



HAL
open science

Assessing the epidemiological effect of wolbachia for dengue control.

Louis Lambrechts, Neil M Ferguson, Eva Harris, Edward C Holmes, Elizabeth A McGraw, Scott L O'Neill, Eng E Ooi, Scott A Ritchie, Peter A Ryan, Thomas W Scott, et al.

► **To cite this version:**

Louis Lambrechts, Neil M Ferguson, Eva Harris, Edward C Holmes, Elizabeth A McGraw, et al.. Assessing the epidemiological effect of wolbachia for dengue control.. The Lancet Infectious Diseases, 2015, 15 (7), pp.862-6. 10.1016/S1473-3099(15)00091-2 . pasteur-01176423

HAL Id: pasteur-01176423

<https://pasteur.hal.science/pasteur-01176423>

Submitted on 16 Jul 2015

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

1 **Assessing the epidemiological impact of *Wolbachia* deployment for**
2 **dengue control**

3

4

5 Louis Lambrechts, PhD,^{1*} Prof. Neil M. Ferguson, PhD,² Eva Harris, PhD,³ Prof. Edward C.
6 Holmes, PhD,⁴ Elizabeth A. McGraw, PhD,⁶ Prof. Scott L. O'Neill, PhD,⁶ Eng E. Ooi, PhD,⁷
7 Prof. Scott A. Ritchie, PhD,⁸ Peter A. Ryan, PhD,⁶ Prof. Thomas W. Scott, PhD,^{9,10} Prof.
8 Cameron P. Simmons, PhD,^{11,12,13} Prof. Scott C. Weaver, PhD¹⁴

9

10

11 ¹ *Insect-Virus Interactions Group, Department of Genomes and Genetics, Institut Pasteur –*
12 *CNRS URA 3012, 28 rue du Docteur Roux, 75724 Paris Cedex 15, France*

13 ² *MRC Centre for Outbreak Analysis and Modelling, School of Public Health, Imperial College*
14 *London, Norfolk Place, London, W2 1PG, United Kingdom*

15 ³ *Division of Infectious Diseases and Vaccinology, School of Public Health, University of*
16 *California, Berkeley, 185 Li Ka Shing Center, Berkeley, CA 94720-3370, USA*

17 ⁴ *Marie Bashir Institute for Infectious Diseases and Biosecurity, Charles Perkins Centre,*
18 *School of Biological Sciences and Sydney Medical School, the University of Sydney, Sydney,*
19 *NSW 2006, Australia*

20 ⁶ *School of Biological Sciences, Monash University, Clayton VIC 3800, Australia*

21 ⁷ *Program in Emerging Infectious Diseases, Duke-National University of Singapore Graduate*
22 *Medical School, 8 College Road, Singapore 169857*

23 ⁸ *School of Public Health and Tropical Medicine and Rehabilitative Sciences, James Cook*
24 *University, Cairns Queensland 4870, Australia*

25 ⁹ *Department of Entomology and Nematology, University of California, Davis, CA 95695, USA*

26 ¹⁰ *Fogarty International Center, National Institutes of Health, Bethesda, MD 20892, USA*

27 ¹¹ *Oxford University Clinical Research Unit, Centre for Tropical Medicine, Ho Chi Minh City,*
28 *District 5, Vietnam*

29 ¹² *Centre for Tropical Medicine, Nuffield Department of Medicine, University of*
30 *Oxford, Oxford OX1 2JD, United Kingdom*

31 ¹³ *Nossal Institute of Global Health, University of Melbourne, Carlton, VIC 3010, Australia*

32 ¹⁴ *Institute for Human Infections and Immunity and Department of Pathology, University of*
33 *Texas Medical Branch, Galveston, TX 77555-0610, USA*

34

35 ** Corresponding author. Telephone: +33140613497. E-mail: louis.lambrechts@pasteur.fr*

36

37 **Summary**

38

39 Dengue viruses cause more human morbidity and mortality than any other arthropod-borne
40 virus. Dengue prevention relies primarily on vector control but the failure of traditional
41 methods has promoted the development of novel entomological approaches. Although use of
42 the intracellular bacterium *Wolbachia* to control mosquito populations was proposed half a
43 century ago, it has only gained significant interest as a potential agent of dengue control in
44 the last decade. Here, we review the evidence that supports a practical approach for dengue
45 reduction through field release of *Wolbachia*-infected mosquitoes and discuss the additional
46 studies that must be conducted before the strategy can be validated and operationally
47 implemented. A critical next step is to assess the efficacy of *Wolbachia* deployment in
48 reducing dengue virus transmission. We argue that a cluster-randomized trial is currently
49 premature because *Wolbachia* strain choice for release as well as deployment strategies are
50 still being optimized. We therefore present a pragmatic approach to acquiring preliminary
51 evidence of efficacy via a suite of complementary methodologies: prospective cohort study,
52 geographical cluster investigation, virus phylogenetic analysis, virus surveillance in
53 mosquitoes, and vector competence assays. This multi-pronged approach could provide
54 valuable intermediate evidence of efficacy to justify a future cluster-randomized trial.

55

56 Dengue is a major public health problem in tropical and sub-tropical regions, where almost
57 400 million infections are estimated to occur each year.¹ The etiological agents are four
58 dengue virus serotypes (DENV-1 to -4) in the genus *Flavivirus* that are transmitted among
59 humans by mosquitoes. These viruses cause a systemic, debilitating though mostly self-
60 limiting illness, which without careful management can lead to hypovolemic shock and
61 death.² In the absence of a licensed vaccine or therapeutic drug, dengue prevention efforts
62 are currently limited to the control of its main mosquito vector, *Aedes aegypti*. With a few
63 exceptions, the implementation of vector control methods has been largely unsuccessful due
64 to the lack of sustained commitment of resources³ and inability to effectively scale-up and
65 successfully apply interventions over large geographic areas and modern mega-cities. Novel
66 entomological approaches to dengue control have been developed⁴ and some are now
67 advancing to field testing.⁵

68

69

70 ***Wolbachia*-based strategies for dengue control**

71

72 One of the most promising entomological strategies being developed for dengue control
73 relies on introduction of the intracellular bacterium *Wolbachia* into *Ae. aegypti*.⁶ *Wolbachia*
74 *pipientis* is a bacterial endosymbiont that was originally identified in ovaries of *Culex*
75 mosquitoes in the 1920s⁷ and is thought to infect two-thirds of all living insect species.⁸ The
76 extraordinary evolutionary success of *Wolbachia* is attributed to their ability to manipulate the
77 biology of their hosts in diverse ways.⁹ For example, *Wolbachia* can induce reproductive
78 abnormalities such as feminization and sperm-egg cytoplasmic incompatibility (CI). Because
79 *Wolbachia* is transmitted vertically via the egg, these female-biased reproductive
80 manipulations can drive *Wolbachia* infections to high frequencies in wild populations. CI, the
81 most common manipulation in insects, occurs when *Wolbachia*-infected males mating with
82 *Wolbachia*-free females lead to the production of non-viable offspring. *Wolbachia*-infected

83 females, in contrast, produce successful offspring regardless of the *Wolbachia* infection
84 status of their mate.

85

86 The potential of *Wolbachia* to control pest insect populations was realized as early as half a
87 century ago (Figure 1). *Wolbachia*-induced CI was then proposed to eliminate *Culex*
88 mosquitoes¹⁰ or to introduce desirable genes into wild vector populations.¹¹ To date,
89 however, *Wolbachia* have never been operationally implemented as a vector control
90 measure. A significant hurdle was the fact that several major vectors of human pathogens
91 are not naturally infected by *Wolbachia*, including the main DENV vector *Ae. aegypti*. The
92 mosquito vectors (*Anopheles* spp.) of human malaria parasites were also thought to be
93 *Wolbachia*-free until a recent study reported evidence for *Wolbachia* in field populations of
94 *An. gambiae*.¹²

95

96 A resurgence of interest for *Wolbachia*-based strategies to control vector-borne diseases
97 occurred about a decade ago with the advent of transinfection techniques (Figure 1). Stable
98 *Wolbachia* infections in naïve hosts can now be established by embryonic microinjections
99 into the developing embryo germline. In general, *Wolbachia* transinfection is more likely to be
100 successful between closely related donor and recipient hosts, and the expression of
101 *Wolbachia*-induced phenotypes is conserved across hosts. In 2005, a stable infection by a
102 *Wolbachia* strain from the mosquito *Aedes albopictus* was established in *Ae. aegypti*, which
103 caused high rates of CI and rapidly spread to high frequencies in experimental populations.¹³
104 This was quickly followed by double transinfections of *Ae. aegypti* with two different
105 *Wolbachia* strains from *Ae. albopictus*.¹⁴

106

107 A second wave of breakthroughs occurred a few years later with the discovery of *Wolbachia*-
108 induced phenotypes in mosquitoes that had a direct effect on pathogen transmission (Figure
109 1). Until then *Wolbachia* was primarily considered a gene drive system, but the possibility to
110 transinfect *Wolbachia* strains from more distant hosts by cell culture adaptation prior to

111 microinjection,¹⁵ combined with the wide diversity of available *Wolbachia* strains and
112 properties, resulted in new mosquito-*Wolbachia* associations. Stable introduction of a life-
113 shortening strain of *Wolbachia* from *Drosophila* into *Ae. aegypti* halved the adult mosquito
114 life-span under laboratory conditions, making mosquitoes unlikely to live long enough to
115 transmit DENV.¹⁶ More importantly, this life-shortening *Wolbachia* strain directly inhibited the
116 ability of a range of pathogens, including DENV, to infect and replicate in *Ae. aegypti*.¹⁷
117 Finally, semi-field and field trials in Australia demonstrated that *Wolbachia* can be
118 persistently established in wild *Ae. aegypti* populations.^{18,19} Together, these properties form
119 the basis of a practical approach for suppression of DENV transmission through field release
120 of *Wolbachia*-infected mosquitoes.

121

122

123 **Current status of *Wolbachia* deployment for dengue control**

124

125 The critical next step is to assess the efficacy of medium-scale *Wolbachia* deployment in
126 reducing human DENV infection. The gold standard, a cluster-randomized trial (CRT) of
127 *Wolbachia*, has been considered in detail previously.²⁰ CRT is a type of randomized
128 controlled trial in which groups of subjects, instead of individual subjects, are randomly
129 assigned to the alternative treatments under study. The CRT design is particularly useful
130 when the intervention cannot be directed toward selected individual subjects, such as the
131 release of *Wolbachia*-infected mosquitoes. In the classical two-armed CRT, clusters without
132 intervention provide contemporaneous controls. In a stepped wedge CRT, the intervention is
133 rolled-out sequentially to all the clusters so that the clusters are their own controls over time.

134

135 We believe that at this time a CRT is premature for the *Wolbachia*-based approach for
136 several reasons. First, there are multiple strains of *Wolbachia* available for deployment, each
137 with its own characteristic effects on DENV blocking and mosquito fitness. A process of
138 selection through field-testing is still required before one or more final strain(s) can be chosen

139 for a particular release area. In addition, while deployment in North Queensland has provided
140 a basic template for release, this environment differs substantially from the large urban
141 centers in Southeast Asia and Latin America where a CRT would likely be carried out. It is
142 crucial to retain the capacity to learn during deployment about the effectiveness of release
143 strategies and community engagement and to adjust practice accordingly. Past examples of
144 adaptive changes made during deployment include releasing larger numbers of mosquitoes,
145 changing the intensity of trap grids to monitor *Wolbachia* spread, supplementing releases
146 with different mosquito developmental stages, and altering locations of deployment based on
147 community concerns.^{18,21} In contrast, the standard CRT approach would lock-in all aspects of
148 the release, preventing ‘on the fly’ improvements in design. Finally, a classical two-armed
149 CRT would have to be large, with >40 clusters that each include approximately 100 study
150 subjects who are monitored for infection to detect a 50% reduction in dengue with 90%
151 power.²⁰ Rough estimates of cost for such a design suggest it would exceed 5-10 million
152 USD.

153

154

155 **A pragmatic approach to optimize *Wolbachia* deployment**

156

157 Here, we argue that well-designed observational studies could provide a suite of valuable
158 indirect evidence that supports *Wolbachia* as a dengue intervention and, hence, justifies
159 continued development, ultimately leading to a definitive efficacy trial. Ideally, several
160 observational studies would be conducted in different settings and their outcomes combined
161 in a meta-analytic framework to assess the impact on disease and infection incidence. Below
162 we describe five possible approaches that could be used separately or in combination for
163 acquiring such evidence.

164

165 **Pediatric cohort study.** A prospective longitudinal cohort study that tracks seroconversion
166 rates in children could measure both the true incidence of DENV infections and the relative

167 risk of infection between *Wolbachia*-treated and untreated areas.²⁰ Because the overall
168 DENV seroconversion rate is generally 5-10% per annum in endemic countries,²² a cohort
169 would need to include at least several thousand individuals to be compatible with the
170 statistical requirements of a CRT with sufficient power to detect a moderate intervention
171 effect.²⁰ A smaller cohort of 1,000-1,500 children, although underpowered in the context of a
172 CRT, could be significantly enhanced by the concurrent approaches described below. Fine-
173 scale entomological surveillance (e.g., a grid of traps) would allow monitoring the spatio-
174 temporal dynamics of *Wolbachia* prevalence to distinguish, in real time, *Wolbachia*-free areas
175 from areas where *Wolbachia* had established. The raw entomological data could be
176 interpolated over time and space using standard methodology and serve as a covariate for
177 DENV seroconversion. As in other epidemiological investigations, participants residing in the
178 study area, but acquiring infections outside of the intervention area, represent a complication
179 to this approach.^{23,24} However, geographical cluster studies of dengue cases and fine-scale
180 spatiotemporal phylogenetic analyses of genomic DENV strain sequences (see below) would
181 help to address this concern.

182

183 **Geographical cluster investigation.** DENV infections are acute, often mild, inapparent or
184 with non-specific signs and symptoms, and thus are difficult to detect across populations in
185 real time. Active surveillance of human infections can be efficiently achieved using
186 geographical cluster sampling around dengue index cases.^{25,26} Here, 'index case' refers to
187 the laboratory-diagnosed clinical dengue case that initiates a cluster investigation within a
188 geographically restricted area around the home of a person with a documented DENV
189 infection. Geographical cluster investigations could be used to compare the fine-scale spatial
190 signature of DENV transmission in areas with and without *Wolbachia* (Figure 2). This
191 methodology would test the hypothesis that concurrent and/or subsequent infections around
192 an index case are reduced in areas where *Wolbachia*-infected mosquitoes are established.
193 Inward migration of dengue infections acquired outside the treatment area would also be a
194 confounding factor,^{23,24} although again potentially resolvable through detailed phylogenetic

195 analysis of virus sequences and/or monitoring movement patterns of study participants.
196 Nonetheless, if a *Wolbachia* intervention reduces local transmission at a micro-scale, it
197 should be detectable by a cluster investigation methodology. An efficacious intervention
198 would result in a lower overall number of index cases in the *Wolbachia*-treated areas and/or
199 reduction in concurrent infections measured by a lower frequency of cases that are
200 spatiotemporally linked to the index case.

201

202 **Virus sequence analysis.** Increasing access to viral genome sequence data has promoted
203 the development of new methodologies to infer dengue epidemiological dynamics based on
204 analyses of changing patterns in viral genetic diversity in time and space.^{27,28} Assuming that
205 multiple lineages of various DENV serotypes co-circulate prior to an intervention, a reduction
206 in local DENV transmission is expected to result in a decrease in viral genetic diversity
207 across serotypes in the intervention area due to a major viral demographic bottleneck, and in
208 an increase in the average dispersion distances travelled by DENV into the intervention area
209 (Figure 3). Phylogenetic analysis provides a simple means to identify importation of 'foreign'
210 viral lineages into the study area, provided that genetic diversity accumulates at a sufficiently
211 high rate. Previous studies on DENV microevolution in Southeast Asia suggested that spatial
212 patterns of genetic diversity are shaped by frequent virus immigration and highly focal
213 transmission.²⁸⁻³⁰ Although the level of phylogenetic resolution to be obtained is uncertain,
214 deep sequencing methods have recently undergone dramatic improvement, increasing the
215 power of this approach. We expect that if local DENV transmission is reduced in *Wolbachia*-
216 treated areas, some viruses will continue to be imported by human-mediated dispersal but
217 will not persist locally, reducing the strong spatial clustering that is typically observed in
218 DENV phylogenies.

219

220 **Virus detection in mosquitoes.** The release of *Wolbachia*-infected mosquitoes will require
221 monitoring of the local *Ae. aegypti* population for changes in *Wolbachia* frequency and
222 possibly in mosquito density. Recently, several sampling methods that effectively capture

223 female *Ae. aegypti* have been developed.³¹⁻³⁴ Virus detection could be combined with routine
224 molecular tests for *Wolbachia* presence. Detecting DENV-infected *Ae. aegypti* mosquitoes is
225 challenging because of the low infection rates (typically ~0.1%) in the adult females across
226 the population, although infection rates can be higher in locations of geographical cluster
227 investigations.²⁵ Because mosquitoes that test positive for virus are not necessarily
228 infectious, the proportion of DENV-infected mosquitoes does not directly translate into an
229 estimate of virus transmission unless virus disseminated from the mosquito midgut or in
230 saliva is also assayed, and even this approach is limited by the sensitivity of assays and
231 variation of *in vitro* saliva collections. Nonetheless, a successful intervention is expected to
232 reduce the incidence of viremic and infectious humans and, therefore, similarly reduce the
233 incidence of DENV infection in mosquitoes in areas where *Wolbachia* infection predominates.

234

235 **Vector competence assays.** Following the release of *Wolbachia*-infected mosquitoes, it will
236 be necessary to verify that the phenotype of reduced vector competence is maintained over
237 time in field-collected mosquitoes.³⁵ Vector competence assays consist of experimentally
238 exposing laboratory-reared mosquitoes to either an artificial infectious blood meal or the
239 blood of a viremic person.³⁶ The proportion of infectious mosquitoes (i.e., with virus detected
240 in saliva) is then measured over time. *Wolbachia*-infected mosquitoes have a strongly
241 reduced ability to deliver DENV in their saliva compared to *Wolbachia*-free mosquitoes.¹⁹
242 Ideally, vector competence experiments would be extended to human-to-mosquito-to-human
243 transmission experiments in a human challenge model.³⁷ Vector competence assays will
244 provide additional indirect evidence on the impact of the intervention, especially if the virus
245 interference effect is strong.

246

247

248 **Conclusions and perspectives**

249

250 The current challenge is to convert a promising strategy into a validated public health
251 intervention through rigorous assessment of its epidemiological impact. The suite of
252 approaches described above is not a substitute for a CRT. Nonetheless, this strategy has at
253 least two major strengths that can lay the foundations for a future CRT. First, the proposed
254 investigations are not dependent on the uniform application of the intervention, which by
255 nature will vary through time and space. Instead, an association between *Wolbachia*
256 presence and proxies of DENV transmission (e.g., DENV seroconversion or occurrence of
257 secondary cases around index cases) can be inferred dynamically from the spatiotemporal
258 correlation between these factors. Second, comprehensive observation and detection of
259 correlations between multiple environmental and biological factors will likely improve
260 fundamental understanding of dengue epidemiology that will inform and underpin future trial
261 designs. A multi-pronged approach would also help to evaluate potential impacts on other
262 *Ae. aegypti*-borne arboviruses (e.g., chikungunya virus), and the likelihood of unexpected
263 outcomes such as viral evolution to escape the inhibitory effects of *Wolbachia*, or other
264 unanticipated, adverse events.

265

266 Measuring the epidemiological impact of a *Wolbachia* deployment to reduce DENV
267 transmission is challenging. The intervention is not based on individuals, as a vaccine trial
268 would be, but on populations defined by spatial areas. The fundamental test of the impact of
269 the intervention is a comparison between areas where *Wolbachia*-infected mosquitoes are
270 present versus areas where they are not (Figures 2, 3). Although limited dispersal of *Ae.*
271 *aegypti*³⁸ and, therefore, spread of *Wolbachia*, is expected to maintain spatial delineation of
272 the intervention, a buffer zone will be necessary to avoid unanticipated overlap between
273 treatment and control areas. The intervention needs to be deployed over a large enough
274 geographic area to ensure that a sufficient number of dengue cases (or absence of cases if
275 the intervention is effective) is captured. Prior knowledge of the study area will help to assign
276 intervention and control areas with similar baseline transmission trends. Virus importation
277 into the intervention area (through human-mediated dispersal^{23,24}), which is likely to occur

278 and may reduce the signal-to-noise ratio, can be explored with geographic cluster studies
279 and by accounting for movement of study subjects.

280

281 One advantage of our proposed approach is that interpretation of seroconversion data from a
282 small-scale pediatric cohort can be enhanced by data from geographical cluster
283 investigations, viral sequencing and virus detection in mosquitoes, collectively resulting in a
284 body of evidence that could support continued development of *Wolbachia* as public health
285 tool. In any case, virus importation by study participants exposed to infected mosquitoes
286 outside of the treatment area would result in false positive cases in the *Wolbachia*-treated
287 area and conservatively lead to an underestimation of efficacy. A true placebo treatment (i.e.,
288 release of *Wolbachia*-free mosquitoes) is not ethically possible. The human and mosquito
289 samples can, however, be blinded prior to laboratory testing.

290

291 We have described a pragmatic approach for evaluation of novel entomological interventions
292 for dengue control through a coordinated, cross-disciplinary, ecological study that combines
293 several proxies of efficacy at the epidemiological, entomological and virological levels. It
294 relies on a combination of methodologies that have been successfully used to monitor
295 dengue epidemiological dynamics, as well as novel methodologies. Although this approach
296 has no precedent for dengue, it has the potential to provide valuable intermediate evidence
297 of efficacy that supports the *Wolbachia* methodology and justifies funding for a CRT or
298 deployment.

299

300 **Acknowledgements**

301

302 We thank Stephanie James and Fil M. Randazzo for their insights, and two anonymous
303 reviewers for helpful comments. This work was supported by a grant from the Foundation for
304 the National Institutes of Health through the Grand Challenges in Global Health Initiative of
305 the Bill and Melinda Gates Foundation.

306

307

308 **Contributors**

309

310 LL wrote the first draft of the manuscript. All other authors contributed equally to edit the
311 manuscript.

312

313

314 **Conflicts of interest**

315

316 We declare that we have no conflict of interest

317

318 **References**

- 319 1. Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, et al. The global
320 distribution and burden of dengue. *Nature* 2013; **496**: 504-7.
- 321 2. Simmons CP, Farrar JJ, Nguyen v V, Wills B. Dengue. *N Engl J Med* 2012; **366**:
322 1423-32.
- 323 3. Morrison AC, Zielinski-Gutierrez E, Scott TW, Rosenberg R. Defining challenges and
324 proposing solutions for control of the virus vector *Aedes aegypti*. *PLoS Med* 2008; **5**: e68.
- 325 4. McGraw EA, O'Neill SL. Beyond insecticides: new thinking on an ancient problem.
326 *Nat Rev Microbiol* 2013; **11**: 181-93.
- 327 5. James S, Simmons CP, James AA. Ecology. Mosquito trials. *Science* 2011; **334**: 771-
328 2.
- 329 6. Iturbe-Ormaetxe I, Walker T, O'Neill SL. *Wolbachia* and the biological control of
330 mosquito-borne disease. *EMBO Rep* 2011; **12**: 508-18.
- 331 7. Hertig M, Wolbach SB. Studies on *Rickettsia*-like micro-organisms in insects. *J Med*
332 *Res* 1924; **44**: 329-74.
- 333 8. Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH. How
334 many species are infected with *Wolbachia*?--A statistical analysis of current data. *FEMS*
335 *Microbiol Lett* 2008; **281**: 215-20.
- 336 9. Werren JH, Baldo L, Clark ME. *Wolbachia*: master manipulators of invertebrate
337 biology. *Nat Rev Microbiol* 2008; **6**: 741-51.
- 338 10. Laven H. Eradication of *Culex pipiens fatigans* through cytoplasmic incompatibility.
339 *Nature* 1967; **216**: 383-4.
- 340 11. Curtis CF. Population replacement in *Culex fatigans* by means of cytoplasmic
341 incompatibility. 2. Field cage experiments with overlapping generations. *Bull World Health*
342 *Organ* 1976; **53**: 107-19.
- 343 12. Baldini F, Segata N, Pompon J, Marcenac P, Robert Shaw W, Dabire RK, et al.
344 Evidence of natural *Wolbachia* infections in field populations of *Anopheles gambiae*. *Nature*
345 *Commun* 2014; **5**: 3985.

- 346 13. Xi Z, Khoo CC, Dobson SL. *Wolbachia* establishment and invasion in an *Aedes*
347 *aegypti* laboratory population. *Science* 2005; **310**: 326-8.
- 348 14. Ruang-Areerate T, Kittayapong P. *Wolbachia* transinfection in *Aedes aegypti*: a
349 potential gene driver of dengue vectors. *Proc Natl Acad Sci U S A* 2006; **103**: 12534-9.
- 350 15. McMeniman CJ, Lane AM, Fong AW, Voronin DA, Iturbe-Ormaetxe I, Yamada R, et
351 al. Host adaptation of a *Wolbachia* strain after long-term serial passage in mosquito cell lines.
352 *Appl Environ Microbiol* 2008; **74**: 6963-9.
- 353 16. McMeniman CJ, Lane RV, Cass BN, Fong AW, Sidhu M, Wang YF, et al. Stable
354 introduction of a life-shortening *Wolbachia* infection into the mosquito *Aedes aegypti*. *Science*
355 2009; **323**: 141-4.
- 356 17. Moreira LA, Iturbe-Ormaetxe I, Jeffery JA, Lu G, Pyke AT, Hedges LM, et al. A
357 *Wolbachia* symbiont in *Aedes aegypti* limits infection with dengue, Chikungunya, and
358 *Plasmodium*. *Cell* 2009; **139**: 1268-78.
- 359 18. Hoffmann AA, Montgomery BL, Popovici J, Iturbe-Ormaetxe I, Johnson PH, Muzzi F,
360 et al. Successful establishment of *Wolbachia* in *Aedes* populations to suppress dengue
361 transmission. *Nature* 2011; **476**: 454-7.
- 362 19. Walker T, Johnson PH, Moreira LA, Iturbe-Ormaetxe I, Frentiu FD, McMeniman CJ, et
363 al. The wMel *Wolbachia* strain blocks dengue and invades caged *Aedes aegypti* populations.
364 *Nature* 2011; **476**: 450-3.
- 365 20. Wolbers M, Kleinschmidt I, Simmons CP, Donnelly CA. Considerations in the design
366 of clinical trials to test novel entomological approaches to dengue control. *PLoS Negl Trop*
367 *Dis* 2012; **6**: e1937.
- 368 21. Harris AF, McKemey AR, Nimmo D, Curtis Z, Black I, Morgan SA, et al. Successful
369 suppression of a field mosquito population by sustained release of engineered male
370 mosquitoes. *Nature Biotechnol* 2012; **30**: 828-30.
- 371 22. Endy TP, Yoon IK, Mammen MP. Prospective cohort studies of dengue viral
372 transmission and severity of disease. *Curr Top Microbiol Immunol* 2010; **338**: 1-13.

- 373 23. Stoddard ST, Forshey BM, Morrison AC, Paz-Soldan VA, Vazquez-Prokopec GM,
374 Astete H, et al. House-to-house human movement drives dengue virus transmission. *Proc*
375 *Natl Acad Sci U S A* 2013; **110**: 994-9.
- 376 24. Stoddard ST, Morrison AC, Vazquez-Prokopec GM, Paz Soldan V, Kochel TJ, Kitron
377 U, et al. The role of human movement in the transmission of vector-borne pathogens. *PLoS*
378 *Negl Trop Dis* 2009; **3**: e481.
- 379 25. Yoon IK, Getis A, Aldstadt J, Rothman AL, Tannitisupawong D, Koenraadt CJ, et al.
380 Fine scale spatiotemporal clustering of dengue virus transmission in children and *Aedes*
381 *aegypti* in rural Thai villages. *PLoS Negl Trop Dis* 2012; **6**: e1730.
- 382 26. Reyes M, Mercado JC, Standish K, Matute JC, Ortega O, Moraga B, et al. Index
383 cluster study of dengue virus infection in Nicaragua. *Am J Trop Med Hyg* 2010; **83**: 683-9.
- 384 27. Rabaa MA, Ty Hang VT, Wills B, Farrar J, Simmons CP, Holmes EC. Phylogeography
385 of recently emerged DENV-2 in southern Viet Nam. *PLoS Negl Trop Dis* 2010; **4**: e766.
- 386 28. Raghwani J, Rambaut A, Holmes EC, Hang VT, Hien TT, Farrar J, et al. Endemic
387 dengue associated with the co-circulation of multiple viral lineages and localized density-
388 dependent transmission. *PLoS Pathog* 2011; **7**: e1002064.
- 389 29. Jarman RG, Holmes EC, Rodpradit P, Klungthong C, Gibbons RV, Nisalak A, et al.
390 Microevolution of dengue viruses circulating among primary school children in Kamphaeng
391 Phet, Thailand. *J Virol* 2008; **82**: 5494-500.
- 392 30. Rabaa MA, Klungthong C, Yoon IK, Holmes EC, Chinnawirotpisan P,
393 Thaisomboonsuk B, et al. Frequent in-migration and highly focal transmission of dengue
394 viruses among children in Kamphaeng Phet, Thailand. *PLoS Negl Trop Dis* 2013; **7**: e1990.
- 395 31. Barrera R, Mackay AJ, Amador M. An improved trap to capture adult container-
396 inhabiting mosquitoes. *J Am Mosq Control Assoc* 2013; **29**: 358-68.
- 397 32. Krockel U, Rose A, Eiras AE, Geier M. New tools for surveillance of adult yellow fever
398 mosquitoes: comparison of trap catches with human landing rates in an urban environment. *J*
399 *Am Mosq Control Assoc* 2006; **22**: 229-38.

- 400 33. Ritchie SA, Buhagiar TS, Townsend M, Hoffmann A, Van Den Hurk AF, McMahon JL,
401 et al. Field validation of the gravid Aedes trap (GAT) for collection of *Aedes aegypti* (Diptera:
402 Culicidae). *J Med Entomol* 2014; **51**: 210-9.
- 403 34. Vazquez-Prokopec GM, Galvin WA, Kelly R, Kitron U. A new, cost-effective, battery-
404 powered aspirator for adult mosquito collections. *J Med Entomol* 2009; **46**: 1256-9.
- 405 35. Frentiu FD, Zakir T, Walker T, Popovici J, Pyke AT, van den Hurk A, et al. Limited
406 dengue virus replication in field-collected *Aedes aegypti* mosquitoes infected with *Wolbachia*.
407 *PLoS Negl Trop Dis* 2014; **8**: e2688.
- 408 36. Nguyen N, Thi Hue Kien D, Tuan T, Quyen N, Tran C, Vo Thi L, et al. Host and viral
409 features of human dengue cases shape the population of infected and infectious *Aedes*
410 *aegypti* mosquitoes. *Proc Natl Acad Sci U S A* 2013; **110**: 9072-7.
- 411 37. Thomas SJ. Dengue human infection model: re-establishing a tool for understanding
412 dengue immunology and advancing vaccine development. *Hum Vaccin Immunother* 2013; **9**:
413 1587-90.
- 414 38. Harrington LC, Scott TW, Lerdthusnee K, Coleman RC, Costero A, Clark GG, et al.
415 Dispersal of the dengue vector *Aedes aegypti* within and between rural communities. *Am J*
416 *Trop Med Hyg* 2005; **72**: 209-20.
- 417
- 418

419 **Figure Legends**

420

421 **Figure 1. Key dates in the development of *Wolbachia*-based dengue control strategies.**

422 The timeline shows major achievements over the last century that have supported the
423 development of *Wolbachia* as a dengue control tool.

424

425

426 **Figure 2. Geographical cluster methodology.** The central dot represents the home of a
427 confirmed dengue case (orange: area with *Wolbachia*; green: area without *Wolbachia*).

428 People living within a 100-m radius (black dots) are screened for concomitant and/or
429 secondary DENV infection (crosses denote homes of additional DENV-infected individuals).

430

431

432 **Figure 3. Schematic representation of how *Wolbachia* intervention might change
433 patterns of virus genetic diversity.** Assuming that multiple lineages of various DENV

434 serotypes (colored dots) co-circulate prior to the intervention, a reduction in local DENV
435 transmission is expected to result in a decrease in viral genetic diversity in the intervention

436 area and a relative increase in the average dispersion distances.

437

438