R.W., Kissling, R.E., Stamm, D.D., Nelson, D.B., Sikes, R.K., 1956a.
40 Venezuelan equine encephalomyelitis in wild birds. *Am. J. Hyg.* 63, 261–273.
41
42
43
44 Chamberlain, R.W., Sikes, R.K., Nelson, D.B., 1956b. Infection of *Mansonia perturbans* and
45 *Psorophora ferox* mosquitoes with Venezuelan equine encephalomyelitis virus. *Proc. Soc.*
46 *Exp. Biol. Med.* 91, 215–216.
47
48
49
50
51 Chamberlain, R.W., Sudia, W.D., 1957. The North American arthropod-borne encephalitis
52 viruses in *Culex tarsalis* Coquillett. *Am. J. Hyg.* 66, 151–159.
53
54
55
56 Chamberlain, R.W., Sudia, W.D., Gillett, J.D., 1959. St. Louis encephalitis virus in
57 mosquitoes. *Am. J. Hyg.* 70, 221–236.
58
59
60
61
62
63
64
65

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
- Davis, N.C., Shannon, R.C., 1930. The location of yellow fever virus in infected mosquitoes and the possibility of hereditary transmission. *Am. J. Epidemiol.* 11, 335–344.
- Diaz, L.A., Flores, F.S., Quaglia, A., Contigiani, M.S., 2012. Intertwined arbovirus transmission activity: reassessing the transmission cycle paradigm. *Front. Physiol.* 3, 493.
- Ebert, D., 2013. The epidemiology and evolution of symbionts with mixed-mode transmission. *Annu. Rev. Ecol. Evol. Syst.* 44, 623–643.
- Eldridge, B.F., 1966. Environmental control of ovarian development in mosquitoes of the *Culex pipiens* complex. *Science* 151, 826–828.
- Fechter-Leggett, E., Nelms, B.M., Barker, C.M., Reisen, W.K., 2012. West Nile virus cluster analysis and vertical transmission in *Culex pipiens* complex mosquitoes in Sacramento and Yolo Counties, California, 2011. *J. Vector Ecol.* 37, 442–449.
- Fine, P.E., 1975. Vectors and vertical transmission: an epidemiologic perspective. *Ann. N. Y. Acad. Sci.* 266, 173–194.
- Finlay, C.J., 1899. Mosquitoes considered as transmitters of yellow fever and malaria. *Med. Record* 55, 737–739.
- Frobisher, M., Jr, Davis, N.C., Shannon, R.C., 1931. On the failure of yellow fever virus to persist in a colony of *Aedes aegypti*. *Am. J. Epidemiol.* 14, 142–146.
- Gubler, D.J., 2001. Human arbovirus infections worldwide. *Ann. N. Y. Acad. Sci.* 951, 13–24.
- Gubler, D.J., 2002. The global emergence/resurgence of arboviral diseases as public health problems. *Arch. Med. Res.* 33, 330–342.
- Guiteras, J., 1901. Experimental yellow fever at the inoculation station of the Sanitary Department of Havana with a view to producing immunization. *Am. Med.* 2, 809–817.
- Guzmán, M.G., Kourí, G., Vázquez, S., Rosario, D., Bravo, J., Valdés, L., 1999. DHF epidemics in Cuba, 1981 and 1997: some interesting observations. *Dengue Bull.* 23, 39–

43.

1
2 Hastie, J.T., Pregibon, D., 1991. Generalized linear models, in: Chambers, J.M., J, H.T.
3
4 (Eds.), Statistical Models in S. Wadsworth & Brooks/Cole.
5

6
7 Hindle, E., 1930. The transmission of yellow fever. *Lancet* 216, 835–842.
8

9
10 Joshi, V., Mourya, D.T., Sharma, R.C., 2002. Persistence of dengue-3 virus through
11
12 transovarial transmission passage in successive generations of *Aedes aegypti* mosquitoes.
13
14 *Am. J. Trop. Med. Hyg.* 67, 158–161.
15

16
17 Kilpatrick, A.M., 2011. Globalization, land use, and the invasion of West Nile virus. *Science*
18
19 334, 323–327.
20

21
22 Kissling, R.E., Chamberlain, R.W., Nelson, D.B., Stamm, D.D., 1956. Venezuelan equine
23
24 encephalomyelitis in horses. *Am. J. Hyg.* 63, 274–287.
25

26
27 Kissling, R.E., Chamberlain, R.W., Sudia, W.D., Stamm, D.D., 1957. Western equine
28
29 encephalitis in wild birds. *Am. J. Hyg.* 66, 48–55.
30

31
32 Lambrechts, L., Scott, T.W., 2009. Mode of transmission and the evolution of arbovirus
33
34 virulence in mosquito vectors. *Proc. Biol. Sci.* 276, 1369–1378.
35

36
37 Leake, C.J., 1984. Transovarial transmission of arboviruses by mosquitoes, in: Mayo, M.A.,
38
39 Harrap, K.A. (Eds.), *Vectors in virus biology*. Academic Press, New-York & London, pp.
40
41 63–91.
42

43
44 Leparc-Goffart, I., Nougairede, A., Cassadou, S., Prat, C., de Lamballerie, X., 2014.
45
46 *Chikungunya in the Americas*. *Lancet* 383, 514.
47

48
49 Littell, J.H., Corcoran, J., Pillai, V., 2008. *Systematic reviews and meta-analysis*. Oxford Univ
50
51 Press, New-York.
52

53
54 Marchoux, É., Simond, P.-L., 1905. La transmission héréditaire du virus de la fièvre jaune
55
56 chez le *Stegomyia fasciata*. *C. R. Soc. Biol. Paris* 259–260.
57

58
59 Marchoux, É., Simond, P.-L., 1906. Etudes sur la fièvre jaune - Deuxième mémoire. *Ann.*
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Inst. Pasteur 20, 1–25.

- Messina, J.P., Brady, O.J., Scott, T.W., Zou, C., Pigott, D.M., Duda, K.A., Bhatt, S., Katzelnick, L., Howes, R.E., Battle, K.E., Simmons, C.P., Hay, S.I., 2014. Global spread of dengue virus types: mapping the 70 year history. *Trends Microbiol.* 22, 138–146.
- Mitamura, T., Kitaoka, M., Watanabe, S., Hosoi, T., Tenjin, S., Seki, O., Nagahata, K., Jo, K., Shimizu, M., 1939. Weitere Untersuchungen über die Übertragung der japanischen epidemischen enzephalitis durch Mücken. *Trans. Soc. Pathol. Jpn.* 29, 92–105.
- Mitchell, C.J.J., Briegel, H., 1989. Fate of the blood meal in force-fed, diapausing *Culex pipiens* (Diptera: Culicidae). *J. Med. Entomol.* 26, 332–341.
- Nelms, B.M., Macedo, P.A., Kothera, L., Savage, H.M., Reisen, W.K., 2013. Overwintering biology of *Culex* (Diptera: Culicidae) mosquitoes in the Sacramento valley of California. *J. Med. Entomol.* 50, 773–790.
- Pialoux, G., Gaüzère, B.-A., Jauréguiberry, S., Strobel, M., 2007. Chikungunya, an epidemic arbovirolosis. *Lancet Infect. Dis.* 7, 319–327.
- Reed, W., Carroll, J., Agramonte, A., 1901. Experimental yellow fever. *Am. Med.* 2, 15–23.
- Reeves, W.C., 2004. Overwintering of arboviruses, in: *Epidemiology and control of mosquito-borne arboviruses in CA, 1943-1987*. Calif. Mosq. Vector Control Assoc., Sacramento, California, pp. 357–382.
- Reisen, W.K., 2013. Ecology of West Nile virus in North America. *Viruses* 5, 2079–2105.
- Rezza, G., Nicoletti, L., Angelini, R., Romi, R., Finarelli, A.C., Panning, M., Cordioli, P., Fortuna, C., Boros, S., Magurano, F., Silvi, G., Angelini, P., Dottori, M., Ciufolini, M.G., Majori, G.C., Cassone, A., CHIKV study group, 2007. Infection with chikungunya virus in Italy: an outbreak in a temperate region. *Lancet* 370, 1840–1846.
- Root, J.J., Oesterle, P.T., Nemeth, N.M., Klenk, K., Gould, D.H., McLean, R.G., Clark, L., Hall, J.S., 2006. Experimental infection of fox squirrels (*Sciurus niger*) with West Nile

virus. *Am. J. Trop. Med. Hyg.* 75, 697–701.

1
2 Root, J.J., Oesterle, P.T., Sullivan, H.J., Hall, J.S., Marlenee, N.L., McLean, R.G.,
3
4 Montenieri, J.A., Clark, L., 2007. Fox squirrel (*Sciurus niger*) associations with West Nile

5
6
7 virus. *Am. J. Trop. Med. Hyg.* 76, 782–784.

8
9
10 Rosen, L., 1988. Further observations on the mechanism of vertical transmission of
11
12 flaviviruses by *Aedes* mosquitoes. *Am. J. Trop. Med. Hyg.* 39, 123–126.

13
14 Rosenau, M.J., Goldberger, J., 1906. Attempts to grow the yellow fever parasite - The
15
16 hereditary transmission of the yellow fever parasite in the mosquito. *Yellow Fever*
17
18 *Institute, Bulletin* 15, 103–115.

19
20
21 Sheather, S.J., Jones, M.C., 1991. A reliable data-based bandwidth selection method for
22
23
24 kernel density estimation. *J. Roy. Stat. Soc. B* 53, 683–690.

25
26 Siler, J.F., Hall, M.W., Hitchens, A.P., 1926. Dengue: its history, epidemiology, mechanism
27
28 of transmission, etiology, clinical manifestations, immunity, and prevention. *Philippine J.*
29
30 *Sci.* 29, 1–322.

31
32
33 Simmons, J.S., St John, J.H., Reynolds, F.H.K., 1931. Experimental studies of dengue.
34
35
36 *Philippine J. Sci.* 44, 1–255.

37
38
39 Tesh, R.B., 1984. Transovarial transmission of arboviruses in their invertebrate vectors, in:
40
41 Harris, K.F. (Ed.), *Current topics in vector research*. New York, pp. 57–76.

42
43
44 Tesh, R.B., Chaniotis, B.N., Johnson, K.M., 1972. Vesicular stomatitis virus (Indiana
45
46 serotype): transovarial transmission by phlebotomine sandflies. *Science* 175, 1477–1479.

47
48
49 Tesh, R.B., Shroyer, D.A., 1980. The mechanism of arbovirus transovarial transmission in
50
51 mosquitoes: San Angelo virus in *Aedes albopictus*. *Am. J. Trop. Med. Hyg.* 29, 1394.

52
53
54 Turell, M.J., 1988. Horizontal and vertical transmission of viruses by insect and tick vectors.,
55
56 in: Monath, T.P. (Ed.), *The arboviruses: epidemiology and ecology*. CRC Press, Boca
57
58 Raton, Florida, pp. 127–152.

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
- Turell, M.J., Hardy, J.L., Reeves, W.C., 1982. Stabilized infection of California encephalitis virus in *Aedes dorsalis*, and its implications for viral maintenance in nature. *Am. J. Trop. Med. Hyg.* 31, 1252–1259.
- Vasilakis, N., Weaver, S.C., 2008. The history and evolution of human dengue emergence. *Adv. Virus Res.* 72, 1–76.
- Vazeille, M., Mousson, L., Failloux, A.-B., 2009. Failure to demonstrate experimental vertical transmission of the epidemic strain of chikungunya virus in *Aedes albopictus* from La Réunion Island, Indian Ocean. *Mem. Inst. Oswaldo Cruz* 104, 632–635.
- Watts, D.M., Harrison, B.A., Pantuwatana, S., Klein, T.A., Burke, D.S., 1985. Failure to detect natural transovarial transmission of dengue viruses by *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae). *J. Med. Entomol.* 22, 261–265.
- Watts, D.M., Pantuwatana, S., DeFoliart, G.R., Yuill, T.M., Thompson, W.H., 1973. Transovarial transmission of LaCrosse virus (California encephalitis group) in the mosquito, *Aedes triseriatus*. *Science* 182, 1140–1141.
- World Health Organization, 2011. West Nile virus [WWW Document]. World Health Organization. URL <http://www.who.int/mediacentre/factsheets/fs354/en/> (accessed 3.8.14).
- Yoon, I.-K., Getis, A., Aldstadt, J., Rothman, A.L., Tannitisupawong, D., Koenraadt, C.J.M., Fansiri, T., Jones, J.W., Morrison, A.C., Jarman, R.G., Nisalak, A., Mammen, M.P., Thammapalo, S., Srikiatkachorn, A., Green, S., Libraty, D.H., Gibbons, R.V., Endy, T., Pimgate, C., Scott, T.W., 2012. Fine scale spatiotemporal clustering of dengue virus transmission in children and *Aedes aegypti* in rural Thai villages. *PLoS Negl. Trop. Dis.* 6, e1730.

Figure Legends

Figure 1. Evolution of the yearly number of publications on arbovirus VT by mosquitoes. Bars show the yearly number of publications in each of five broad categories represented by different colors. The dashed vertical line indicates the publication of an influential *Science* article on LaCrosse virus VT by Watts et al. in 1973.

Figure 2. Evolution of the yearly number of publications on dengue virus VT. Bars represent the normalized number of yearly publications on dengue virus VT. The red curve shows the overall number of dengue publications used for normalization. The vertical dashed line indicates the start of the 1981 DHF epidemic in the Americas.

Figure 3. Evolution of the yearly number of publications on West Nile virus VT. Bars represent the normalized number of yearly publications on West Nile virus VT. The red curve shows the overall number of West Nile virus publications used for normalization. The vertical dashed line indicates the start of the 1999 West Nile virus emergence in North America.

Figure 4. Evolution of the yearly number of publications on chikungunya virus VT. Bars represent the normalized number of yearly publications on chikungunya virus VT. The red curve shows the overall number of chikungunya publications used for normalization. The vertical dashed line indicates the start of the 2005 chikungunya epidemic in the Indian Ocean region.

Figure 5. Evolution of virological assays used for arbovirus VT detection in (A) experimental and (B) natural studies. In each panel, colored areas represent the relative

1
2 likelihood of each assay category at a given time-point on the x -axis. The yearly proportion of
3 each assay is calculated relative to the total number of individual mosquitoes tested for VT.
4
5
6

7
8 **Figure 6. Evolution of VT detection in (A) experimental and (B) natural studies of the**
9 ***Aedes-Flavivirus* pair.** In each panel, the curve represents the relative likelihood of VT
10 occurrence at a given time-point on the x -axis. The yearly detection rate is calculated relative
11 to the total number of database entries.
12
13
14
15
16

17
18
19
20 **Figure 7. Distribution of sample sizes by assay category in (A) experimental and (B)**
21 **natural studies of arbovirus VT.** Sample size was \log_{10} -transformed prior to analysis and
22 graphical representation. Boxplots represent the 75th percentile, median and 25th percentile;
23 whiskers extend to the highest and lowest value in the 1.5x interquartile range.
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Figure 1

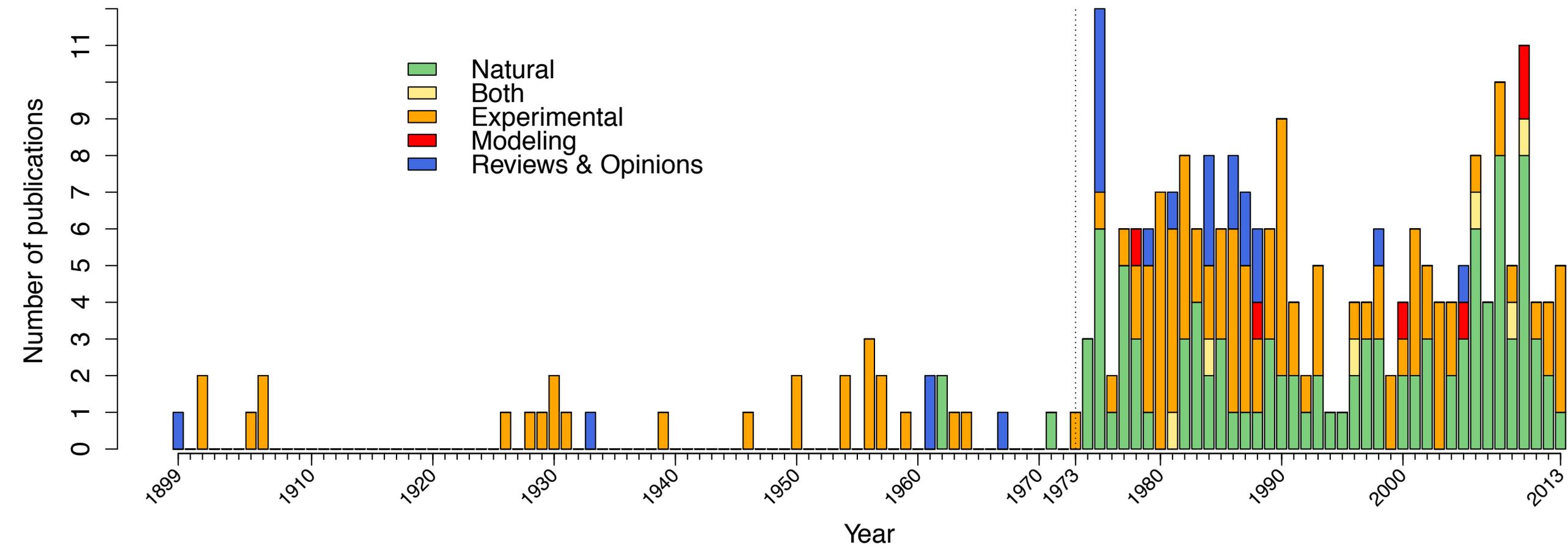


Figure 2

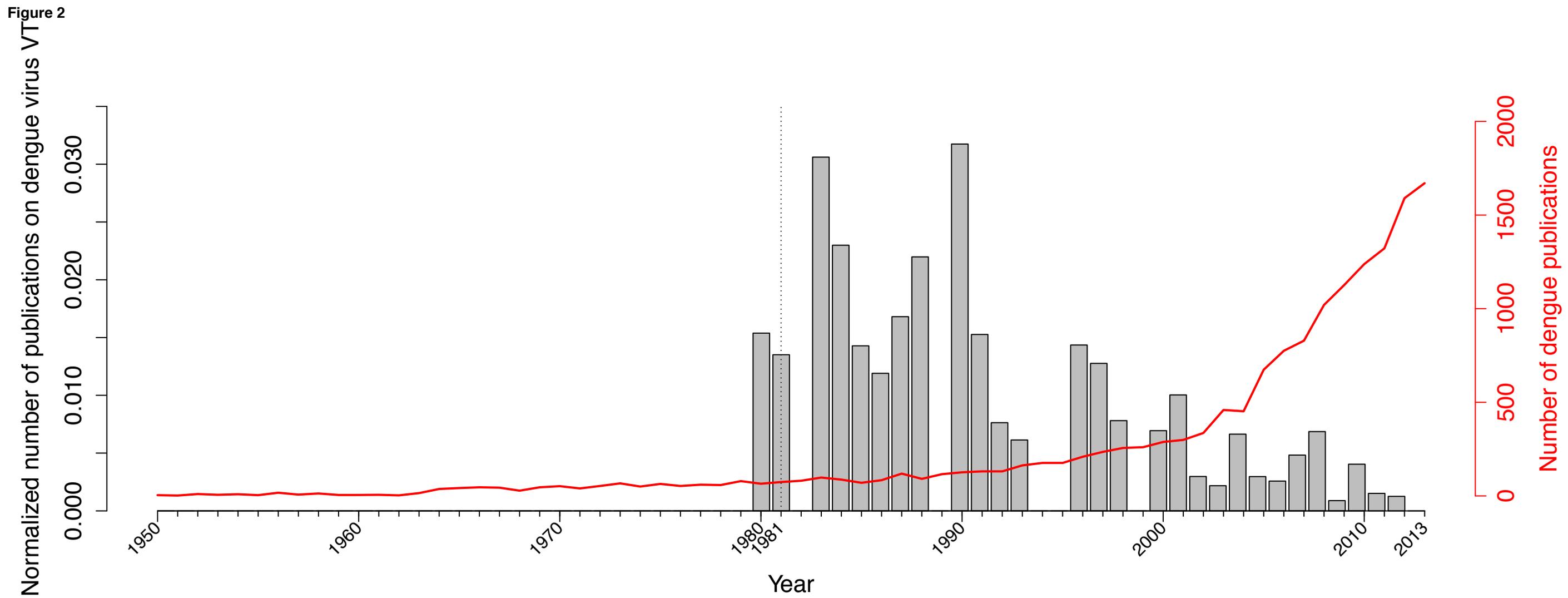


Figure 3

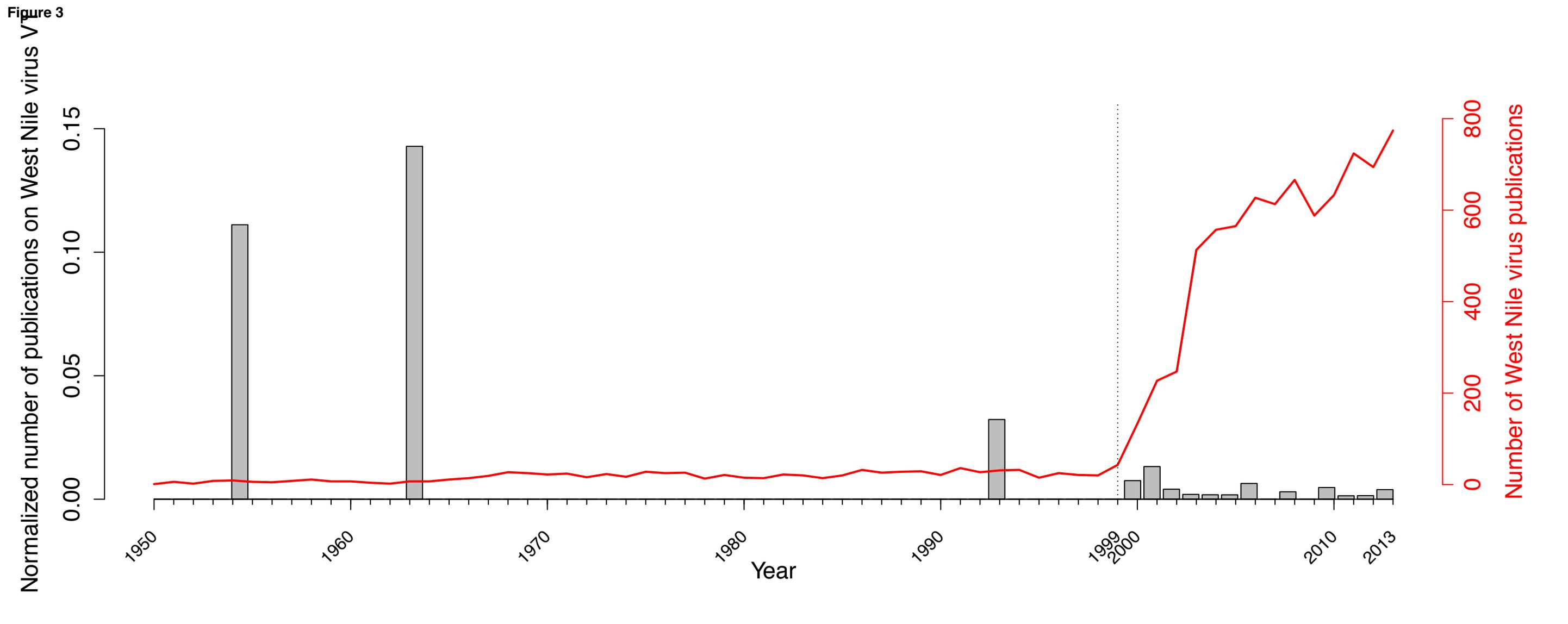


Figure 4

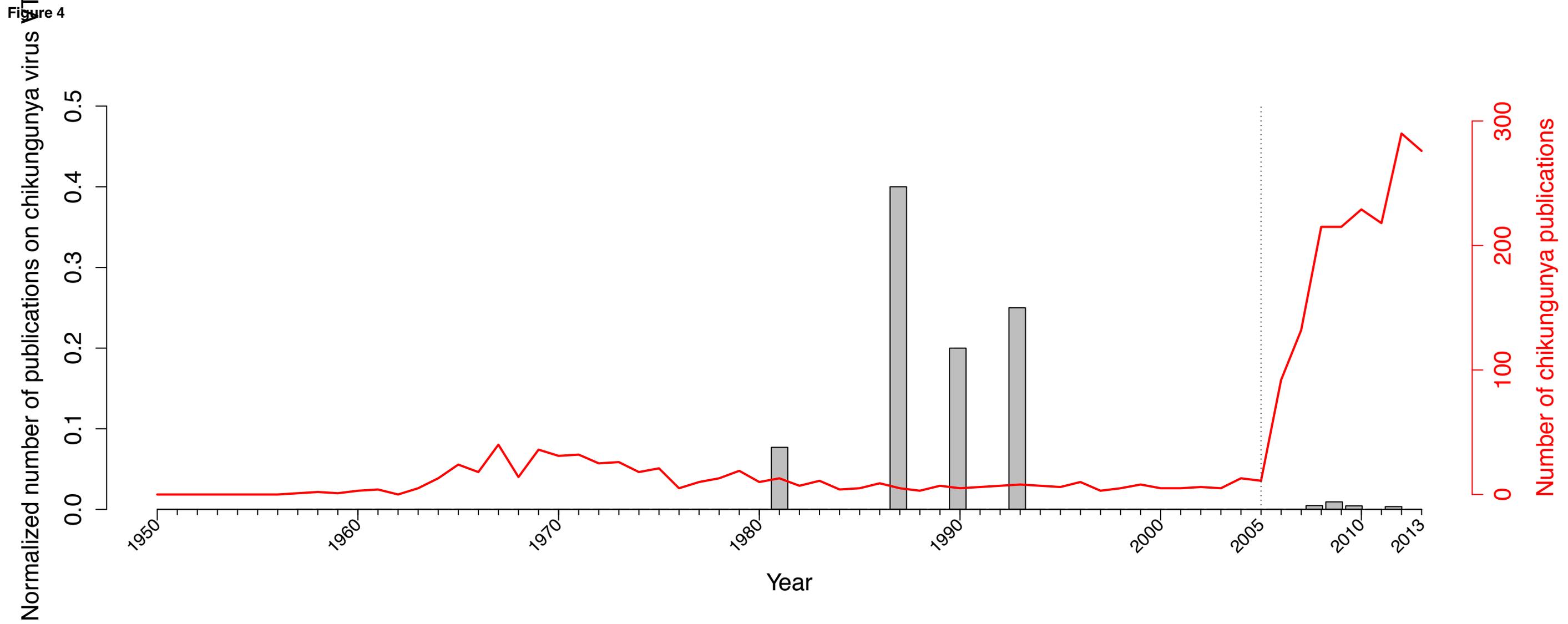
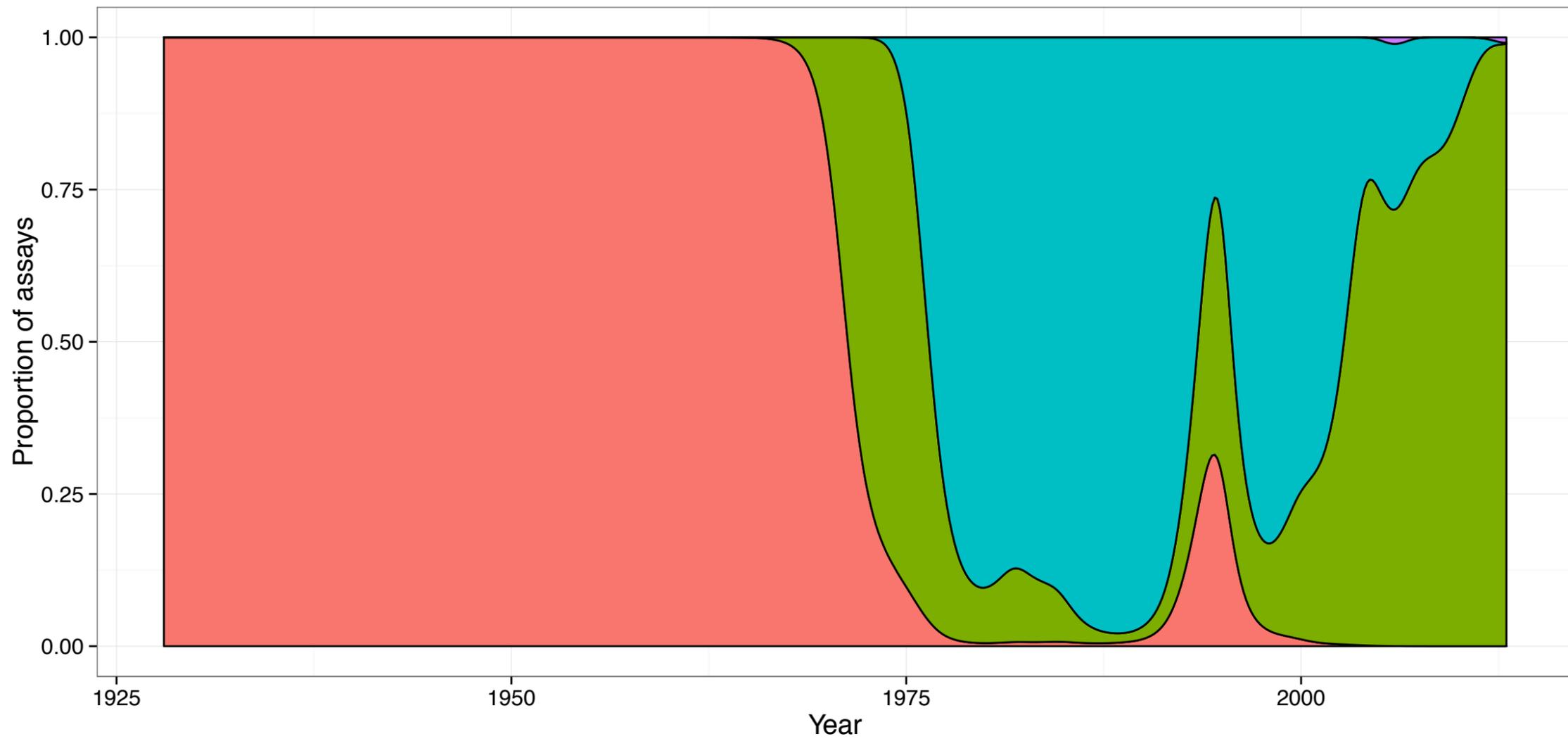


Figure 5

A



B

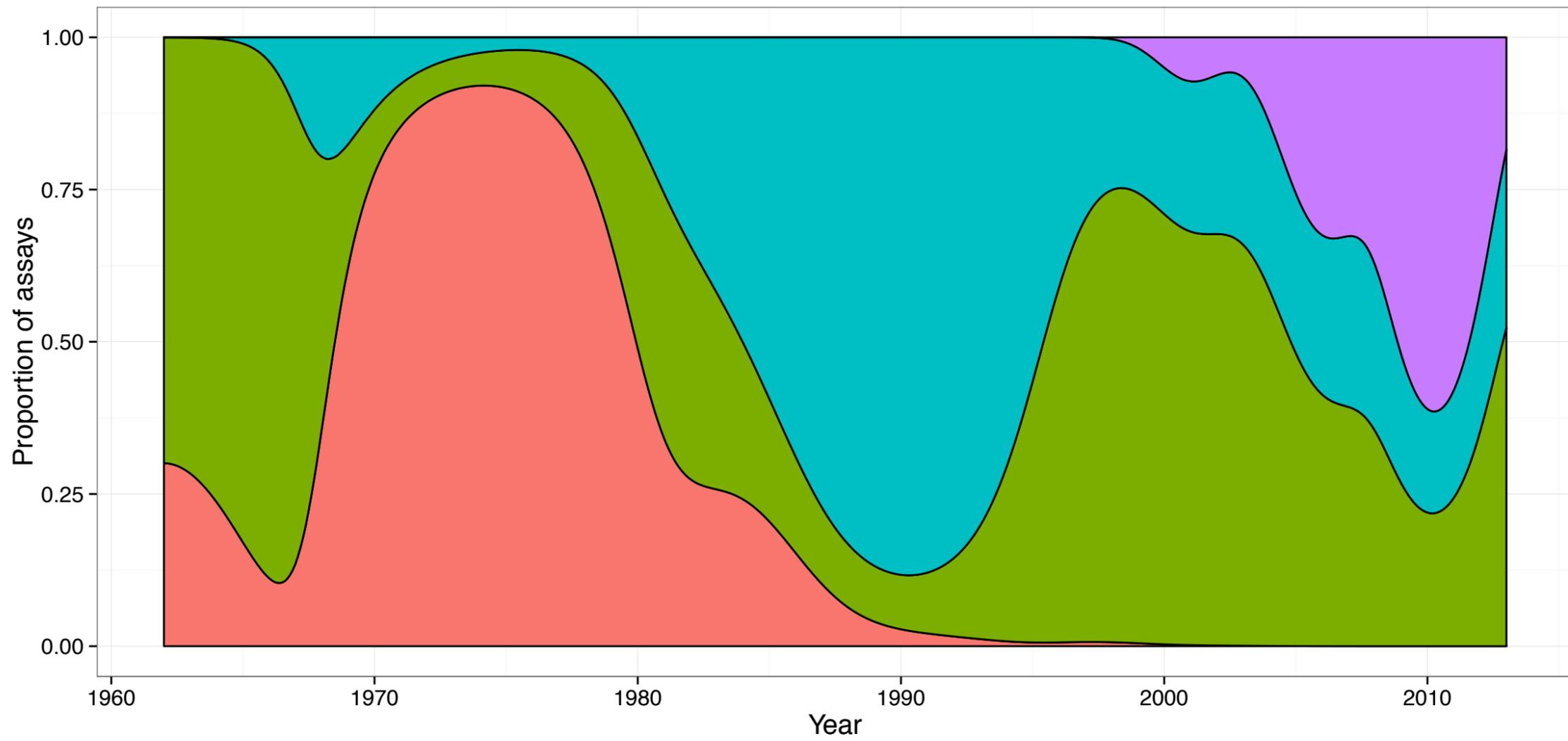


Figure 6

A



B

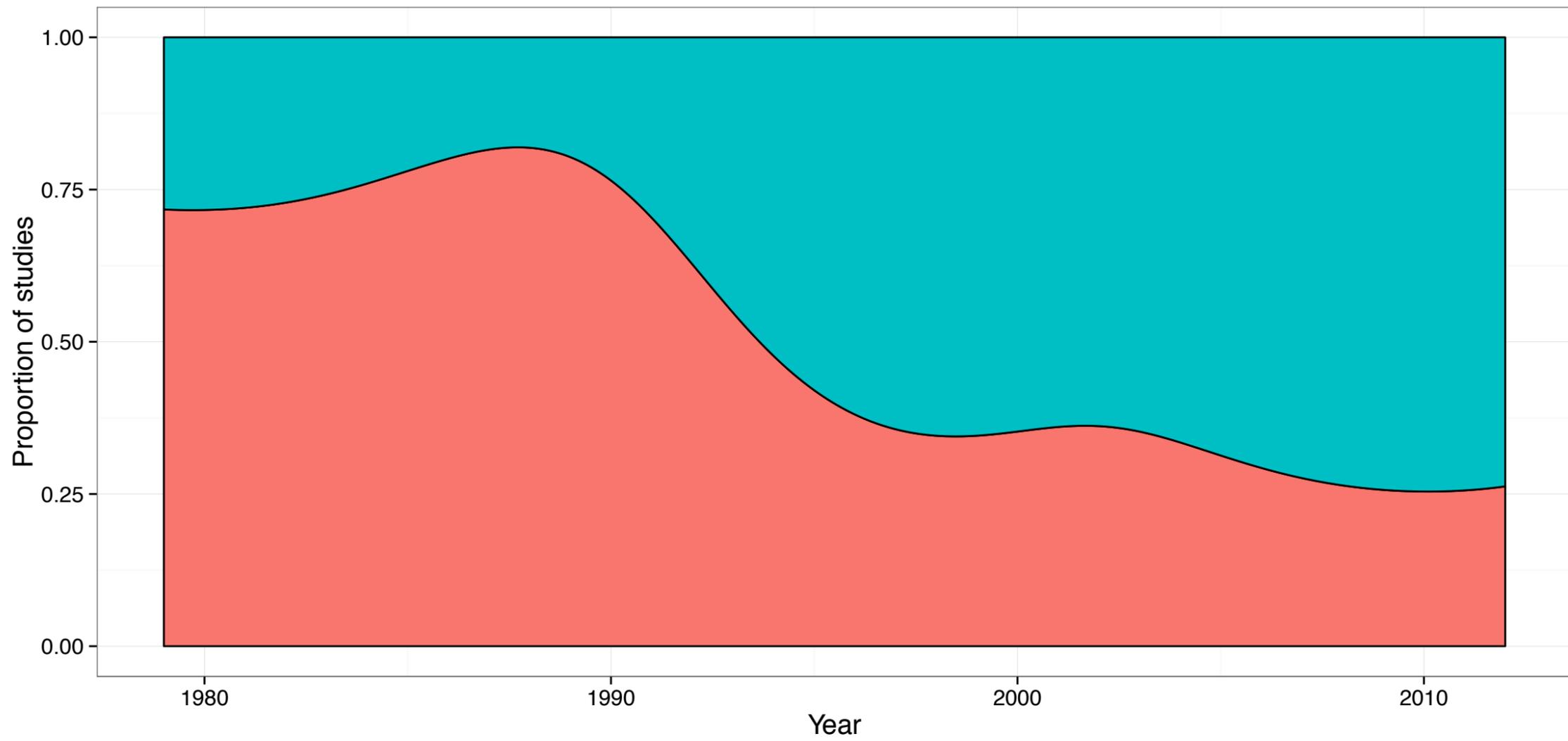
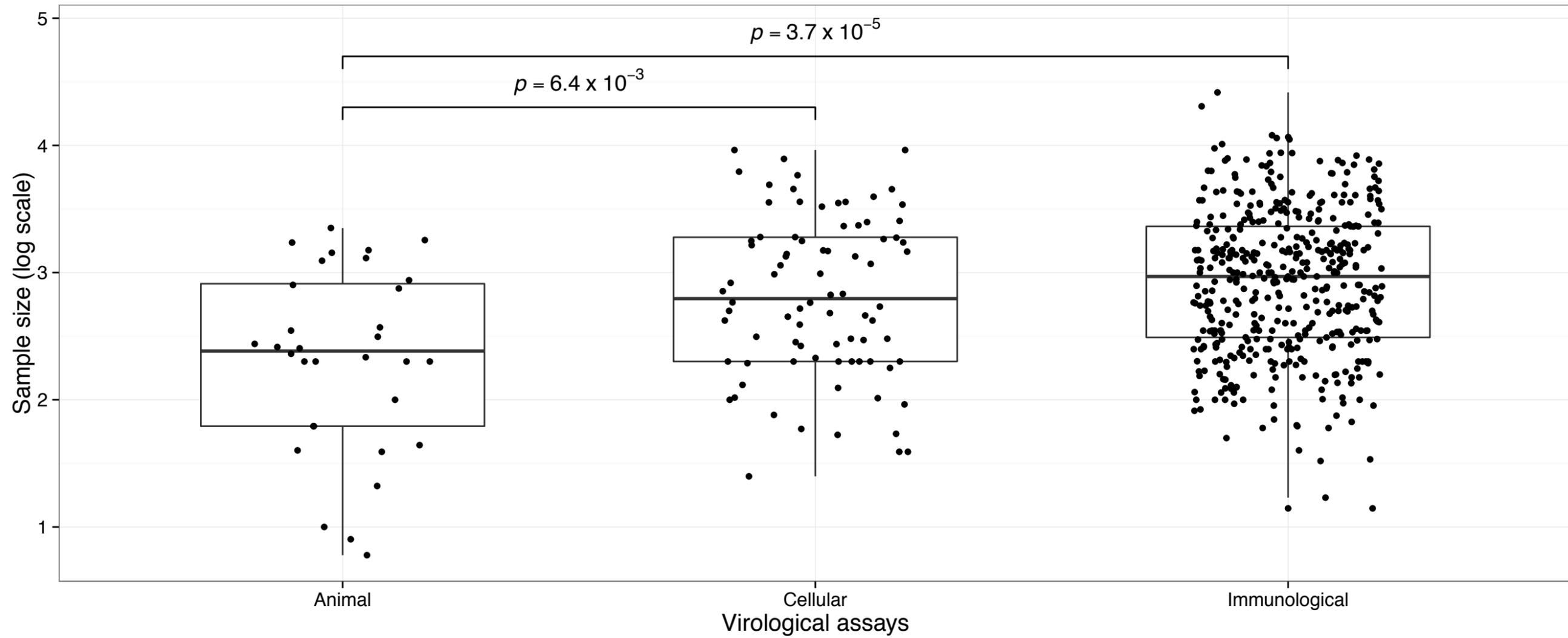


Figure 7

A



B

