

Human herpesvirus 8, Southern Siberia.

Olivier Cassar, Sylviane Bassot, Sabine Plancoulaine, Lluis Quintana-Murci, Christine Harmant, Vladimir Gurtsevitch, Natalia B Senyuta, Larissa S Yakovleva, Guy de The, Antoine Gessain

▶ To cite this version:

Olivier Cassar, Sylviane Bassot, Sabine Plancoulaine, Lluis Quintana-Murci, Christine Harmant, et al.. Human herpesvirus 8, Southern Siberia.. Emerging Infectious Diseases, 2010, 16 (3), pp.580-2. 10.3201/eid1603.091390 . pasteur-01370962

HAL Id: pasteur-01370962 https://pasteur.hal.science/pasteur-01370962

Submitted on 23 Sep 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

LETTERS

Acknowledgments

We thank Nadine Jahn, Doris Krumnow, Stephan Schatz, and Robert Schneider for excellent technical assistance and Martin Pfeffer and Holger Scholz for editorial help.

This study was supported by the Adolf and Hildegard Isler Stiftung and the Klara Samariter Stiftung. The multilocus sequence typing database is publicly available at http://mlst.ucc.ie, which is currently supported by a grant from the Science Foundation of Ireland (05/FE1/B882).

Kristin Mühldorfer, Gudrun Wibbelt, Joachim Haensel, Julia Riehm, and Stephanie Speck¹

Author affiliations: Leibniz Institute for Zoo and Wildlife Research, Berlin, Germany (K. Mühldorfer, G. Wibbelt, S. Speck); Berlin, (J. Haensel); and Federal Armed Forces Institute of Microbiology, Munich, Germany (J. Riehm)

DOI: 10.3201/eid1603.091035

¹Current affiliation: Federal Armed Forces Institute of Microbiology, Munich, Germany.

References

- Calisher CH, Childs JE, Field HE, Holmes KV, Schountz T. Bats: important reservoir hosts of emerging viruses. Clin Microbiol Rev. 2006;19:531–45. DOI: 10.1128/ CMR.00017-06
- Wong S, Lau S, Woo P, Yuen K-Y. Bats as a continuing source of emerging infections in humans. Rev Med Virol. 2007;17:67– 91. DOI: 10.1002/rmv.520
- Wibbelt G, Speck S, Field H. Methods for assessing diseases in bats. In: Kunz TH, Parsons S, editors. Ecological and behavioral methods for the study of bats. Baltimore: The Johns Hopkins University Press; 2009. p. 775–94
- MLST Databases at the ERI. University College Cork [cited 2009 Jul 1]. http://mlst. ucc.ie/mlst/dbs/Ypseudotuberculosis/ documents/primersPseudotuberculosis_ html
- Mair NS. Yersiniosis in wildlife and its public health implications. J Wildl Dis. 1973;9:64–71.

- Williams CH. A review of pseudotuberculosis at a European zoo: epidemiology and approaches to control. In: Health and conservation of captive and free-ranging wildlife. Proceedings of the American Association of Zoo Veterinarians, American Association of Wildlife Veterinarians, Wildlife Disease Association Joint Conference; 2004 Aug 28–Sep 3; San Diego, California. p. 303–9.
- Childs-Sanford SE, Kollias GV, Abou-Madi N, McDonough PL, Garner MM, Mohammed HO. *Yersinia pseudotuberculosis* in a closed colony of Egyptian fruit bats (*Rousettus aegyptiacus*). J Zoo Wildl Med. 2009;40:8–14. DOI: 10.1638/2007-0033.1
- Tennant SM, Grant TH, Robins-Browne RM. Pathogenicity of *Yersinia enterocolitica* biotype 1A. FEMS Immunol Med Microbiol. 2003;38:127–37. DOI: 10.1016/ S0928-8244(03)00180-9
- Corbel MJ, Brewer RA, Hunter D. Characterisation of *Yersinia enterocolitica* strains associated with bovine abortion. Vet Rec. 1990;127:526–7.
- Rahuma N, Ghenghesh KS, Ben Aissa R, Elamaari A. Carriage by the housefly (*Musca domestica*) of multiple-antibiotic-resistant bacteria that are potentially pathogenic to humans, in hospital and other urban environments in Misurata, Libya. Ann Trop Med Parasitol. 2005;99:795– 802. DOI: 10.1179/136485905X65134

Address for correspondence: Kristin Mühldorfer, Leibniz Institute for Zoo and Wildlife Research, Research Group Wildlife Diseases, Alfred-Kowalke-Str. 17, D-10315 Berlin, Germany; email: muehldorfer@izw-berlin.de

Letters

Letters commenting on recent articles as well as letters reporting cases, outbreaks, or original research are welcome. Letters commenting on articles should contain no more than 300 words and 5 references; they are more likely to be published if submitted within 4 weeks of the original article's publication. Letters reporting cases, outbreaks, or original research should contain no more than 800 words and 10 references. They may have 1 Figure or Table and should not be divided into sections. All letters should contain material not previously published and include a word count.

Human Herpesvirus 8, Southern Siberia

To the Editor: Human herpesvirus 8 (HHV-8) is the etiologic agent of Kaposi sarcoma. Sequence analysis of the highly variable open reading frame (ORF)–K1 of HHV-8 has enabled the identification of 5 main molecular subtypes, A–E (I). A and C subtypes are prevalent in persons in Europe, Mediterranean countries, northwestern China, and the United States; subtype B, in persons in sub-Saharan Africa; subtype D, in persons in the Pacific Islands and Japan (2–6); and subtype E, in Native Americans in the United States.

Considering that K1 gene polymorphisms of HHV-8-infected persons reflect the divergence accumulated during the early migrations of modern humans out of Africa (1), it is tempting to put the polymorphisms observed in the different subtypes into an evolutionary perspective with their geographic distribution. It is thought that Native Americans infected by subtype E and Pacific Islanders, including those infected by subtype D in the Japanese archipelago, originated from a common ancestral genetic stock in continental Asia. Because Siberia constitutes the geographic link between mainland Asia, North America, and the Pacific (online Technical Appendix, www.cdc. gov/eid/content/16/3/585-Techapp. pdf), it is likely that the Siberian region has served as a source or a corridor of human dispersals to these regions. Thus, we conducted a molecular epidemiology HHV-8 survey of the Buryat population, a major indigenous group in southern Siberia, to gain new insights into the origins, possibly common, of HHV-8 subtypes D and E.

After consent of local authorities and participants, we collected 745 human blood samples in 1995 in 17 medicosocial structures (homes for elderly persons, veterans of the Russian army,

LETTERS

hospitalized persons, blood donors) located near Lake Baïkal and originating from Ulan Ude (344), Ust Orda (216), and Chita (185), Siberia, Russia (additional data can be obtained directly from the authors). The median age of those included was 52 years (range 25-98 years); 489 (66%) were women. Antibodies against HHV-8 latency-associated nuclear antigen were identified by immunofluorescent antibody assay by using the BC3 cell line (3). Punctuate nuclear staining of BC3 cells at a 1:160 dilution was observed for 187 (25.1%) patients with no difference according to investigated regions $(p = 0.32 \text{ by } \chi^2 \text{ test})$ or between men (25.8%) and women (24.7%) (p = 0.76 by χ^2 test; online Technical Appendix). However, HHV-8 seroprevalence increased with patient age, rising from 12.9% (25-43 years) to 46.4% (≥61 years) (p = 1.8×10^{-13} by χ^2 test for trend) (Figure; online Technical Appendix). No significant difference was

observed in antibody titers according to age (p = 0.45 by Fisher exact test). These results demonstrate that HHV-8 infection is highly prevalent in the Siberian adult population tested.

HHV-8 infection was determined by nested PCR that amplified a 737-bp fragment of the ORFK1 in peripheral blood buffy coats of 85 HHV-8-seropositive and 10 HHV-8-seronegative persons (3). Amplification was positive in 19/85 (22.4%) samples; sequences were obtained for 18 of these samples (online Technical Appendix). These sequences showed 0%-7.31% nucleotide divergence and 0%-3.55% amino acid divergence. Nevertheless, 17 strains were found to be closely related with <1.75% nucleotide differences for 684 nt, and only 1 sequence (1445 strain) displayed higher nucleotide divergence.

A comparative sequence analysis, including 66 representatives of K1 gene sequences of the HHV-8 A/C

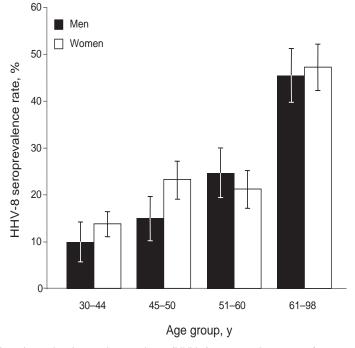


Figure. Age-dependent human herpesvirus 8 (HHV-8) seroprevalence rates for 745 persons in southern Siberia 25–98 years of age who lived in the Ust Orda, Ulan Ude, or Chita districts during 1995. Seropositivity was based on strict criteria; only samples showing punctuate nuclear staining clearly reactive at a dilution ≥1:160 were considered HHV-8 positive. All 187 HHV-8–seropositive samples were tested for antibodies directed against HIV-1/2 by using Genscreen HIV-1/2 Antibody Assay (Bio-Rad Laboratories, Marnes-la-Coquette, France); only 2 were seropositive. Error bars indicate 95% confidence intervals.

subtypes/subgroups, and sequences obtained from persons originating from Russia, was performed (7–9). Seventeen of the 18 HHV-8 strains from Siberia belonged to the A subtype; 15 clustered in a newly identified specific subclade (online Technical Appendix). Notably, the 1445-Siberian strain, which exhibits the typical 5 aa deletion at positions 201–205, belongs to subtype C and clustered with the 7848 strain previously described by Lacoste et al. (9). Furthermore, both strains originate from Chita.

Our results indicate that HHV-8 infection is highly prevalent in the population tested in southern Siberia and extend current knowledge on the worldwide distribution of HHV-8 genotypes. The presence of a Siberian strain monophyletic subclade suggests the existence of HHV-8 strains preferentially spreading among this population in southern Siberia.

To ascertain the maternal ancestry of these persons, we sequenced the hypervariable region I (HVS-I) of the maternally-inherited mitochondrial DNA (mtDNA) and assigned haplogroups on the basis of the HVS-I motifs. Our analyses showed that 17/18 persons analyzed showed a mtDNA motif of clear continental east Asian origin (e.g., A, D correspond to different mtDNA haplogroups). One person (1474-strain) had a lineage (i.e., HV1) that is thought to have a western Eurasian origin. Overall, these mtDNA analyses indicate that the maternal ancestry of the persons examined here can be unambiguously attributed to East Asia, and not to Western Eurasia. K1 subtype A sequences recently found in the Xinjiang Uygur region in China (10) do not correspond to the specific Siberian clade described in our study. Thus, we must now consider that the widely distributed HHV-8 A/C subtype, so far mainly observed in Europe and Mediterranean countries, is also largely predominant in continental Asia.

Olivier Cassar, Sylviane Bassot, Sabine Plancoulaine, Lluis Quintana-Murci, Christine Harmant, Vladimir Gurtsevitch, Natalia B. Senyuta, Larissa S. Yakovleva, Guy de Thé, and Antoine Gessain

Author affiliations: Institut Pasteur, Paris, France (O. Cassar, S. Bassot, S. Plancoulaine, L. Quintana-Murci, G. de Thé, C. Harmant, A. Gessain); Institut National de la Santé et de la Recherche Médicale (INSERM), Paris (S. Plancoulaine); Centre National de la Recherche Scientifique (CNRS), Paris (L. Quintana-Murci); and Blokhin Cancer Research Center, Moscow, Russia (V. Gurtsevitch, N.B. Senyuta, L.S. Yakovleva)

DOI: 10.3201/eid1603.091390

References

- Hayward GS, Zong JC. Modern evolutionary history of the human KSHV genome. Curr Top Microbiol Immunol. 2007;312:1–42. DOI: 10.1007/978-3-540-34344-8_1
- Duprez R, Cassar O, Hbid O, Rougier Y, Morisse L, Bassot S, et al. Cutaneous disseminated endemic Kaposi's sarcoma in a Polynesian man infected with a new divergent human herpesvirus 8 subtype D. J Clin Virol. 2006;37:222–6. DOI: 10.1016/j.jcv.2006.07.013
- Cassar O, Afonso PV, Bassot S, Plancoulaine S, Duprez R, Capuano C, et al. Novel human herpesvirus 8 subtype D strains in Vanuatu, Melanesia. Emerg Infect Dis. 2007;13:1745–8.
- 4. Zong JC, Ciufo DM, Alcendor DJ, Wan X, Nicholas J, Browning PJ, et al. High-level variability in the ORF-K1 membrane protein gene at the left end of the Kaposi's sarcoma–associated herpesvirus genome defines four major virus subtypes and multiple variants or clades in different human populations. J Virol. 1999;73:4156–70.
- Meng YX, Spira TJ, Bhat GJ, Birch CJ, Druce JD, Edlin BR, et al. Individuals from North America, Australasia, and Africa are infected with four different genotypes of human herpesvirus 8. Virology. 1999;261:106–19. DOI: 10.1006/viro.1999.9853
- Cook PM, Whitby D, Calabro ML, Luppi M, Kakoola DN, Hjalgrim H, et al. Variability and evolution of Kaposi's sarcoma–associated herpesvirus in Europe and Africa. International Collaborative Group. AIDS. 1999;13:1165–76. DOI: 10.1097/00002030-199907090-00004
- Davidovici B, Karakis I, Bourboulia D, Ariad S, Zong J, Benharroch D. et al. Seroepidemiology and molecular epidemiology of Kaposi's sarcoma–associated herpesvirus among Jewish population groups in Israel. J Natl Cancer Inst. 2001;93:194–202. DOI: 10.1093/ jnci/93.3.194
- Kadyrova E, Lacoste V, Duprez R, Pozharissky K, Molochkov V, Huerre M, et al. Molecular epidemiology of Kaposi's sarcomaassociated herpesvirus/human herpesvirus 8 strains from Russian patients with classic, posttransplant, and AIDS-associated Kaposi's sarcoma. J Med Virol. 2003;71:548–56. DOI: 10.1002/jmv.10530
- Lacoste V, Kadyrova E, Chistiakova I, Gurtsevitch V, Judde JG, Gessain A. Molecular characterization of Kaposi's sarcoma–associated herpesvirus/human herpesvirus-8 strains from Russia. J Gen Virol. 2000;81:1217–22.
- Fu B, Sun F, Li B, Yang L, Zeng Y, Sun X, et al. Seroprevalence of Kaposi's sarcoma–associated herpesvirus and risk factors in Xinjiang, China. J Med Virol. 2009;81:1422–31. DOI: 10.1002/jmv.21550

Address for correspondence: Antoine Gessain, Unité d'Epidémiologie et Physiopathologie des Virus Oncogènes – Unité de Recherche Appliquée, 3015, Département de Virologie, Bâtiment Lwoff, Institut Pasteur, 28 rue du Dr. Roux, 75724 Paris CEDEX 15, France; email: agessain@pasteur. fr

Sus-Pense

R.L. Bernstein

Don't call it swine flu:

if you dine on pork, you

can't catch the flu, as we know.

But the pork people say

some customers may

be confused by the name and just go.

Now look at the news:

around the world views

show people with a mask on their face.

Despite the name change,

folks who find H1N1 strange

still want to keep swine in their place.

For me it is clear:

I don't have the fear

of swallowing bacon-wrapped figs.

While others may hurry

to wear masks, I don't worry.

I really don't plan to kiss pigs.

Dr. Bernstein is associated with the molecular biology program at New Mexico State University. He carries out research and consults in bioinformatics, biotechnology, and regulatory microbiology.

Author affiliation: New Mexico State University, Las Cruces, New Mexico, USA

Address for correspondence: R.L. Bernstein, New Mexico State University, Molecular Biology Chemistry/Biochem, 1175 N Horseshoe Drive, Las Cruces, NM 88003-8001, USA; email: rbernst@nmsu.edu