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***Non-human primates in HIV research: Achievements, limits and alternatives***

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## ***Abstract***

An ideal model for HIV-1 research is still unavailable. However, infection of non-human primates (NHP), such as macaques, with Simian Immunodeficiency Virus (SIV) recapitulates most virological, immunological and clinical hallmarks of HIV infection in humans. It has become the most suitable model to study the mechanisms of transmission and physiopathology of HIV/AIDS. On the other hand, natural hosts of SIV, such as African green monkeys and sooty mangabeys that when infected do not progress to AIDS, represent an excellent model to elucidate the mechanisms involved in the capacity of controlling inflammation and disease progression. The use of NHP-SIV models has indeed enriched our knowledge in the fields of: i) viral transmission and viral reservoirs, ii) early immune responses, iii) host cell-virus interactions in tissues, iv) AIDS pathogenesis, v) virulence factors, vi) prevention and vii) drug development. The possibility to control many variables during experimental SIV infection, together with the resemblance between SIV and HIV infections, make the NHP model the most appropriate, so far, for HIV/AIDS research. Nonetheless, some limitations in using these models have to be considered. Alternative models for HIV/AIDS research, such as humanized mice and recombinant forms of HIV-SIV viruses (SHIV) for NHP infection, have been developed. The improvement of SHIV viruses that mimic even better the natural history of HIV infection and of humanized mice that develop a greater variety of human immune cell lineages, is ongoing. None of these models is perfect, but they are contributing to the progress in managing or preventing HIV infection.

**Keywords:** HIV, SIV, AIDS, non-human primates, animal models

## **Highlights**

- <sup>35</sup><sub>17</sub>NHP allow studying viral transmission, immunopathology, immune response and prevention.
- <sup>35</sup><sub>17</sub>Natural SIV host models help to reveal mechanisms of HIV disease control.
- <sup>35</sup><sub>17</sub>NHP models of spontaneous or ART-induced control provide insights for HIV cure.
- <sup>35</sup><sub>17</sub>NHP models have allowed major advances in HIV/AIDS field but they are not perfect.
- <sup>35</sup><sub>17</sub>Alternative models (humanized mice, SHIV) are useful for specific research questions.

## INTRODUCTION

HIV/AIDS is still a major public health issue. According to the World Health Organization, HIV infection figures, even today, among the ten major leading causes of death and is the second cause of mortality in adolescents. Since the first report in 1981 and the identification of HIV as a causative agent in 1983<sup>1</sup>, AIDS has claimed more than 35 million of lives and only in 2015, 2.1 million of people became newly infected with HIV. HIV infection is characterized by a slow and progressive loss of CD4<sup>+</sup>T cells that, in absence of treatment, generally leads to an immunosuppressive condition. Nowadays, it is admitted that chronic immune activation is the driving force of such immunodeficiency<sup>2</sup>. Under successful combined antiretroviral therapy (cART), the virus is controlled up to undetectable levels in blood, but a residual chronic inflammation persists and is associated with the morbidity and mortality observed in the antiretroviral-treated patients.

Despite of the great advances obtained in HIV/AIDS knowledge, there are still key problems to solve, in particular the lack of a vaccine and a cure and the absence of treatments for resolution of HIV-induced inflammation. Animal models for HIV have already contributed to answer major questions, but they also have several limitations. The “perfect animal model” for HIV-1 research is indeed still unavailable. However, infection of non-human primates (NHP), such as macaques, with Simian Immunodeficiency Virus (SIV) leads to a disease that is similar to AIDS induced by HIV in humans. So far, this is the most suitable model to study the mechanisms of transmission and physiopathology of the disease. Indeed, SIV infection in macaques fulfills numerous conditions generally requested to constitute a reliable animal model for a human disease:

1. The virus causing disease in the model should cause the same disease in humans;
2. The course of the disease in the animals should resemble that in humans;
3. The range of cells, tissues and organs involved should be similar in humans and in the animal;
4. Immune responses to infection in the animal model should be similar to those in humans.

These conditions are not fulfilled by other animal models, such as FIV in cats or HIV-1 in humanized mice (see below). Here, we review characteristics of the SIV infection in NHP that have favored its use as a model for HIV/AIDS research and summarize some of the major past and recent advances in the field obtained thanks to the NHP models (Table 1). We will briefly evoke advances in other models, such as humanized mice, in research toward a HIV vaccine and cure. Finally, we will explain the limits of NHP models and discuss how these models could nonetheless help in the global effort to achieve the development of efficient preventive and cure approaches.

## 1. SIV MODELS AND THEIR CONTRIBUTIONS TO RESEARCH ON HIV PREVENTION AND TREATMENT

Human AIDS is caused by two types of HIV (HIV-1 and HIV-2). These viruses are subdivided into groups (M, N, O, and P for HIV-1; A to I for HIV-2), subtypes, circulating recombinant forms (CRF) and unique recombinant forms (URF), HIV-1 group M subtype C being the most prevalent in the world<sup>3,4</sup>.

Since the first cases of HIV infection, there has been interest in identifying the origin of the epidemic. The first insight came to light when a close relationship between HIV-2 and a virus that infects macaques was found<sup>5</sup>. This virus was called SIVmac in analogy to HIV and according to the species from which it was isolated. Thereafter, another phylogenetic association was discovered between HIV-2 and SIVsmm, a virus that infects sooty mangabeys in West Africa<sup>6</sup>. Subsequent studies confirmed that SIVmac and HIV-2 derived both from SIVsmm<sup>6-9</sup>.

The origin of HIV-1 was traced on non-human primates, as well. The human virus is closely related to SIVcpz, which infects West-Central Africa chimpanzees (*Pan troglodytes troglodytes*)<sup>7,8,10-12</sup> and from which HIV-1 M and N derived<sup>11-16</sup>. On the other hand, the analysis of fecal samples from Cameroonian gorillas revealed the existence of SIVgor<sup>17,18</sup>. The latter virus is related to HIV-1 O and P. Phylogenetic analyses indicate that chimpanzees constitute most likely the original reservoir and source of SIVgor as well as of HIV-1 O and P<sup>18-20</sup> (Figure 1).

More than 40 NHP species have been found to carry SIV in the wild. Noteworthy, natural carriers of SIV are all African species. In contrast, Asian monkeys, such as macaques, are

only infected in captivity.

The first report of AIDS in a NHP was provided by Letvin and colleagues in 1983<sup>21</sup>, soon after the discovery of HIV. This syndrome was detected in captive macaques (*Macaca cyclopis* and *Macaca mulatta*) that died of lymphomas or opportunistic infections like *Pneumocystis carinii*. The revision of autopsy records and laboratory studies revealed that these animals suffered of anemia, neutropenia and monocytosis before death. A lymphocyte ratio (CD4/CD8) reversion and also of T cell numbers and functionality were observed as well before death. The causes of death included necrotizing gingivitis, *Pneumocystis carinii* and cytomegalovirus infections, as well as three atypical cases of lymphoma.

Macaques infected by SIVmac became the most important animal model for HIV/AIDS research. The disease progression profile during SIV infections in macaques depends both on the macaque species and the SIVmac strain used (Figure 2). The most frequently used animals are Indian and Chinese rhesus macaques as well as cynomolgus macaques infected by the SIVmac239 molecular clone or the SIVmac251 viral isolate. The most rapid disease progression is generally observed in Indian rhesus macaques infected with SIVmac239.

This macaque model revealed the massive CD4<sup>+</sup>T cell depletion in the gut in the very early phase of infection<sup>22-24</sup>. While it was already known that HIV replicates in the gut<sup>25</sup>, the macaque model helped to underscore the rapidity of the events in this tissue (within the first 24 h upon infection) and to what extent the degree of T lymphocyte loss in the gut is associated with disease progression. It was subsequently shown that resting memory CD4<sup>+</sup> T cells were the most frequently infected cells within the gastrointestinal (GI) tract<sup>26,27</sup>. These cells can live for decades and are now considered as a major reservoir for HIV-1<sup>28,29</sup>. Experimental SIV infections can be performed through the intravenous route. Several additional experimental protocols have been developed for infection through the rectal, vaginal or oral route in order to mimic the predominant routes of transmission encountered in humans, i.e. sexual or mother-to-child transmission by breast feeding<sup>20,22-24</sup>. To better resemble even more what is happening during infection and dissemination of the virus throughout the body in humans, protocols for repetitive low-dose challenges have been set up as well. These models contribute to evaluate vaccine candidates and already lead to the demonstration of two constructions that can confer strong control, i.e. SIVΔnef and rhesus cytomegalovirus (CMV)-based vectors. However, the degree of protection correlates

inversely with the level of attenuation, the least-attenuated strain giving the greatest protection<sup>33,34</sup>.

Vaccine candidates based on CMV-vectors generate every strong and persistent T effector memory responses in half of the animals leading to a controlled infection and elimination of viral reservoirs<sup>35</sup>. While it is at the moment unclear if such constructions can be used as a vaccine, the studies of these vaccine candidates have

already and will continue to provide important clues about the correlates of protection against HIV<sup>36,37</sup>.

The macaque/SIVmac models also allow to examine the very first immunological events after viral exposure in relation to the transmission route or to study the selection mechanisms of transmitted/founder virus resulting from the genetic bottleneck that occurs during transmission (Table 1)<sup>38-42</sup>. It was shown for instance in the macaque model, that while in most cases the transmission is based on the selection and persistence of only one viral variant from the donor, the number of transmitted founder variants increases with the viral dose in the challenge<sup>31,43-45</sup>.

In the search for a vaccine against HIV-1, there is a need for an animal model that can be infected with the human virus. Such a model would help, for instance, in the study of neutralizing antibodies (Nabs) against HIV-1. Macaques are susceptible to HIV-2 but not to HIV-1 infection. Chimpanzees constitute the original reservoir of HIV-1 and therefore are naturally susceptible to HIV-1 viruses. However, HIV-1 had adapted to the new human host, most likely by circumventing restriction factors, among other reasons. Therefore, the results obtained from HIV-1 infection in these primates have been non conclusive. Some groups have reported the development of AIDS in this model,<sup>46-49</sup> characterized by marked depletion of CD4+T cells, sustained viremia, severe CD4:CD8 inversion and increased T cell apoptosis. Others have observed animals that do not or only slowly progress to AIDS, maintaining their normal CD4+T cell counts and displaying undetectable viremia<sup>50</sup>. The scientific limitations of the model, together with the new guidelines for the use of chimpanzees in biomedical research, led to the interruption of its use for HIV<sup>51</sup>. Infection of macaques with HIV-1 has also revealed to be challenging. Indeed, HIV-1 faces in macaques the existence of cellular proteins that restrict the replication of the human virus, such as TRIM5 $\square$ <sup>52</sup>, APOBEC3G and Tetherin<sup>53</sup>. This makes it difficult to infect macaques or their CD4 $^{+}$ T cells by HIV-1 *invivo* and *in vitro*, but allowed to contribute to

the discovery and extensive characterization of restriction factors (Table 1). Several TRIM5 $\alpha$  alleles have been identified and characterized in monkeys. It has been clearly shown that the viral replication differs depending on the TRIM5 $\alpha$  allele present in the simian host<sup>54,55</sup>. The only macaques susceptible to be infected by HIV-1 is the pig-tailed macaque (*Macaca nemestrina*). This occurs as a result of a 2-nucleotide deletion in the 5'-end of TRIM5 $\alpha$  transcript, which impairs the action of this RF<sup>16</sup>. However, the viral replication is not efficient and there is no disease progression.

A recombinant virus has been engineered, which consists of a HIV-1 backbone where target sites of restriction factors were replaced by the corresponding SIVmac sequences<sup>56</sup>. While this virus induces persistent viremia, disease progression is only observed after *in vivo* depletion of CD8+ T cells in acute infection. This modified model allows the study of some virus-host interactions, but needs further development for its application in HIV research.

Another attempt to circumvent the lack of a macaque model for HIV-1 consists in the construction of other types of recombinant viruses of HIV and SIV, called SHIV<sup>57</sup>. These have been used since many years to overcome the lack of an animal model by constructing for instance SIVmac viruses coding for HIV-1 Env instead of SIVmacEnv. Such macaque/SHIV models were essential for providing the proof of concept that antibodies alone can protect against infection by taking advantage of the possibility to passively transfer the antibodies and challenge the animals experimentally<sup>58</sup> (Table 1). However, these types of approaches are challenged by the large diversity of the envelope glycoprotein, their glycans shields and by the high plasma titers of NAbs needed for protection. Lately, the improvement of broadly neutralizing monoclonal antibodies (bNAbs) allowed to enhance their neutralization potency and to protect, for instance, SHIV-challenged macaques with much lower NAb concentrations<sup>59</sup>. This new generation of very broad and potent Abs might have a potential not only as pre-exposure blocking agents but also in view of a cure or remission of HIV. The treatment with those Abs resulted indeed in control of viremia and reduced proviral DNA in blood and tissues<sup>60,61</sup>.

Very early on, SIVs that encode for HIV-1 Pol have also been engineered. Indeed, SIVmac viruses are not susceptible to some drugs used against HIV-1, such as non-nucleoside

reversetranscriptaseinhibitors (NNRTIs), fusion inhibitors and someintegraseinhibitors. The engineeringof SHIVsexpressingthe HIV-1 Pol protein makes them susceptible to such antiretrovirals.Theycontributed to thedevelopment oftherapeutic approaches.As anexample, NHPinfection models havedemonstrated the effectiveness of subcutaneous,gel, and oral formulations oftenofovir (PMPA)in preventingtransmission ofSIVor SHIV

viruses, even when applied several hours after viral challenge<sup>62</sup>. Such *in vivo*activityof tenofovir madeitapromisingagentforpreventionof HIV-1 infection.

Most SHIV viruses, however, do not reproduce all characteristics ofHIV-1infection in humans. The firstgeneration of SHIVwas composed of X4 virusesand thereforedisplayed a distinct cellular tropismthan primaryHIVin earlyinfection.CD8+T celldepletion in monkeys isstilloften needed fordisease progressionto AIDS.Nevertheless, SHIVmodels have allowed overcomingspecific hurdlesand cannot be missedso farforpre-clinical trials ofvaccine and drug candidates<sup>63-67</sup>.

As wehave recapitulated, SIV infection in NHP constitutes agood model,in manyaspects, forHIVresearch.It is, however, not perfect. Onecannot ignorethat SIVand HIV are close but different viruses,which can lead to differencesin certain aspects oftheir interactions with thehost or susceptibilityto drugsand vaccinecandidates.

## 2. MODELSFORSPONTANEOUSSIV CONTROL

The majorityof theindividualsinfected byHIVprogresses to AIDSwithin7 to 10yearsin the absenceof cART. There exist, however,afewindividuals (<0.2%)known as HIV- controllers who spontaneouslyandefficientlycontrol viral replication. Thedep deep characterization oftheseindividuals and the factors that areinvolved in thecontrol of the infection havebeen thefocus of manyresearches<sup>68,69</sup>. Multiple causes aresuspected, and arenot mutuallyexclusive. In particular thegenetic backgroundof theindividuals enriched in MHCalleles such asB27 and/or B57, strongCD8+T cell responses, reduced susceptibilityof CD4+Tcells to HIV infectionandearlycontrol ofviral replicationare likelyinvolved<sup>70,71</sup>. All distinctprofiles of diseaseprogression in humans are recapitulated bymacaquesafter SIVmacinfection, includingtheoccurrenceofa spontaneously controlled infection in aminorityof macaques thatcan also be related to aspecific MHC background<sup>51,97</sup>. Thediseaseprogressionratedepends both on the virus and host species

(Figure2), as well as on individual host features, as in humans. Studies in the macaque

model allow deciphering, one by one, the role of each viral gene *in vivo* by infecting macaques with genetically engineered SIVmac clones containing defective or lacked accessory genes. This is how it was discovered that Nef is essential for high viral replication *in vivo*, while *in vitro*, Nef was dispensable<sup>72</sup>. Mutations in Nef indeed are associated with attenuated HIV-1 infection in humans.

The role of MHC alleles (*Mamu* alleles in rhesus macaque) has been extensively studied in the spontaneous control in macaques. Some *Mamu* alleles (e.g. *Mamu-A\*01*)<sup>73</sup> are associated with a better containment of viral replication while others are linked to a faster disease progression<sup>74</sup>. Furthermore, the combination of *Mamu* alleles and KIR receptors in NK cells increases the complexity of the response<sup>75,76</sup>, as showed for example in an epistatic analysis of KIR3DL05, KIR3DS05, and KIR3DL10 in association with *Mamu-B\*012* that contributes to elevate the viral load in rhesus macaques<sup>76</sup>.

The NHP models allow investigators to address not only the very early events that follow SIV exposure, but also to study these events in tissues, such as lymphoid tissues, genital tract, lung, liver, intestine, and central nervous system (CNS)<sup>77</sup>. Latent reservoirs of HIV are located throughout the body and persist during infection even in presence highly efficient cART. NHP models allow to study the establishment of these reservoirs in tissues, which occurs in the first hours and days following infection<sup>78-80</sup>. Advances in our understanding of the role of the different immune cell populations as viral target cells, have been provided by experiments in which these populations are depleted in NHP models. Depletion of CD4<sup>+</sup>T-cells in macaques prior to SIV infection was associated with higher viral load, massive activation and infection of macrophages and microglia that become the predominant infected cells<sup>81</sup>. The SIVmac-infected macaque model was used as a proof-of-concept to demonstrate that CD8+ cells participate in the control of infection, using *in vivo* CD8+T cell depletion (Table 1)<sup>82-84</sup>. Additionally, this model allowed to demonstrate the presence of a CTL response as soon as 7 days p.i., and helped to understand how quickly HIV/SIV evades this cellular response through the acquisition of mutations very early on in acute infection<sup>85</sup>. The macaque model also provided the concept that CD8+T cell responses arrive “too little” and “too late” into vaginal mucosa to control viral dissemination and that the “window of opportunity” to prevent infection is very short<sup>86,87</sup>. Furthermore, depletion of NK cells resulted in modest changes in plasma viral load but in a

significant increase in the gut<sup>88,89</sup>. However, no tools exist so far to specifically deplete NK cells and CD8+T cells. Noteworthy, cell depletion induces homeostatic proliferations of other cells, this side effect in addition to others need to be taken into consideration in the interpretation of the results.

### 3. MODELS FOR DISEASE CONTROL

About 40 distinct African NHP are natural hosts of SIV. Only in a few species though, studies could be conducted to evaluate the natural history of SIV infection and its outcome in these animals. Studies based on non-invasive techniques have shown that SIVcpz infection in wild chimpanzees reduces their lifespan, when compared to non-infected animals in the same habitat<sup>16,90</sup>. One case of AIDS-related symptoms was also reported in a naturally infected Central African chimpanzee, which presented lymphopenia, weight loss and opportunistic infections 7 years after SIV positive screening. Efforts should be made to better characterize the pathogenicity of SIV in natural hosts and to find out whether SIV infection also plays a role in ape population decline<sup>91</sup>. Other studies in semi-free and drills infected by SIV and in their natural habitat revealed, in contrast, a strong protection against AIDS. Similarly, other African NHP, such as African Green Monkeys (AGM) and Sooty mangabeys (SM) revealed to be resistant to AIDS<sup>92</sup>. The latter serve today as a model to study the control of the disease, in particular SM and AGM, and are called the “natural hosts” models. Cross-species transmissions demonstrated that the same SIV strain can cause opposite outcomes of infection, depending on the host species (Figure 3). On the other hand, not all SIV strains induce AIDS in macaques, some SIV viruses being easily controlled in the animals, while others not. This demonstrates that the outcome of the infection depends on a combination of both viral and host determinants.

Natural hosts of SIV have been used to better understand the mechanisms responsible for disease progression in HIV infection<sup>92,93</sup>. The most striking difference between non-pathogenic SIV and pathogenic HIV-1/SIVmac infections is the lack of chronic immune activation. Paradoxically, natural hosts of SIV exhibit high titers of virus in plasma, but they do not display chronic activation of the adaptive and innate immune system<sup>94,95</sup>. This feature resembles the ‘viremic non-progressors’, very rare human individuals who display elevated viremia but maintain CD4 T-cell counts and avoid disease progression for

years<sup>96</sup>. Extensive studies are ongoing in order to elucidate the mechanisms involved in the capacity for controlling inflammation and disease progression. Some non-mutually exclusive hypotheses are listed below<sup>97</sup>:

<sup>35</sup><sub>17</sub> Intact mucosa barrier, thereby, lack of microbial translocation and lack of microbial products that would drive the continuous stimulation of the immune system<sup>98</sup>. Experimentally mimicking microbial translocation in these non-pathogenic models provided evidence of the link between this phenomenon and immune activation. LPS administration in AGMs led indeed to elevated T cell activation levels<sup>99</sup>. However, it is unlikely that this is the only event involved in the deleterious chronic inflammation as it has been shown that infection of macaques with a mutated virus still leads to AIDS without acute loss of CD4+ T cells and microbial translocation<sup>100</sup>.

<sup>35</sup><sub>17</sub> Ancient infection that led to an adaptation of the host's immune response responsible for differences in the intensity or nature of SIV sensing. The capacity to produce IFN- $\beta$  is normal in natural hosts, but other sensing pathways have only poorly been explored in natural hosts so far<sup>101,102</sup>.

<sup>35</sup><sub>17</sub> The ability of the Nef protein to block the activation of infected T cells by downregulation of CD3<sup>103</sup>.

<sup>35</sup><sub>17</sub> Natural hosts show low levels of CCR5 surface expression on CD4+ T cells and apparent protection of central memory T cells from SIV infection<sup>104–106</sup>.

<sup>35</sup><sub>17</sub> Controlled viral replication in secondary lymphoid organs<sup>107</sup>. One hallmark of SIV infection in natural hosts is the low viral burden in lymph nodes in the chronic phase of infection in contrast to other tissues<sup>108–112</sup>. This constitutes one of the major differences in lymph nodes of AGMs and SMs. Furthermore, lymph nodes in these natural hosts are characterized by: (i) lack of lymphadenopathy and thus no extensive sequestration of lymphocytes in AGMLNs, (ii) no or less follicular dendritic cell deposition of virus in AGM and SM compared to SIVmac infections<sup>110,112,113</sup>; (iii) more rapid control of interferon-stimulated genes (ISGs) in LN as compared to blood and gut<sup>101,109–111</sup>. The control of viral replication and immune activation in lymph nodes prevents fibrosis and maintains normal immune

function<sup>108,114,115</sup>in this organ which is essential for theinduction and shapingof adaptiveimmune responses.Maintenanceof normal immune functions mightlead to a better control of other infections in the host (i.e. CMV, virome)that otherwise could alsocontributeto immune activation.

<sup>35</sup><sub>17</sub>Depletion of CD4+T cells through apoptosis leadingto homeostaticproliferation of T cells<sup>116,117</sup>

#### 4. SMALLANIMALMODELSFOR HIV RESEARCH

Besides NHPs, other animalmodels havebeen usedforthe studyof HIV-1.In the past,felineimmunodeficiencyvirus (FIV) infection in cats servedas a model ofnaturallyoccurringimmunodeficiency.DuringFIV infection, CD4+T cells aredepleted and a chronic inflammatorystate is established,such asithappens during HIV/SIVinfection<sup>118</sup>. TheFIVmodel has alsobeenuseful in thepastto test some antiretroviral drugs<sup>119</sup>. Nowadays, however, thismodelisgenerallynot usedanymoreforHIVresearch since

some importantfeatures of theinfection areclearlydifferent.Forexample, FIV is able to infect CD8+T cells andBcells in addition to CD4+T cells andmacrophages, dueto its affinityforCD134 asareceptor. Thus,FIVestablishes different viral reservoirs in comparison with thehuman orprimate infection byHIV/SIV.

Other attempts forstudyingHIV-1 infection in small animals such asrats,miceor rabbits wereperformed withoutsuccess.Indeed, rodent cells arerefractoryto HIVinfection. This could not be circumvented bytheexpression ofthe human viralreceptor on the cellsurface. When the murine cells wereengineered forallowing HIV to enter, thevirus encountered additional problems to effectivelyreplicate,mostlikelydueto restriction factors, among others.

Thedevelopment of humanized mice arose thepossibilitytoinfect them with HIV-1 and answer some specificresearch questions. Examples of humanized mice used inHIV research comprise, *interalia*, SCID-hu, SCID-hu-PBL,NOD-SCID, Hu-HSCand BLT mice<sup>67,68,120–122</sup>. Humanized micehavebeen used to studypunctual aspectsofHIV-1 infection, such as Tcell exhaustion<sup>123</sup>, mucosal immunity<sup>120,124,125</sup>, viral latency, antiretroviral drug efficacies and infection ofmacrophages<sup>124,126,127</sup>, amongothers(Table

2). Humanized mice have already contributed to the knowledge of specific aspects of HIV infection. However, their development requires complex surgical engineering. Moreover, they need to be generated *de novo* for each experiment, being tailored according to the immunological requirements of the research, which increases the cost of their establishment and maintenance. In addition, there exist anatomical differences, for example, at the level of lymphoid organs that are not fully developed and lack of several human immune cell lineages, in particular related to innate immune responses. Of note, the field is evolving fast and much effort is put on the improvement of humanized mouse models<sup>121</sup>.

## 5. LIMITATIONS, PERSPECTIVES AND CONCLUSIONS

Despite the numerous advantages that NHP represent, it is important not to forget that choosing the right model for each research question is an essential point. Macaque models of transmission were established very early on as models for HIV<sup>128</sup>. Animal models offer the possibility to control the viral strain, viral dose and infection route. However, while it is well admitted that rectal SIV infection mimics what is happening in humans, vaginal infections are more difficult to modelize as menstrual cycle variations impact the thickness of the epithelium and the efficacy of infection<sup>32,129</sup>. Progesterone is used to control the cycle but to what extent this impacts the relevance of the studies is still under debate.

Also, humans are infected through biological fluids that do not only contain free virus, but also infected cells and other components, such as inflammatory cytokines. Studies are ongoing to evaluate if there are differences depending on the composition of the inoculum<sup>130,131</sup>.

Natural transmission of SIV in African NHPs is predominantly horizontal and thought to occur through sexual contacts or bite wounds. In contrast to HIV infections, vertical transmission of SIV is extremely rare. The expression of the co-receptor CCR5 on CD4<sup>+</sup>T cells in AGMs is low in infants and dramatically increases in adults<sup>132</sup>. Susceptibility to infection through the vaginal and rectal routes was linked to the availability of memory CCR5<sup>+</sup>CD4<sup>+</sup>T cells in the targeted mucosa and it has been suggested that the low rate of vertical SIV infection is associated with the few target cell availability in newborns and infants<sup>133,134</sup>. Thus, while HIV and SIV infections are distinct in the frequency of mother-to-

child transmission, NHPmodels can be used to better understand the underlying mechanism of this difference<sup>133,135,136</sup>.

Once infection takes place at mucosal sites, the virus crosses the epithelial barrier, establishes a first pool of infected cells (foci) and disseminates thereafter in draining lymph nodes (LN<sub>s</sub>)<sup>30</sup>. It is still under debate which cells are the first targets of the virus. It has been suggested that CD4+CCR5+T cells are the primary targets of infection in mucosa<sup>24,26,27,137,138</sup>. Dendritic cells (DCs) might also play a role in the process of viral dissemination. The use of macaque models permitted to analyze their role in virus dissemination during the first hours and days *in vivo*<sup>139</sup>. Nonetheless, the events leading to the establishment of infection are still not fully elucidated.

Repeated low-dose viral challenges might provide a benefit for the evaluation of vaccine candidates or the understanding of the establishment of viral reservoirs. The low number of animals for ethical and economic reasons renders interpretation of the results however sometimes difficult. Taken this into account, technical improvements are being made. For instance, novel techniques, such as *in vivo* imaging, are highly promising<sup>77</sup>. It is important to mention that not only the number of specimens allowed for each research is often limited, but the tools are limited as well. Thus, there are still markers for which no monoclonal antibody exists in monkeys. This is the case for example for most of the KIRs. NHP are the closest animal model for HIV/AIDS research, but an ideal model for HIV-1 is still not available. SIV and HIV are close but distinct viruses, which cause differences in certain aspects of their interactions with the host, vaccines and drugs. To overcome these potential discrepancies between SIV and HIV, recombinant viruses, including SHIVs, have been generated. SHIVs have been and still are particularly useful in the study of humoral responses, vaccine candidates and antiretroviral drugs<sup>67</sup>. In the future, it would be useful however to dispose of SHIV viruses that mimic even better the natural history of HIV infection. The development of new generations of SHIV strains is already ongoing. In conclusion, non-human primates infected by SIV represent a model with numerous advantages for the study of HIV infection. Its close resemblance to HIV infection in humans probably makes it the best or at least the most deeply characterized animal model for a human disease. NHP-SIV models have allowed the study of: i) viral transmission, ii)

early immune responses, iii) host cell-virus interactions in tissues, iv) prevention and v) drug development (Table 3).

Despite of all the caveats noted and the potential difficulty in translating the findings from primates to humans, these models can be and have been highly instructive in establishing certain basic principles that would have been difficult or impossible to determine by experimentation in humans for safety reasons.

Of note, the suitability of each model depends on the study's specific question, the available tools and appropriate interpretation of the results. Nonetheless, if used correctly, it constitutes an essential model, in many aspects, for HIV research until a vaccine and therapy for HIV cure or HIV remission are discovered.

Table 1. Examples of major contributions of NHP models to HIV/AIDS research

□ Discovery of initial founder populations of infected cells (foci) <sup>30</sup>
□ CD8+T cell response: «too little, too late» to clear infection <sup>86</sup>
□ Short window of opportunity to prevent infection <sup>38</sup>
□ CD8+T cells : impact on viral set-point <sup>74,140</sup>
□ Proof of concept that Nab can protect against infection <sup>141</sup>
□ Nef viral protein: necessary for high viral load <i>invivo</i> <sup>72</sup>
□ Resting memory T cells: main target of the virus in lymphoid tissues <sup>24,27</sup>
□ Rapid and dramatic depletion of CD4+T cells in gut <sup>22,27</sup>
□ Trafficking of Treg, PDC, NK cells to the gut <sup>142–144</sup>
□ Loss of Tcm associated with disease progression <sup>24,145</sup>
□ Events in acute infection determine disease progression <sup>70</sup>
□ TRIM5α alleles restrict viral replication <i>invivo</i> <sup>86</sup>
□ Analyses of the virome and microbiome in tissues <sup>31–134</sup>
□ <i>In vivo</i> imaging of SIV <sup>77</sup>

<b>Model</b>	<b>Characteristics</b>	<b>Examples of utility</b>	<b>Drawbacks</b>
<b><i>Alternative viruses</i></b>			
SHIV	Chimeric viruses that express HIV proteins in aSIV backbone to replicate in NHP.	Antiretroviral development Vaccination strategies Study of viral restriction	Variability in the infectivity Lack of AIDS induction Unusual pathophysiology
<b><i>Alternative hosts</i></b>			
SCID-hu <sup>150</sup>	Human hematopoietic progenitor stem cells that mature mainly into T cells.	Study of CD4+ T cell loss Test of antiretroviral drugs <sup>126</sup>	Absence of innate immunity. Not suitable to study viral transmission.
Hu-HSC	Irradiated non-obese diabetic (NOD) mice injected with human hematopoietic stem cells. Produces human T and B cells.	Neuropathogenesis <sup>151</sup> Viral latency <sup>151</sup>	Absence of innate immune response Not suitable to study primary infection
BLT <sup>152</sup> (Bone-liver-thymus)	Human T and B lymphocytes, monocytes, macrophages, NK and dendritic cells. Develops mucosal immunity, including GALT.	Antiretroviral treatment tests <sup>124</sup> Studies of viral transmission and replication in mucosa <sup>124,125,153</sup>	Defect in class-switched antibody production. Predominance of pre-mature B cells <sup>154</sup>

Table 3. NHPuse towards a cureforHIV: advantagesand applications

<b>Aim</b>	<b>AdvantageofNHPuse</b>	<b>Examples ofstrategies studied</b>
<b>Pharmacological viral suppressionanddiminution ofviral reservoirs<sup>78,155,156</sup></b>	Control of viral strain Control of viral dose andnatureof inoculum Control of route of administrationand timing	Diffusion of antiretroviral drugs in tissues Veryearlytreatment initiation Treatment withIFN- <input type="checkbox"/> Treatment with bNabs
<b>Restaurationandenhancing ofimmune response<sup>85,86</sup></b>	Control of duration, typeand combination of antiretrovirals Extensive characterization ofviral reservoirsin tissues	Treatment withInterleukins (IL-7,IL-15,IL-21) Treatment with TLR7agonists Treatment with anti PD-1
<b>Purgelatent HIV-reservoir<sup>159–163</sup></b>	Analyses of immune responses in tissues includingimmediatelyafterinfection Intervention in the earlystageof primary infection Administration ofpotentiallyriskysubstances	Administration ofinhibitors of chromatin-remodelingmolecules (SAHA-inhibitors) Gene editing Nanoparticles

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## CONFLICT OF INTERESTS

The authors declare not to have any conflict of interests.

1. Barré-Sinoussi, F.*et al.* Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immunodeficiency syndrome (AIDS). 1983. *Rev. Investig. Clínica Organo Hosp. Enfermedades Nutr.* **56**, 126–129 (2004).
2. Paiardini, M. & Müller-Trutwin, M. HIV-associated chronic immune activation. *Immunol. Rev.* **254**, 78–101 (2013).
3. Peeters, M., Jung, M. & Ayoub, A. The origin and molecular epidemiology of HIV. *Expert Rev. Anti Infect. Ther.* **11**, 885–896 (2013).
4. Hemelaar, J. The origin and diversity of the HIV-1 pandemic. *Trends Mol. Med.* **18**, 182–192 (2012).
5. Chakrabarti, L. *et al.* Sequence of simian immunodeficiency virus from macaque and its relationship to other human and simian retroviruses. *Nature* **328**, 543–547 (1987).
6. Hirsch, V. M., Olmsted, R. A., Murphey-Corb, M., Purcell, R. H. & Johnson, P. R. An African primate lentivirus (SIVsm) closely related to HIV-2. *Nature* **339**, 389–392 (1989).
7. Gao, F. *et al.* Origin of HIV-1 in the chimpanzee Pan troglodytes. *Nature* **397**, 436–441 (1999).
8. Bailes, E. *et al.* Hybrid origin of SIV in chimpanzees. *Science* **300**, 1713 (2003).
9. Chen, Z. *et al.* Genetic characterization of new West African simian immunodeficiency virus SIVsm: geographic clustering of household-derived SIV strains with human immunodeficiency virus type 2 subtypes and genetically diverse viruses from a single feral sooty mangabey troop. *J. Virol.* **70**, 3617–3627 (1996).
10. Peeters, M. *et al.* Isolation and partial characterization of an HIV-related virus occurring naturally in chimpanzees in Gabon. *AIDS* **3**, 625–630 (1989).
11. Simon, F. *et al.* Identification of a new human immunodeficiency virus type 1 distinct from group M and group O. *Nat. Med.* **4**, 1032–1037 (1998).

12. Corbet,S. *et al.* envSequencesof SimianImmunodeficiencyVirusesfromChimpanzees in CameroonAre StronglyRelatedto ThoseofHumanImmunodeficiencyVirus GroupN from the SameGeographicArea.*J. Virol.* **74**,529–534(2000).
13. Müller-Trutwin,M. C.*et al.* SIVcpzfromanaturallyinfectedCameroonianchimpanzee: biologicalandgenetic comparisonwith HIV-1N. *J.Med. Primatol.* **29**,166–172(2000).
14. Nerrienet,E.*et al.* SimianImmunodeficiencyVirus Infectionin Wild-CaughtChimpanzees fromCameroon.*J. Virol.* **79**,1312–1319(2005).
15. Keele, B. F. *et al.* ChimpanzeereservoirsofpandemicandnonpandemicHIV-1.*Science* **313**, 523–526(2006).
16. Peeters,M.&Delaporte,E.Simianretroviruses in Africanapes.*Clin. Microbiol. Infect. Off. Publ.Eur.Soc. Clin. Microbiol. Infect. Dis.* **18**,514–520(2012).
17. VanHeuverswyn,F.*et al.* Humanimmunodeficiencyviruses:SIVinfectionin wildgorillas. *Nature* **444**,164(2006).
18. Plantier,J.-C. *et al.*A newhumanimmunodeficiencyvirus derivedfromgorillas.*Nat. Med.* **15**,871–872(2009).
19. Takehisa,J. *et al.*Originandbiology ofsimianimmunodeficiencyvirus inwild-livingwestern gorillas. *J. Virol.* **83**, 1635–1648(2009).
20. D'arc,M.*et al.* Originof theHIV-1groupOepidemicinwestern lowlandgorillas. *Proc. Natl. Acad. Sci. U. S. A.* **112**, E1343–1352(2015).
21. Letvin, N.L.*et al.* Acquiredimmunodeficienciesyndromeinacolonyofmacaque monkeys. *Proc. Natl. Acad.Sci.U. S.A.* **80**,2718–2722(1983).
22. Veazey,R.S. *et al.* Gastrointestinaltractas amajor site ofCD4+Tcelldepletionandviral replicationin SIV infection.*Science* **280**,427–431(1998).

23. Kewenig, S. *et al.* Rapid mucosal CD4(+) T-cell depletion and enteropathy in simian immunodeficiency virus-infected rhesus macaques. *Gastroenterology* **116**, 1115–1123 (1999).
24. Mattapallil, J.J. *et al.* Massive infection and loss of memory CD4+ T cells in multiple tissues during acute SIV infection. *Nature* **434**, 1093–1097 (2005).
25. Schneider, T. *et al.* Loss of CD4 T lymphocytes in patients infected with human immunodeficiency virus type 1 is more pronounced in the duodenal mucosa than in the peripheral blood. Berlin Diarrhea/Wasting Syndrome Study Group. *Gut* **37**, 524–529 (1995).
26. Brenchley, J.M. *et al.* CD4+ T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract. *J. Exp. Med.* **200**, 749–759 (2004).
27. Li, Q. *et al.* Peak SIV replication in resting memory CD4+ T cells depletes gut lamina propria CD4+ T cells. *Nature* **434**, 1148–1152 (2005).
28. Schmitz, J.E. *et al.* Memory CD4+ T lymphocytes in the gastrointestinal tract: A major source of cell-associated simian immunodeficiency virus in chronic nonpathogenic infection of African green monkeys. *J. Virol.* **86**, 11380–11385 (2012).
29. Chomont, N. *et al.* HIV reservoir size and persistence are driven by T cell survival and homeostatic proliferation. *Nat. Med.* **15**, 893–900 (2009).
30. Haase, A.T. Targeting early infection to prevent HIV-1 mucosal transmission. *Nature* **464**, 217–223 (2010).
31. Fennessey, C.M. & Keele, B.F. Using Nonhuman Primates to Model HIV Transmission. *Curr. Opin. HIV AIDS* **8**, 280–287 (2013).
32. Evans, D.T. & Silvestri, G. Non-Human Primate Models in AIDS Research. *Curr. Opin. HIV AIDS* **8**, 255–261 (2013).

33. Johnson, R.P.&Desrosiers, R.C. Protective immunity induced by live attenuated simian immunodeficiency virus. *Curr. Opin. Immunol.* **10**, 436–443(1998).
34. Koff, W. C. *et al.* HIV vaccine design: insights from live attenuated SIV vaccines. *Nat. Immunol.* **7**, 19–23(2006).
35. Hansen, S.G. *et al.* Immune clearance of highly pathogenic SIV infection. *Nature* **502**, 100–104(2013).
36. Daniel, M.D., Kirchhoff, F., Czajak, S. C., Sehgal, P.K. & Desrosiers, R.C. Protective effects of a live attenuated SIV vaccine with a deletion in the nef gene. *Science* **258**, 1938–1941(1992).
37. Billingsley, J.M. *et al.* Characterization of CD8+ T Cell Differentiation following SIVΔnef Vaccination by Transcription Factor Expression Profiling. *PLoS Pathog.* **11**, (2015).
38. Sodora, D.L., Gettie, A., Miller, C. J. & Marx, P. A. Vaginal transmission of SIV: assessing infectivity and hormonal influences in macaques inoculated with cell-free and cell-associated viral stocks. *AIDS Res. Hum. Retroviruses* **14 Suppl 1**, S119–123(1998).
39. Milush, J. M. *et al.* Rapid dissemination of SIV following oral inoculation. *AIDS Lond. Engl.* **18**, 2371–2380(2004).
40. Keele, B. F. *et al.* Low-dose rectal inoculation of rhesus macaques by SIVsmE660 or SIVmac251 recapitulates human mucosal infection by HIV-1. *J. Exp. Med.* **206**, 1117–1134 (2009).
41. Tsai, L., Tasovski, I., Leda, A. R., Chin, M.P. & Cheng-Mayer, C. The number and genetic relatedness of transmitted/founder virus impact clinical outcome in vaginal R5 SHIVSF162P3N infection. *Retrovirology* **11**, 22(2014).
42. Tully, D. C. *et al.* Differences in the Selection Bottleneck between Modes of Sexual Transmission Influence the Genetic Composition of the HIV-1 Founder Virus. *PLoS Pathog.* **12**, e1005619(2016).

43. Ma, Z.-M. *et al.* SIVmac251 Is Inefficiently Transmitted to Rhesus Macaques by Penile Inoculation with a Single SIVenv Variant Found in Ramp-up Phase Plasma. *AIDS Res. Hum. Retroviruses* **27**, 1259–1269 (2011).
44. Keele, B. F. & Estes, J. D. Barrier-to-mucosal transmission of immunodeficiency viruses. *Blood* **118**, 839–846 (2011).
45. Gambhira, R. *et al.* Transmitted/Founder Simian Immunodeficiency Virus Envelope Sequences in Vesicular Stomatitis and Semliki Forest Virus Vector Immunized Rhesus Macaques. *PLoS ONE* **9**, (2014).
46. Nombenge, F. J. *et al.* Development of AIDS in a chimpanzee infected with human immunodeficiency virus type 1. *J. Virol.* **71**, 4086–4091 (1997).
47. Davis, I. C., Girard, M. & Fultz, P. N. Loss of CD4+ T cells in human immunodeficiency virus type 1-infected chimpanzees associated with increased lymphocyte apoptosis. *J. Virol.* **72**, 4623–4632 (1998).
48. O’Neil, S. P. *et al.* Progressive infection in a subset of HIV-1-positive chimpanzees. *J. Infect. Dis.* **182**, 1051–1062 (2000).
49. Ferrari, G. *et al.* The impact of HIV-1 infection on phenotypic and functional parameters of cellular immunity in chimpanzees. *AIDS Res. Hum. Retroviruses* **9**, 647–656 (1993).
50. Juompan, L. Y. *et al.* Analysis of the immune responses in chimpanzees infected with HIV type 1 isolates. *AIDS Res. Hum. Retroviruses* **24**, 573–586 (2008).
51. Institute of Medicine (US) and National Research Council (US) Committee on the Use of Chimpanzees in Biomedical and Behavioral Research. *Chimpanzees in Biomedical and Behavioral Research: Assessing the Necessity*. (National Academies Press (US), 2011).
52. Stremlau, M. *et al.* The cytoplasmic body component TRIM5alpha restricts HIV-1 infection in Old World monkeys. *Nature* **427**, 848–853 (2004).

53. Misra, A., Thippeshappa, R. & Kimata, J.T. Macaques as model hosts for studies of HIV-1 infection. *Front. Microbiol.* **4**, 176 (2013).
54. Wu, F. *et al.* TRIM5 $\alpha$  restriction affects clinical outcome and disease progression in simian immunodeficiency virus-infected rhesus macaques. *J. Virol.* **89**, 2233–2240 (2015).
55. Lim, S.-Y. *et al.* TRIM5 $\alpha$  Modulates Immunodeficiency Virus Control in Rhesus Monkeys. *PLoS Pathog.* **6**, (2010).
56. Hatzioannou, T. *et al.* HIV-1-induced AIDS in monkeys. *Science* **344**, 1401–1405 (2014).
57. Thippeshappa, R., Ruan, H. & Kimata, J.T. Breaking Barriers to an AIDS Model with Macaque-Tropic HIV-1 Derivatives. *Biology* **1**, 134–164 (2012).
58. Mascola, J. R. *et al.* Protection of macaques against vaginal transmission of a pathogenic HIV-1/SIV chimeric virus by passive infusion of neutralizing antibodies. *Nat. Med.* **6**, 207–210 (2000).
59. Rudicell, R. S. *et al.* Enhanced Potency of a Broadly Neutralizing HIV-1 Antibody In Vitro Improves Protection against Lentiviral Infection In Vivo. *J. Virol.* **88**, 12669–12682 (2014).
60. Barouch, D. H. *et al.* Therapeutic Efficacy of Potent Neutralizing HIV-1-Specific Monoclonal Antibodies in SHIV-Infected Rhesus Monkeys. *Nature* **503**, 224–228 (2013).
61. Shingai, M. *et al.* ANTIBODY-MEDIATED IMMUNOTHERAPY OF MACAQUES CHRONICALLY INFECTED WITH SHIV SUPPRESSES VIREMIA. *Nature* **503**, 277–280 (2013).
62. Gengiah, T. N. *et al.* A drug evaluation of 1% tenofovir gel and tenofovir disoproxil fumarate tablets for the prevention of HIV infection. *Expert Opin. Investig. Drugs* **21**, 695–715 (2012).
63. Hessell, A. J. & Haigwood, N. L. Animal Models in HIV-1 Protection and Therapy. *Curr. Opin. HIV/AIDS* **10**, 170–176 (2015).
64. Veazey, R. S. Animal Models for Microbicide Safety and Efficacy Testing. *Curr. Opin. HIV/AIDS* **8**, 295–303 (2013).

65. Wilson, N.A.&Watkins,D.I. Is an HIVVaccinePossible?*Braz. J. Infect. Dis. Off.Publ.Braz. Soc. Infect. Dis.***13**, 304–310(2009).
66. Dezzutti, C.S. Animalandhumanmucosal tissuemodels to studyHIVbiomedical interventions:canwepredictsuccess?*J. Int. AIDS Soc.***18**,(2015).
67. Hatzioannou,T. &Evans,D. T. Animalmodels for HIV/AIDS research.*Nat. Rev.Microbiol.***10**, 852–867(2012).
68. Policicchio,B. B.,Pandrea, I. &Apetrei, C. Animal Models forHIV CureResearch.*Front. Immunol.* **7**,(2016).
69. Passaes, C.P.&Sáez-Cirión, A. HIVcureresearch:advancesandprospects. *Virology***454-455**, 340–352(2014).
70. Saez-Cirion, A., Jacquelin, B., Barré-Sinoussi,F. &Müller-Trutwin,M. Immuneresponses duringspontaneous control of HIV andAIDS:what is thehopefora cure?*Philos.Trans. R. Soc. B Biol.Sci.* **369**,(2014).
71. Bruel, T. *et al.* Long-TermControlofSimianImmunodeficiencyVirus (SIV) inCynomolgus MacaquesNotAssociatedwith Efficient SIV-Specific CD8+T-CellResponses.*J. Virol.***89**, 3542–3556(2015).
72. Kestler,H. W.*et al.* Importanceof thenef geneformaintenanceof highvirus loads andfor developmentof AIDS.*Cell***65**,651–662(1991).
73. Mothé, B.R. *et al.* ExpressionoftheMajor HistocompatibilityComplexClass I Molecule Mamu-A\*01Is Associatedwith ControlofSimianImmunodeficiencyVirus SIVmac239 Replication. *J. Virol.***77**,2736–2740(2003).
74. Nomura,T.&Matano, T. AssociationofMHC-I genotypeswith diseaseprogressionin HIV/SIV infections.*Front.Microbiol.***3**,(2012).

75. Walter,L.&Ansari, A. A.MHC andKIRPolymorphismsin RhesusMacaqueSIVInfection.  
*Front. Immunol.* **6**, 540(2015).
76. Albrecht, C.*et al.* ProgressiontoAIDSin SIV-InfectedRhesus MacaquesAssociatedwith DistinctKIR andMHCclassI Polymorphisms andNKCell Dysfunction.*Front. Immunol.* **5**,600 (2014).
77. Santangelo,P. J.*et al.* Whole-bodyimmunoPETreveals activeSIV dynamicsinviremicand antiretroviraltherapy-treated macaques.*Nat.Methods* **12**,427–432(2015).
78. Whitney,J.B.*et al.* RapidSeedingofthe ViralReservoir Prior toSIVViremia in Rhesus Monkeys.*Nature* **512**,74–77(2014).
79. Sellier,P. *et al.* Antiretroviral Treatment Start-TimeduringPrimarySIVmac Infectionin MacaquesExertsaDifferentImpactonEarlyViral ReplicationandDissemination.*PLoS ONE* **5**,(2010).
80. Kader,M. *et al.* AntiretroviralTherapyprioro toAcuteViralReplicationPreservesCD4TCells inthePeripherybutNotinRectalMucosa duringAcuteSimianImmunodeficiencyVirus Infection.  
*J. Virol.* **82**, 11467–11471(2008).
81. Micci,L. *et al.* CD4Depletionin SIV-InfectedMacaquesResultsin MacrophageandMicroglia Infectionwith RapidTurnoverof InfectedCells.*PLoS Pathog.* **10**,(2014).
82. Schmitz, J.E.*et al.* Controlof Viremia inSimianImmunodeficiencyVirus InfectionbyCD8+ Lymphocytes.*Science* **283**,857–860(1999).
83. Matano, T.*et al.* Administrationof an anti-CD8monoclonal antibodyinterfereswith the clearanceofchimericsimian/humanimmunodeficiency virusduringprimaryinfectionsof rhesus macaques.*J. Virol.* **72**,164–169(1998).
84. Jin, X. *et al.* DramaticRiseinPlasma Viremia afterCD8+TCellDepletionin Simian ImmunodeficiencyVirus–infectedMacaques.*J. Exp. Med.* **189**,991–998(1999).

85. O'Connor,D. H.*et al.* AcutePhasecytotoxicTlymphocyteescapeisa hallmarkofsimian immunodeficiencyvirus infection.*Nat. Med.* **8**,493–499(2002).
86. Reynolds,M. R.*et al.* CD8+T-LymphocyteResponsetoMajor ImmunodominantEpitopes afterVaginal ExposuretoSimianImmunodeficiencyVirus:TooLateandTooLittle.*J. Virol.* **79**,9228–9235(2005).
87. Haase,A.T.EarlyEventsinSexualTransmissionof HIVandSIVandOpportunitiesfor Interventions.*Annu. Rev.Med.* **62**,127–139(2011).
88. Choi, E. I.,Reimann, K.A. &Letvin, N.L. InVivoNatural KillerCellDepletionduringPrimary SimianImmunodeficiencyVirus Infectionin RhesusMonkeys.*J. Virol.* **82**,6758–6761(2008).
89. Takahashi, Y.*et al.*.InVivoAdministrationof aJAK3Inhibitor toChronicallySIVInfected Rhesus MacaquesLeadstoNKCellDepletionAssociated withTransientModestIncreasein ViralLoads.*PLoS ONE* **8**,(2013).
90. Keele, B. F. *et al.* IncreasedmortalityandAIDS-likeimmunopathologyinwildchimpanzees infectedwith SIVcpz.*Nature* **460**,515–519(2009).
91. Etienne,L.*et al.* Characterizationof anewsimianimmunodeficiencyvirus strainina naturallyinfectedPantroglodytestestroglodyteschimpanzee withAIDS relatedsymptoms. *Retrovirology* **8**,4(2011).
92. Sodora,D.L.*et al.* TowardanAIDS vaccine:lessons fromnaturalsimianimmunodeficiency virus infectionsof Africannonhumanprimatehosts.*Nat. Med.* **15**,861–865(2009).
93. Chahroudi, A., Bosinger,S.E., Vanderford,T. H.,Paiardini, M. &Silvestri, G. Natural SIV Hosts:ShowingAIDStheDoor.*Science* **335**,(2012).
94. Kornfeld,C.*et al.* AntiinflammatoryprofilesduringprimarySIVinfectionin Africangreen monkeys areaassociated with protectionagainstAIDS.*J. Clin. Invest.* **115**,1082–1091(2005).

95. Chakrabarti, L. A. *et al.* Normal T-Cell Turnover in Sooty Mangabeys Harboring Active Simian Immunodeficiency Virus Infection. *J. Virol.* **74**, 1209–1223 (2000).
96. Rotger, M. *et al.* Comparative transcriptomics of extreme phenotypes of human HIV-1 infection and SIV infection in sooty mangabey and rhesus macaque. *J. Clin. Invest.* **121**, 2391–2400 (2011).
97. Kirchhoff, F. Is the high virulence of HIV-1 an unfortunate coincidence of primate lentiviral evolution? *Nat. Rev. Microbiol.* **7**, 467–476 (2009).
98. Estes, J. D. *et al.* Damaged Intestinal Epithelial Integrity Linked to Microbial Translocation in Pathogenic Simian Immunodeficiency Virus Infections. *PLoS Pathog.* **6**, (2010).
99. Pandrea, I. *et al.* Cutting Edge: Experimentally Induced Immune Activation in Natural Hosts of Simian Immunodeficiency Virus Induces Significant Increases in Viral Replication and CD4+T Cell Depletion. *J. Immunol. Baltim. Md 1950* **181**, 6687–6691 (2008).
100. Breed, M. W. *et al.* Loss of a Tyrosine-Dependent Trafficking Motif in the Simian Immunodeficiency Virus Envelope Cytoplasmic Tail Spares Mucosal CD4 Cells but Does Not Prevent Disease Progression. *J. Virol.* **87**, 1528–1543 (2013).
101. Jacqueline, B. *et al.* Nonpathogenic SIV infection of African green monkeys induces a strong but rapidly controlled type I IFN response. *J. Clin. Invest.* **119**, 3544–3555 (2009).
102. Jochems, S. P. *et al.* Plasmacytoid Dendritic Cell Infection and Sensing Capacity during Pathogenic and Nonpathogenic Simian Immunodeficiency Virus Infection. *J. Virol.* **89**, 6918–6927 (2015).
103. Schindler, M. *et al.* Nef-Mediated Suppression of T Cell Activation Was Lost in a Lentiviral Lineage that Gave Rise to HIV-1. *Cell* **125**, 1055–1067 (2006).
104. Paiardini, M. *et al.* Low levels of SIV infection in sooty mangabey central-memory CD4+T cells is associated with limited CCR5 expression. *Nat. Med.* **17**, 830–836 (2011).

105. Veazey, R. *et al.* Decreased CCR5 Expression on CD4+ T Cells of SIV-Infected Sooty Mangabeys. *AIDS Res. Hum. Retroviruses* **19**, 227–233 (2003).
106. Pandrea, I. *et al.* Paucity of CD4+CCR5+T Cells May Prevent Transmission of Simian Immunodeficiency Virus in Natural Nonhuman Primate Hosts by Breast-Feeding. *J. Virol.* **82**, 5501–5509 (2008).
107. Ploquin, M. J., Silvestri, G. & Müller-Trutwin, M. Immune activation in HIV infection: what can the natural hosts of simian immunodeficiency virus teach us? *Curr. Opin. HIV AIDS* **11**, 201–208 (2016).
108. Gueye, A. *et al.* Viral load in tissues during the early and chronic phase of non-pathogenic SIVagm infection. *J. Med. Primatol.* **33**, 83–97 (2004).
109. Martinot, A. J. *et al.* Acute SIV Infection in Sooty Mangabey Monkeys Is Characterized by Rapid Virus Clearance from Lymph Nodes and Absence of Productive Infection in Germinal Centers. *PLoS ONE* **8**, (2013).
110. Brenchley, J. M. *et al.* Differential infection patterns of CD4+T cells and lymphoid tissue viral burden distinguish progressive and non-progressive lentiviral infections. *Blood* **120**, 4172–4181 (2012).
111. Diop, O. M. *et al.* High Levels of Viral Replication during Primary Simian Immunodeficiency Virus SIVagm Infection Are Rapidly and Strongly Controlled in African Green Monkeys. *J. Virol.* **74**, 7538–7547 (2000).
112. Goldstein, S. *et al.* Wide Range of Viral Load in Healthy African Green Monkeys Naturally Infected with Simian Immunodeficiency Virus. *J. Virol.* **74**, 11744–11753 (2000).
113. Beer, B. *et al.* Lack of dichotomy between virus load of peripheral blood and lymph nodes during long-term simian immunodeficiency virus infection of African green monkeys. *Virology* **219**, 367–375 (1996).

114. Ploquin,M. J.-Y., Jacquelin,B., Jochems, S.P., Barré-Sinoussi, F.&Müller-Trutwin, M. C. Innateimmunityin the controlof HIV/AIDS:recent advancesandopenquestions.*AIDS Lond. Engl.* **26**,1269–1279(2012).
115. Ploquin,M. J.-Y.*et al.* DC-SIGNfromAfricanGreenMonkeys Is ExpressedinLymphNodes andMediatesInfectionintrans of SimianImmunodeficiencyVirusSIVagm.*J. Virol.* **78**,798–810(2004).
116. Cumont,M.-C. *et al.* EarlyDivergencein LymphoidTissue Apoptosis betweenPathogenicand NonpathogenicSimianImmunodeficiencyVirus Infectionsof NonhumanPrimates.*J. Virol.* **82**,1175–1184(2008).
117. Hurtrel,B. *et al.* Apoptosis inSIVinfection. *Cell Death Differ.* **12**,979–990(2005).
118. Elder,J. H., Lin,Y.-C., Fink,E. &Grant,C.K. FelineImmunodeficiencyVirus (FIV)as A Model for StudyofLentivirus Infections:Parallelswith HIV.*Curr. HIV Res.* **8**,73–80(2010).
119. Mohammadi, H.&Bienzle,D. Pharmacological InhibitionofFelineImmunodeficiencyVirus (FIV). *Viruses* **4**,708–724(2012).
120. Denton,P. W.&Garcia, J.V. Humanized MouseModels ofHIV Infection.*AIDS Rev.* **13**,135–148(2011).
121. Garcia, J.V. Invivoplatforms for analysisof HIV persistenceanderadication.*J. Clin. Invest.* **126**,424–431(2016).
122. Legrand, N. *et al.* Humanized miceformodelinghumaninfectious disease:challenges, progress, andoutlook.*Cell Host Microbe* **6**,5–9(2009).
123. Seung, E.*et al.* PD-1BlockadeinChronicallyHIV-1-InfectedHumanized MiceSuppressesViral Loads.*PLoS ONE* **8**,(2013).

124. Kovarova,M.*et al.*A long-actingformulationof theintegraseinhibitor raltegravir protects humanized BLTmicefromrepeatedhigh-dosevaginalHIVchallenges.*J. Antimicrob. Chemother.* dkw042(2016). doi:10.1093/jac/dkw042
125. Olesen,R., Wahl,A., Denton, P. W.&Garcia,J. V.ImmuneReconstitutionof theFemale ReproductiveTract ofHumanized BLTMiceandtheirSusceptibilitytoHuman ImmunodeficiencyVirusInfection.*J. Reprod. Immunol.* **88**,195–203(2011).
126. Sundell,I.B., Cortado,R.V.&Koka,P. S.Sulfatide--a newcandidatefor ARTtreatmentin HIV-1infection.*J. StemCells* **7**,61–72(2012).
127. Honeycutt,J. B.*et al.* MacrophagessustainHIV replicationinvivo independentlyof Tcells.*J. Clin. Invest.* **126**,1353–1366(2016).
128. Miller,C.J. *et al.* Genitalmucosal transmissionof simianimmunodeficiencyvirus:animal model for heterosexualtransmissionofhumanimmunodeficiencyvirus.*J. Virol.* **63**,4277–4284(1989).
129. Veazey,R.S., Shattock,R. J., Klasse,P.J.&Moore,J. P. Animalmodels formicrobicide studies. *Curr.HIVRes.* **10**,79–87(2012).
130. Bernard-Stoecklin, S.,Gommet, C., Cavarelli, M.&Grand, R. L. NonhumanPrimateModels for Cell-AssociatedSimianImmunodeficiencyVirus Transmission:TheNeed toBetter UnderstandtheComplexity ofHIV MucosalTransmission.*J. Infect. Dis.* **210**,S660–S666 (2014).
131. Bernard-Stoecklin, S.*et al.*SemenCD4+TCells andMacrophagesAreProductivelyInfected atAll StagesofSIVinfectionin Macaques. *PLoSPathog.* **9**,(2013).
132. Pandrea, I.*et al.*Mucosal SimianImmunodeficiencyVirus TransmissioninAfricanGreen Monkeys:SusceptibilitytoInfectionIs Proportional toTargetCellAvailabilityatMucosal Sites. *J. Virol.* **86**,4158–4168(2012).

133. Chahroudi, A. *et al.* Mother-to-Infant Transmission of Simian Immunodeficiency Virus Is Rare in Sooty Mangabeys and Is Associated with Low Viremia. *J. Virol.* **85**, 5757–5763 (2011).
134. Ma, D. *et al.* Factors Associated with Simian Immunodeficiency Virus Transmission in a Natural African Nonhuman Primate Host in the Wild. *J. Virol.* **88**, 5687–5705 (2014).
135. Marlin, R. *et al.* Decidual soluble factors participate in the control of HIV-1 infection at the materno-fetal interface. *Retrovirology* **8**, 58 (2011).
136. Tripathi, V. *et al.* Differential expression of RDC1/CXCR7 in the human placenta. *J. Clin. Immunol.* **29**, 379–386 (2009).
137. Veazey, R.S. *et al.* Identifying the Target Cell in Primary Simian Immunodeficiency Virus (SIV) Infection: Highly Activated Memory CD4+ T Cells Are Rapidly Eliminated in Early SIV Infection In Vivo. *J. Virol.* **74**, 57–64 (2000).
138. Li, Q. *et al.* Glycerolmonolaurate prevents mucosal SIV transmission. *Nature* **458**, 1034–1038 (2009).
139. Reimann, K. A. *et al.* Immunopathogenic events in acute infection of rhesus monkeys with simian immunodeficiency virus of macaques. *J. Virol.* **68**, 2362–2370 (1994).
140. Letvin, N.L. *et al.* Cytotoxic T lymphocytes specific for the simian immunodeficiency virus. *Immunol. Rev.* **170**, 127–134 (1999).
141. Barouch, D. H. *et al.* Protective efficacy of adenovirus/protein vaccines against SIV challenges in rhesus monkeys. *Science* **349**, 320–324 (2015).
142. Reeves, R.K., Evans, T. I., Gillis, J. & Johnson, R.P. Simian Immunodeficiency Virus Infection Induces Expansion of  $\alpha 4\beta 7+$  and Cytotoxic CD56+ NK Cells. *J. Virol.* **84**, 8959–8963 (2010).
143. Reeves, R.K. *et al.* CD16- natural killer cells: enrichment in mucosal and secondary lymphoid tissues and altered function during chronic SIV infection. *Blood* **115**, 4439–4446 (2010).

144. Reeves,R.K.*et al.*SIV InfectionInducesAccumulationofPlasmacytoidDendriticCells inthe Gut Mucosa.*J. Infect. Dis.***206**,1462–1468(2012).
145. Picker, L.J. *et al.* Insufficient productionandtissue delivery ofCD4+memoryTcells in rapidly progressesimianimmunodeficiencyvirus infection.*J. Exp. Med.***200**,1299–1314(2004).
146. Monaco,C. L.*et al.*AlteredViromeandBacterial Microbiomein HumanImmunodeficiency Virus- Associated Acquired ImmunodeficiencySyndrome.*Cell HostMicrobe***19**,311–322 (2016).
147. Palmer,B.E.,Li, S.X.&Lozupone,C. A.TheHIV-Associated EntericMicrobiomeHas Gone Viral. *Cell HostMicrobe* **19**,270–272(2016).
148. Barbian, H.J. *et al.*Destabilizationof thegutmicrobiomemarkstheend-stageofsimian immunodeficiency virus infectionin wildchimpanzees.*Am. J. Primatol.*(2015).  
doi:10.1002/ajp.22515
149. Handley, S. A. *et al.*SIV Infection-MediatedChangesinGastrointestinalBacterialMicrobiome andViromeAreAssociated with ImmunodeficiencyandPrevented byVaccination. *Cell Host Microbe***19**,323–335(2016).
150. Namikawa,R.,Kaneshima,H., Lieberman,M.,Weissman, I. L. &McCune,J.M. Infectionof the SCID-huMousebyHIV-1. *Science*(1988).
151. Choudhary,S.K.*et al.*LatentHIV-1InfectionofRestingCD4+TCellsintheHumanized Rag2<sup>-/-</sup>γc<sup>-/-</sup>Mouse.*J. Virol.* **86**,114–120(2012).
152. Wege,A.K.,Melkus, M.W., Denton,P.W., Estes, J.D.&Garcia,J.V. Functionaland phenotypiccharacterizationof thehumanizedBLTmousemodel.*Curr. Top. Microbiol. Immunol.* **324**,149–165(2008).
153. Denton,P. W.*et al.*AntiretroviralPre-exposureProphylaxisPrevents Vaginal Transmissionof HIV-1in Humanized BLTMice.*PLoS Med.***5**,(2008).

154. Karpel,M. E., Boutwell,C. L. &Allen,T. M.BLThumanized miceasa small animalmodelof HIVinfection. *Curr.Opin. Virol.***13**,75–80(2015).
155. Shytaj,I.L.*et al.* AHighlyIntensified ARTRegimen InducesLong-TermViralSuppressionand Restrictionofthe ViralReservoir inaSimianAIDS Model. *PLoSPathog.***8**,(2012).
156. DelPrete,G. Q. *et al.* ShortCommunication:ComparativeEvaluationof Coformulated InjectableCombinationAntiretroviralTherapyRegimens in SimianImmundeficiencyVirus-Infected RhesusMacaques. *AIDSRes. Hum. Retroviruses***32**,163–168(2016).
157. Vassena,L. *et al.* Treatmentwith IL-7Prevents theDecline ofCirculatingCD4+TCells during the AcutePhaseof SIVInfectioninRhesusMacaques. *PLoSPathog.***8**,(2012).
158. Xu, H., Wang,X. & Veazey,R. S.Th17Cells Coordinatewith Th22Cells inMaintaining Homeostasisof Intestinal Tissuesandboth areDepleted inSIV-InfectedMacaques. *J. AIDS Clin. Res.* **5**,(2014).
159. Ling, B.*et al.* Effectsof Treatment with SuppressiveCombinationAntiretroviralDrugTherapy andtheHistoneDeacetylaseInhibitor SuberoylanilideHydroxamic Acid; (SAHA)onSIV-Infected ChineseRhesus Macaques. *PLoS ONE***9**,(2014).
160. DelPrete,G. Q.*et al.* Effectof SuberoylanilideHydroxamic Acid(SAHA) Administrationon the Residual Virus Pool inaModelof CombinationAntiretroviralTherapy-Mediated Suppressionin SIVmac239-Infected IndianRhesusMacaques. *Antimicrob. AgentsChemother.* **58**,6790–6806(2014).
161. Trautmann,L. Kill:boostingHIV-specificimmuneresponses. *Curr. Opin. HIVAIDS*(2016).doi:10.1097/COH.0000000000000286
162. Datta,P.K.*et al.* HIV-1LatencyandEradication:Past,Present andFuture. *Curr. HIVRes.* (2016).

163. Wen, J. *et al.* Specific Elimination of Latently HIV-1 Infected Cells Using HIV-1 Protease-Sensitive Toxin Nanocapsules. *PLoS One* **11**, e0151572 (2016).

**Figure 1**

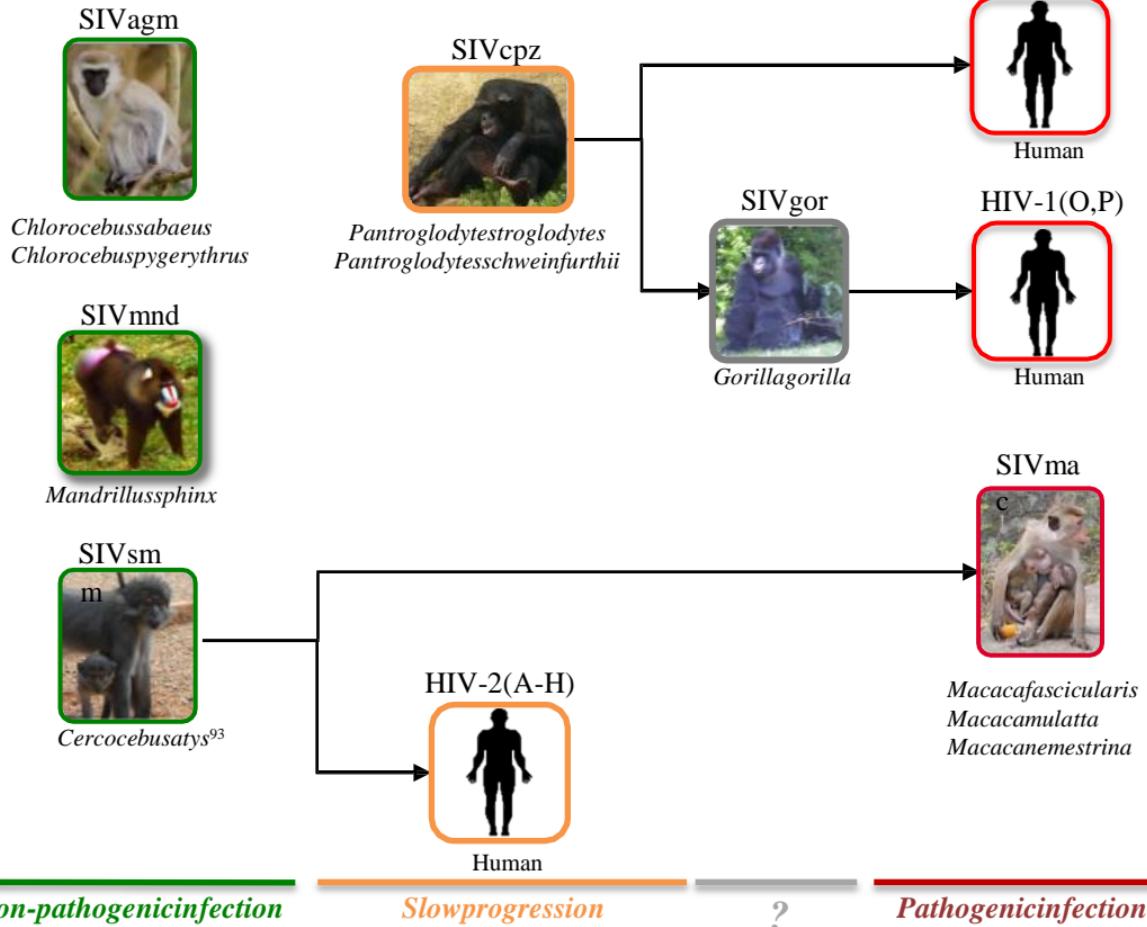


Figure 2

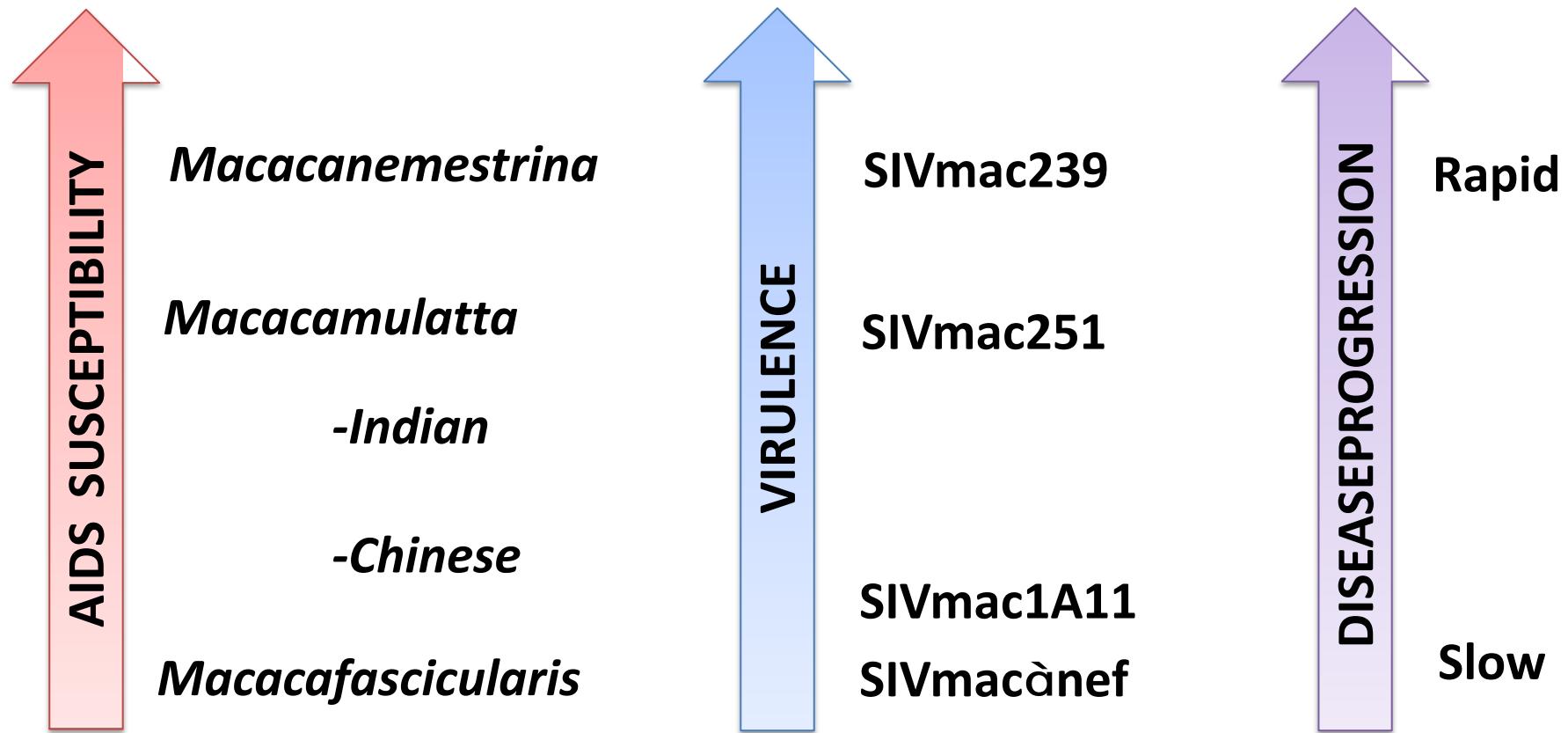
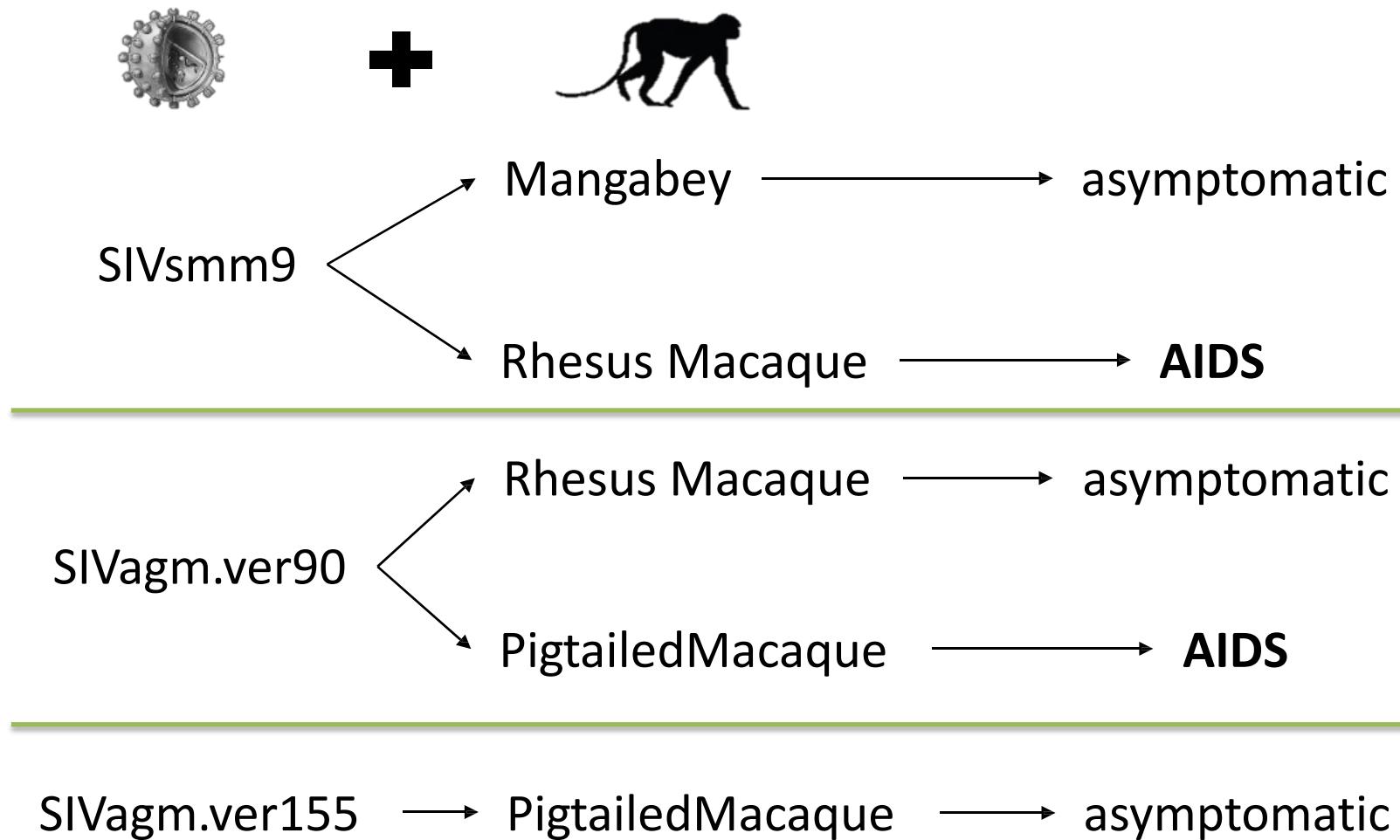


Figure 3



Captions

**Figure1.** Non-human primate models for the study of HIV infection. Non-human primates can be divided into pathogenic (macaques) and non-pathogenic (sooty mangabeys, mandrills and African Green Monkeys) models for HIV research. The figure depicts the relationship between the HIV and SIV viruses and the type of infection caused by them.

**Figure2.** Host and viral determinants of disease progression rate. The progression of the disease depends on the intrinsic susceptibility of each species to develop AIDS and on the virulence of the SIV strain used.

**Figure3** Cross-species transmissions of SIV and fates of infection. The presence or absence of progression depends on both, viral and host factors; i.e. the infection with a given SIV strain will not cause disease in some primates, whereas in others will lead to the development of AIDS. Host and viral determinants interact with each other and a combination of both determines the outcome of infection.